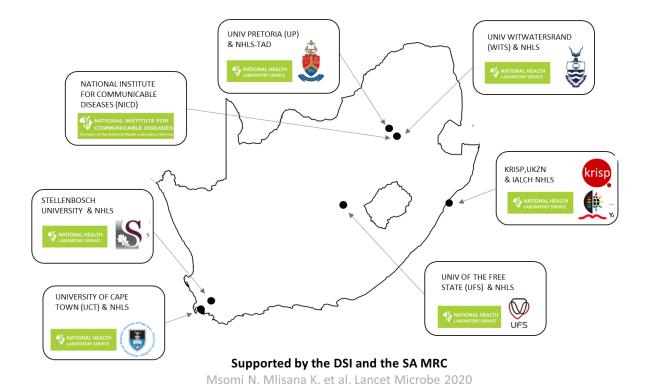


## **Network for Genomic Surveillance South Africa (NGS-SA)**

# SARS-CoV-2 Sequencing Update 29 October 2021

























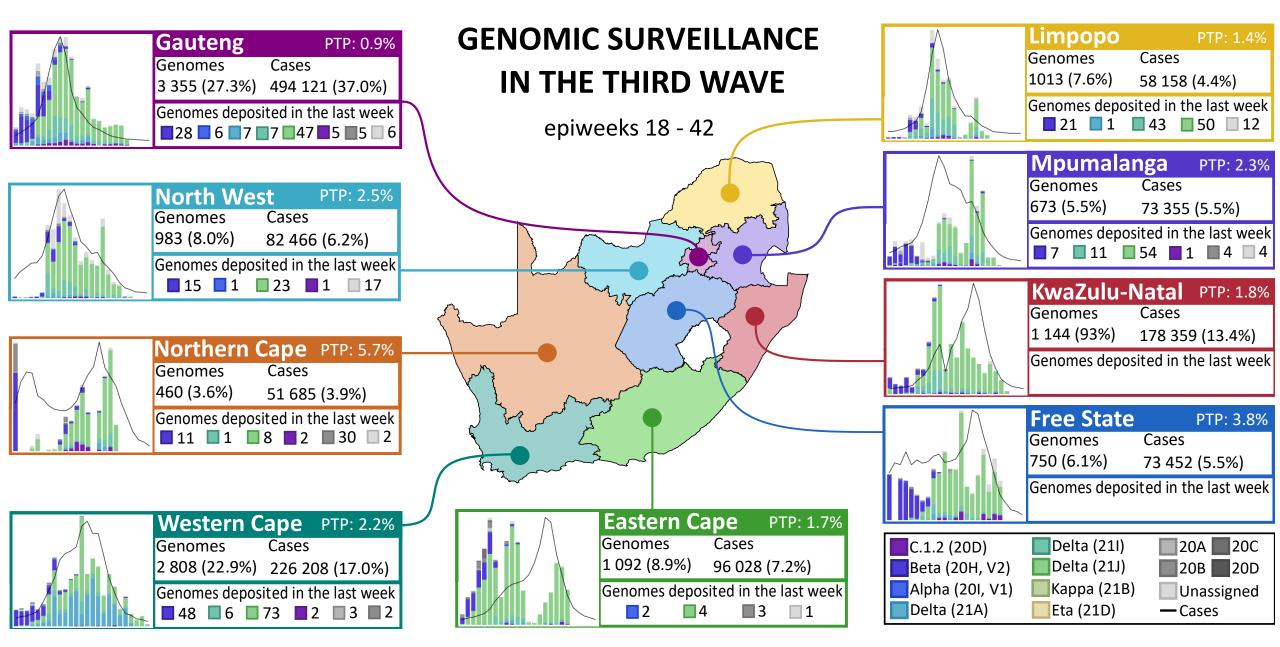
# The genomic data presented here are based on South African SARS-CoV-2 sequence data downloaded from GISAID (www.gisaid.org) on 29 October at 09h16



Data license: https://www.gisaid.org/registration/terms-of-use/

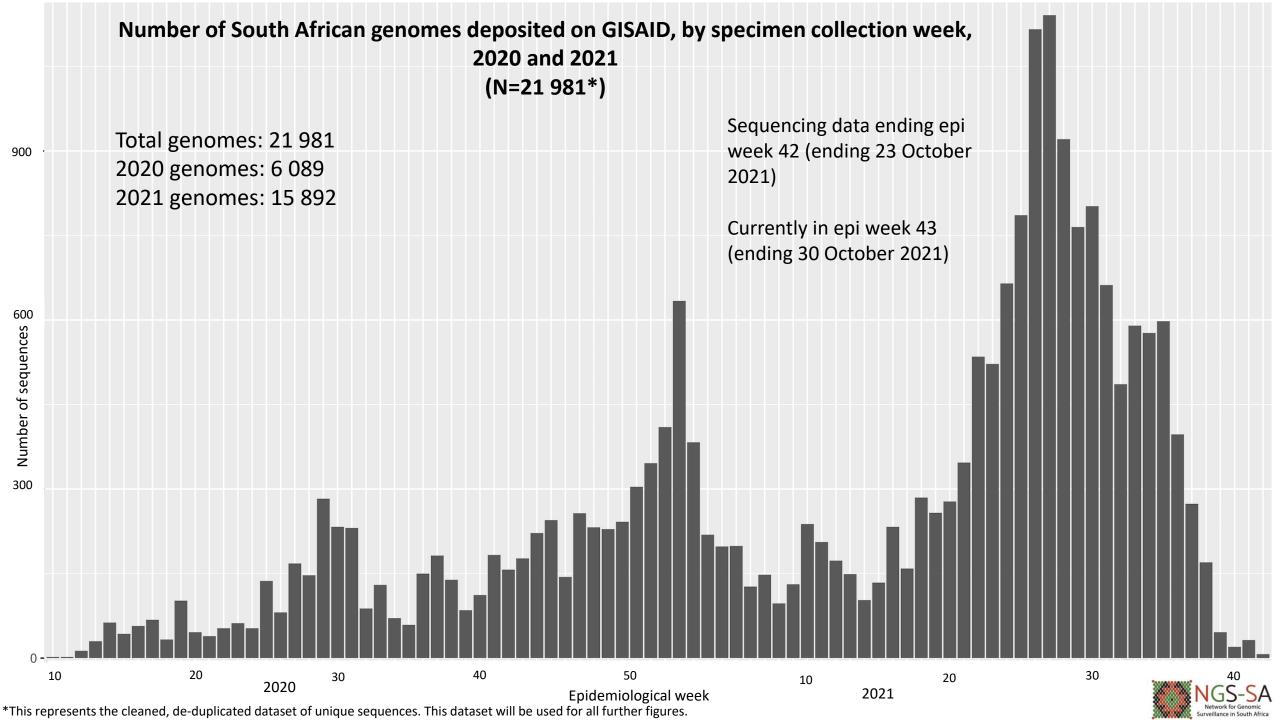
Elbe, S., and Buckland-Merrett, G. (2017) Data, disease and diplomacy: GISAID's innovative contribution to global health. Global Challenges, 1:33-46. DOI: 10.1002/gch2.1018 PMCID: 31565258

Shu, Y., McCauley, J. (2017) GISAID: Global initiative on sharing all influenza data – from vision to reality. EuroSurveillance, 22(13) DOI: 10.2807/1560-7917.ES.2017.22.13.30494 PMCID: PMC5388101

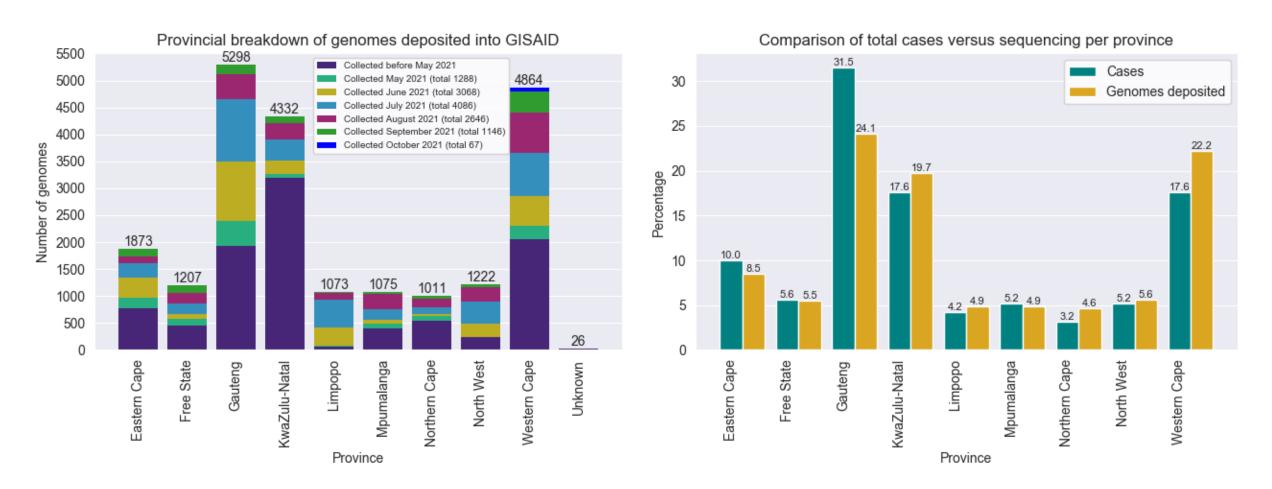


Bar graphs represent genomes sequenced per epiweek, with lines representing cases by collection date (weeks 18-42) Genomes and cases presented as provincial total (percentage of national total) for epiweeks 18-42 PTP: percentage testing positive





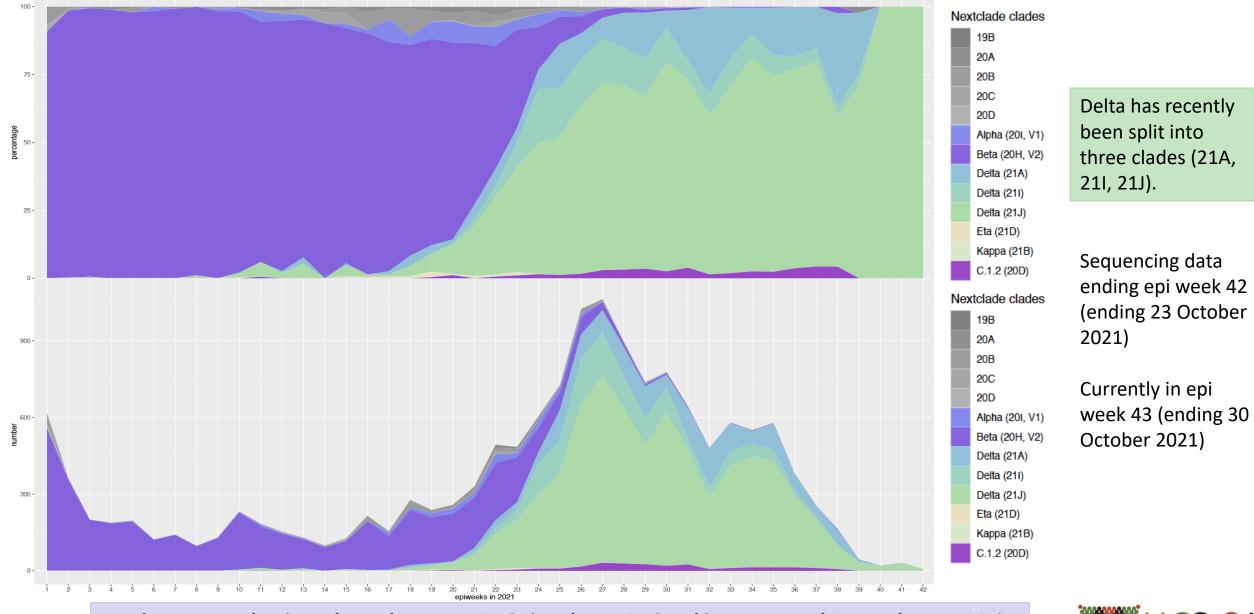
## GISAID genomes vs total cases, 2020 and 2021 (N=21 981)



All provinces, apart from GP, KZN, and WC, have comparable percentage of overall cases and overall sequenced genomes



## Distribution and number of clades in South Africa, 2021 (N= 15 892)

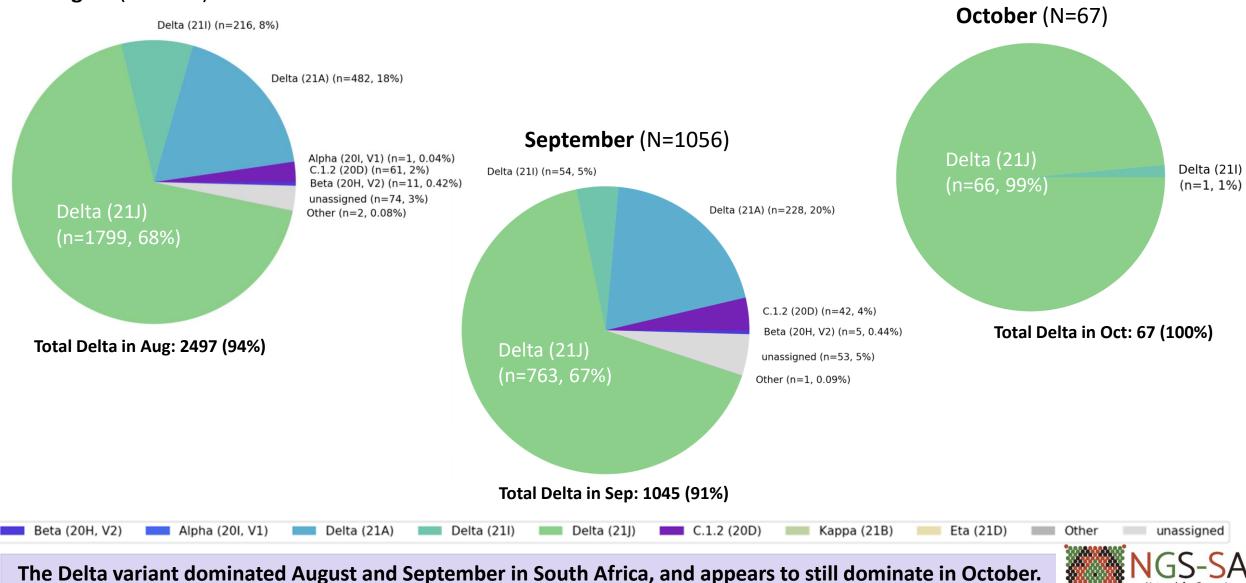


Delta came to dominate by end June at >65%, in July at >85% and in August and September at >90% C.1.2 present at <4% frequency since March

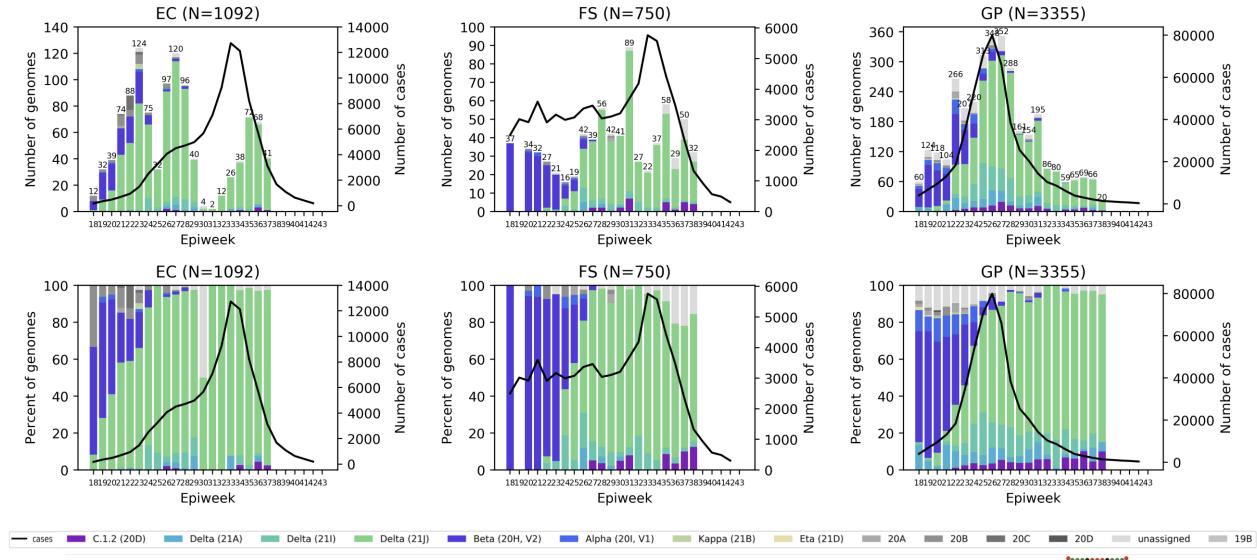


# Prevalence of Variants of Concern (VOC) and Variants of Interest (VOI) in August – October 2021 sequences, South Africa

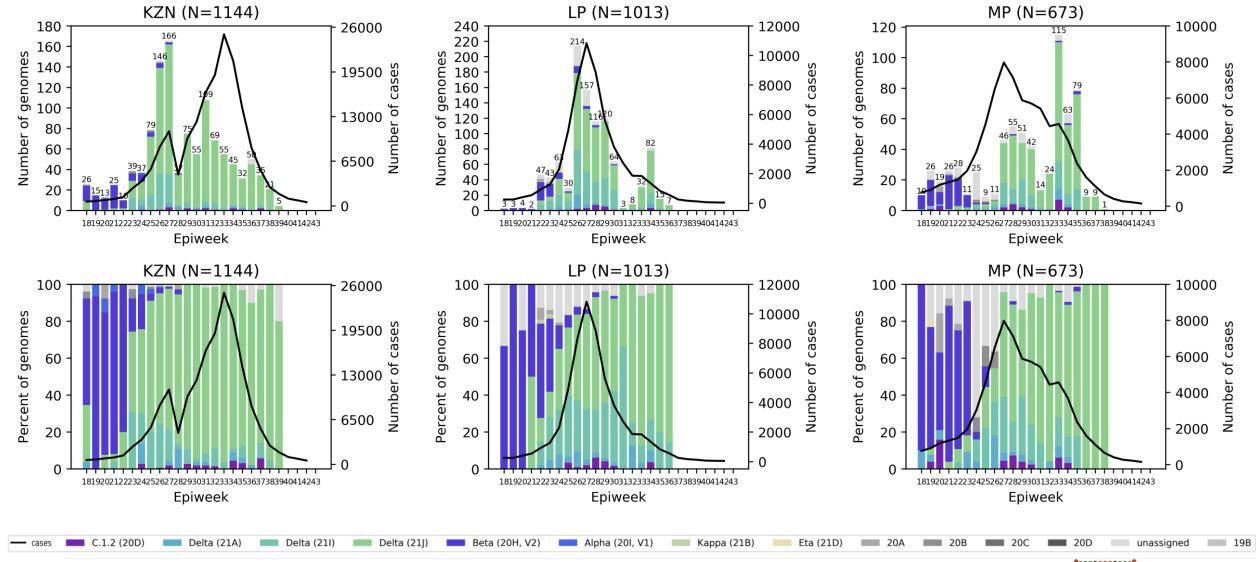
**August** (N=2646)



# Genomes sequenced from specimens collected in May to mid-September 2021 (epiweeks 18 – 43) from Eastern Cape, Free State and Gauteng Provinces

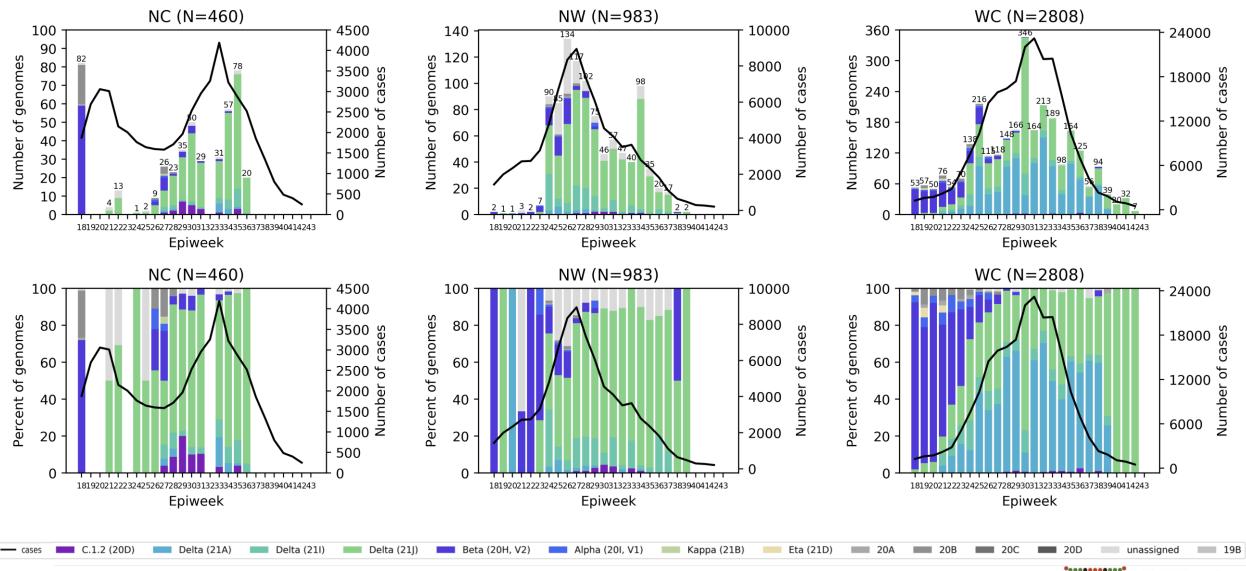


# Genomes sequenced from specimens collected in May to mid-September 2021 (epiweeks 18 – 43) from KwaZulu-Natal, Limpopo and Mpumalanga Provinces

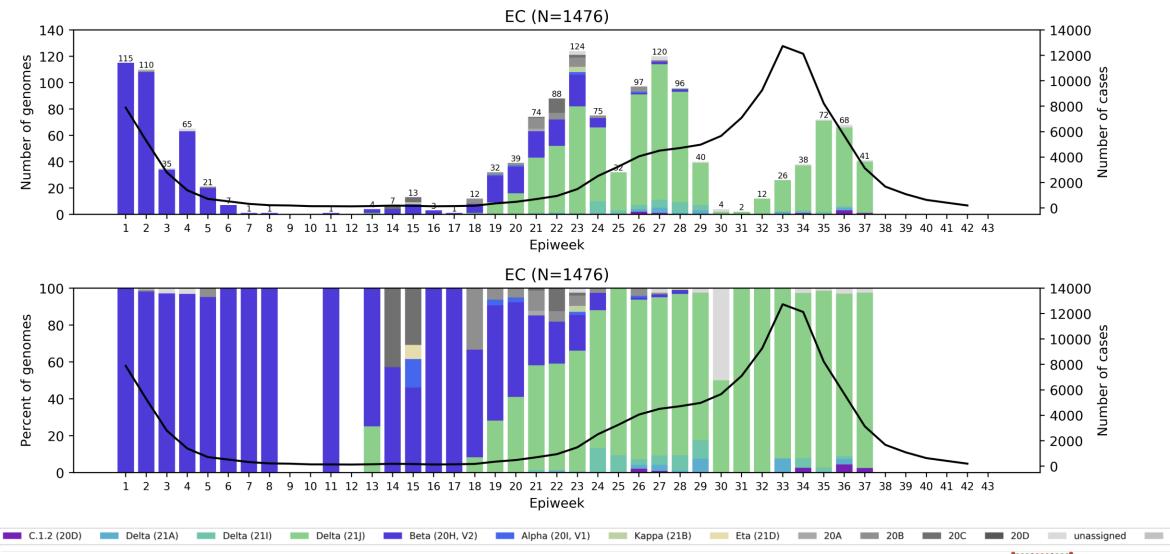




# Genomes sequenced from specimens collected in May to mid-September 2021 (epiweeks 18 – 43) from Northern Cape, North West, and Western Cape Provinces

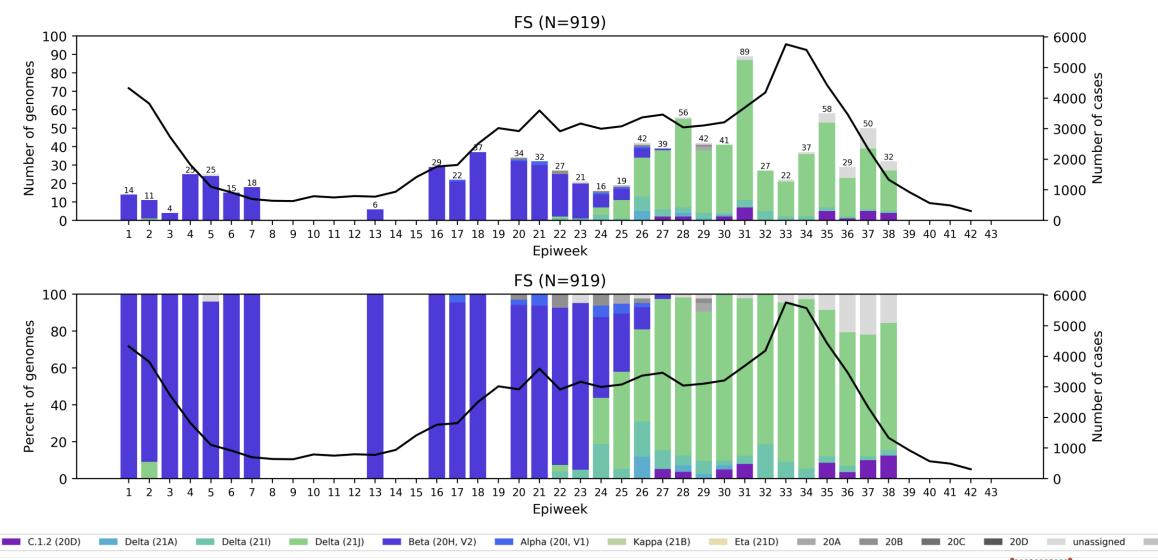


# Eastern Cape Province, 2021, n = 1476



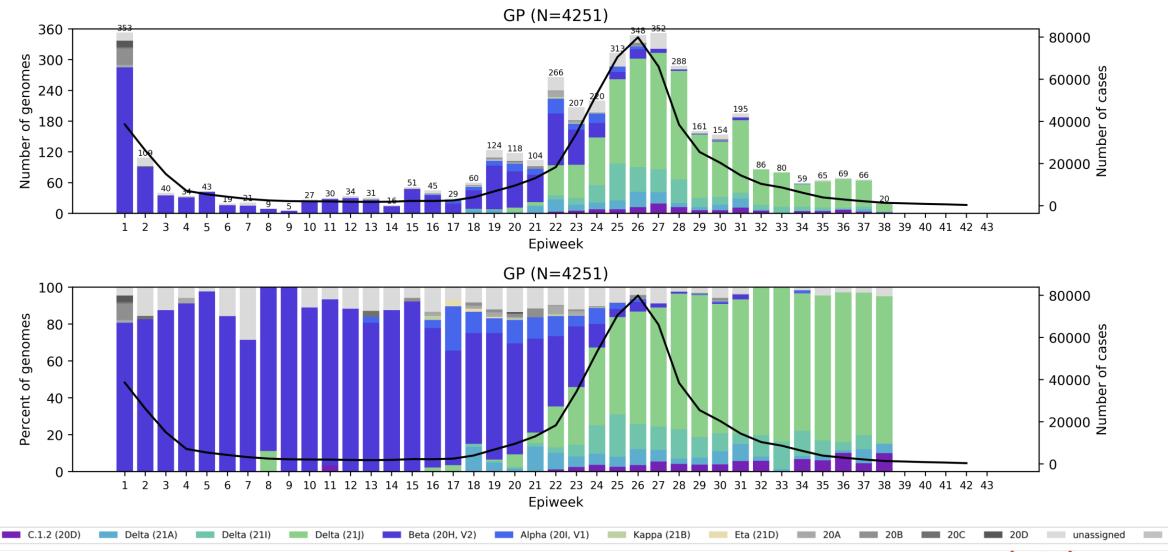


## Free State Province, 2021, n = 919



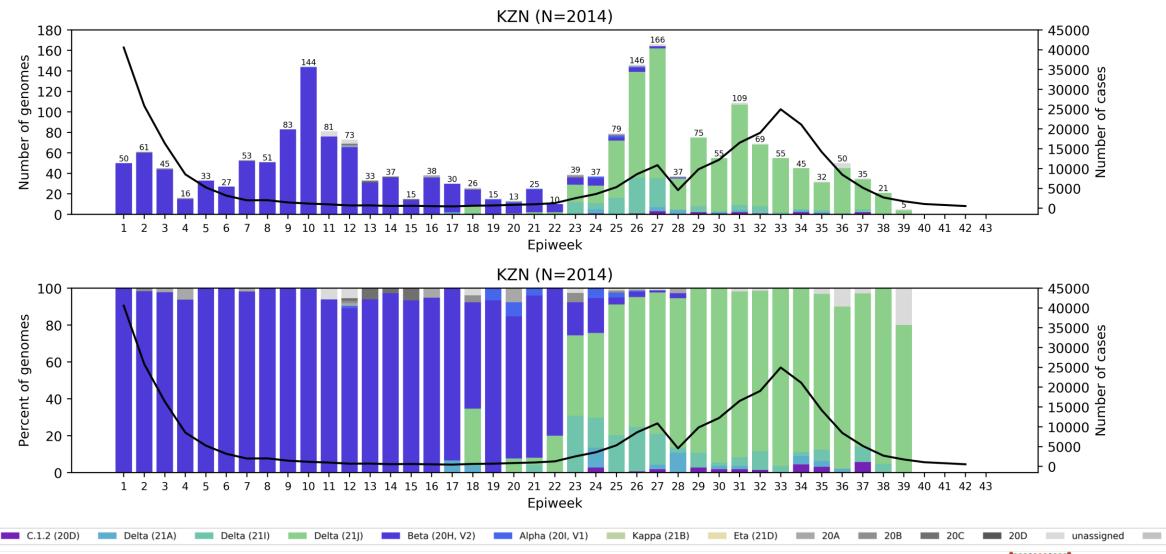


# **Gauteng Province, 2021, n = 4251**



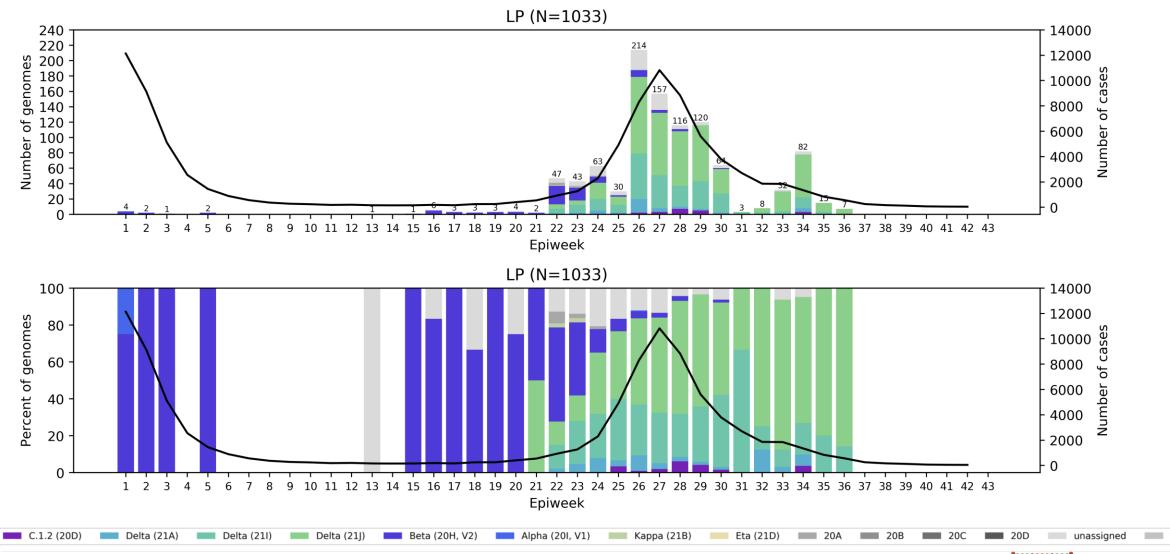


# KwaZulu-Natal Province, 2021, n = 2014



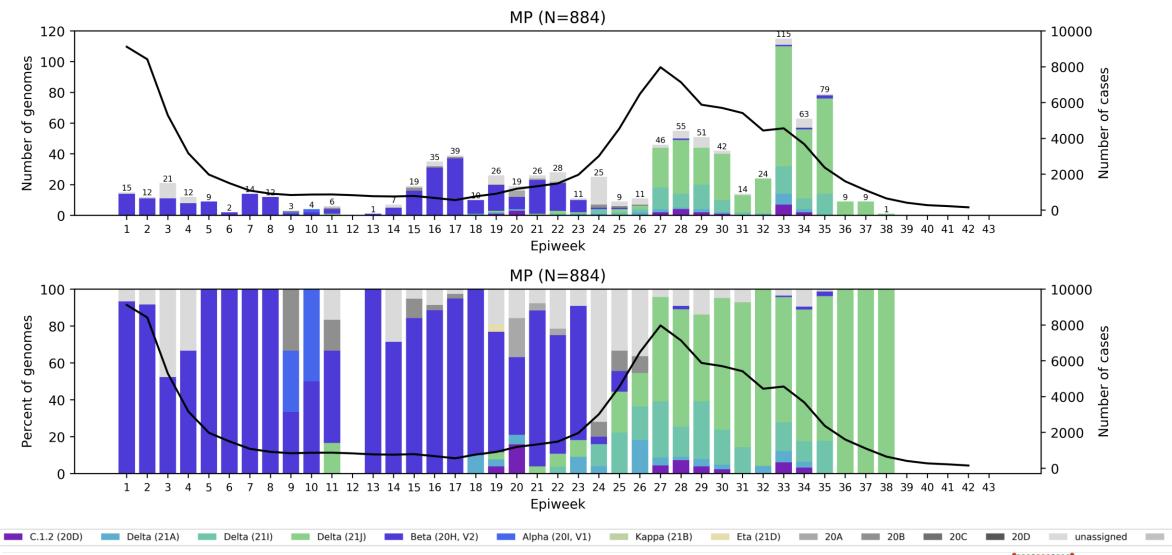


# **Limpopo Province, 2021, n = 1033**



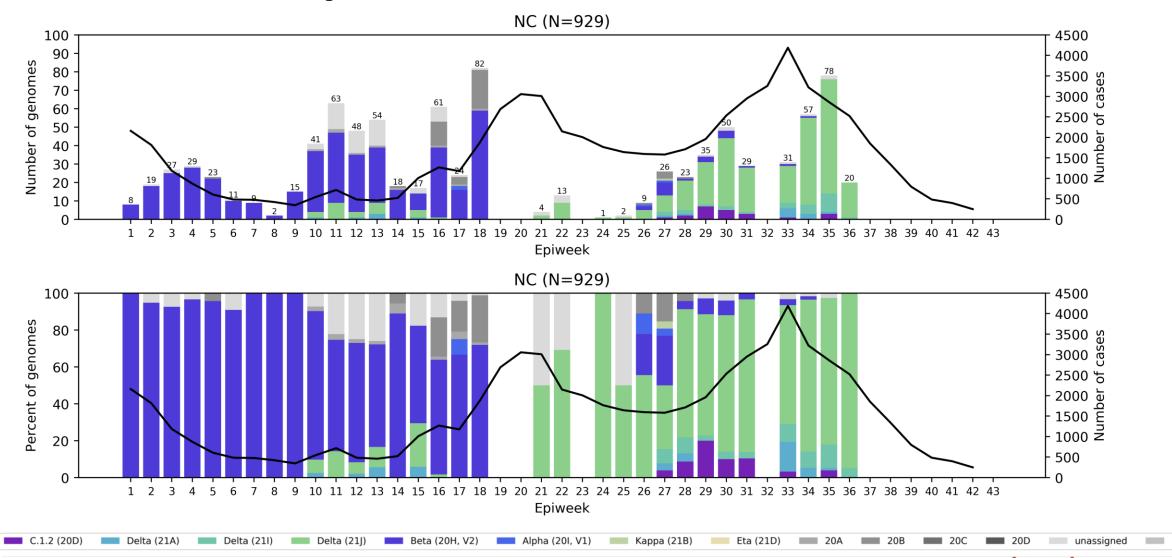


# Mpumalanga Province, 2021, n = 884



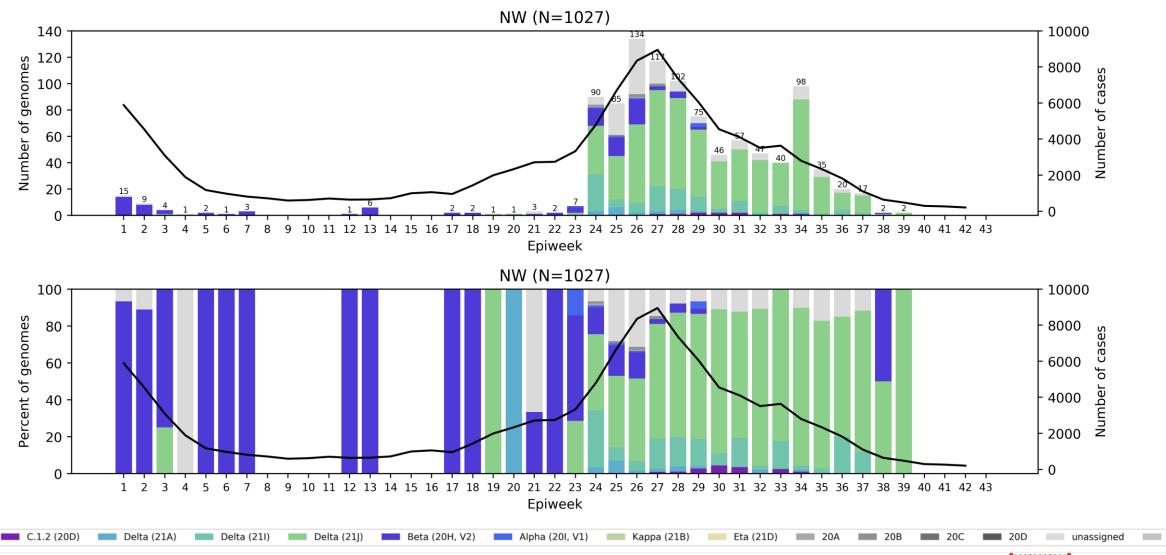


# Northern Cape Province, 2021, n = 929



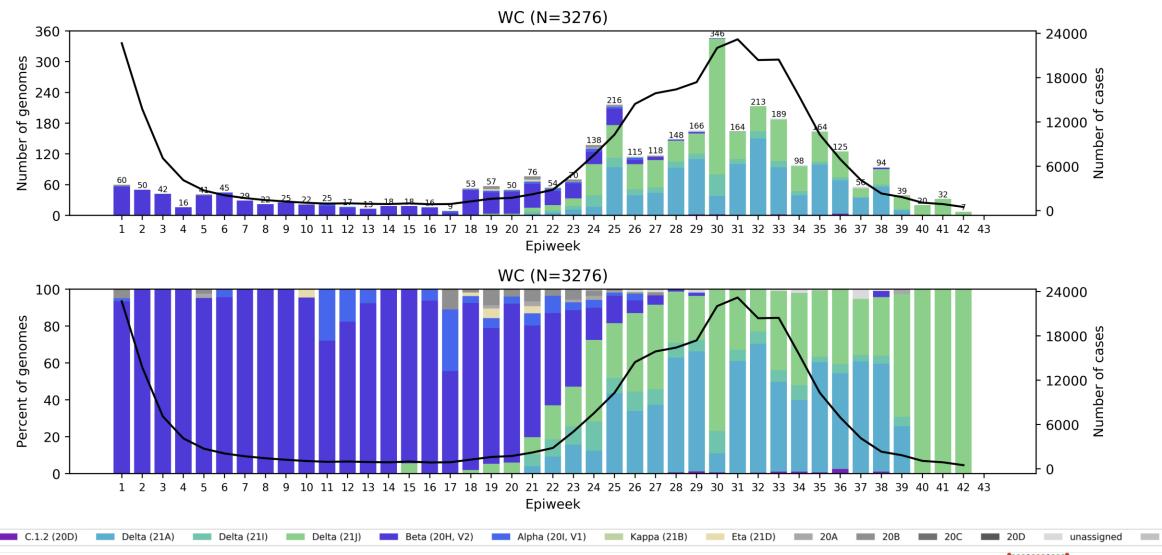


# North West Province, 2021, n = 970





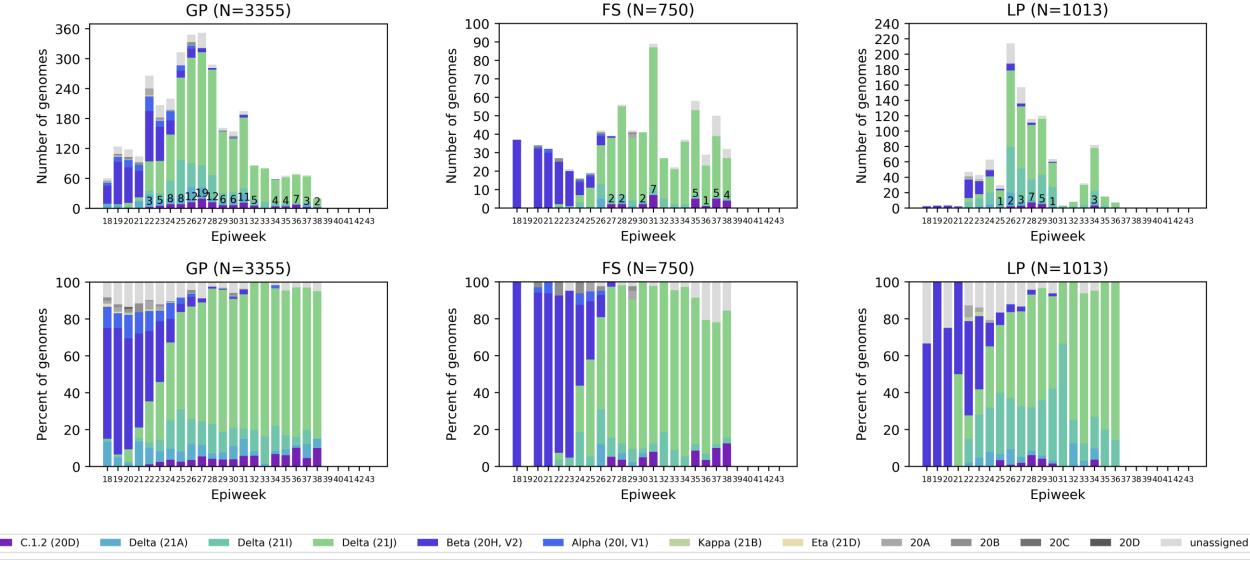
# Western Cape Province, 2021, n = 3276





## C.1.2 (n=257 in SA) in May – August 2021 by epiweek

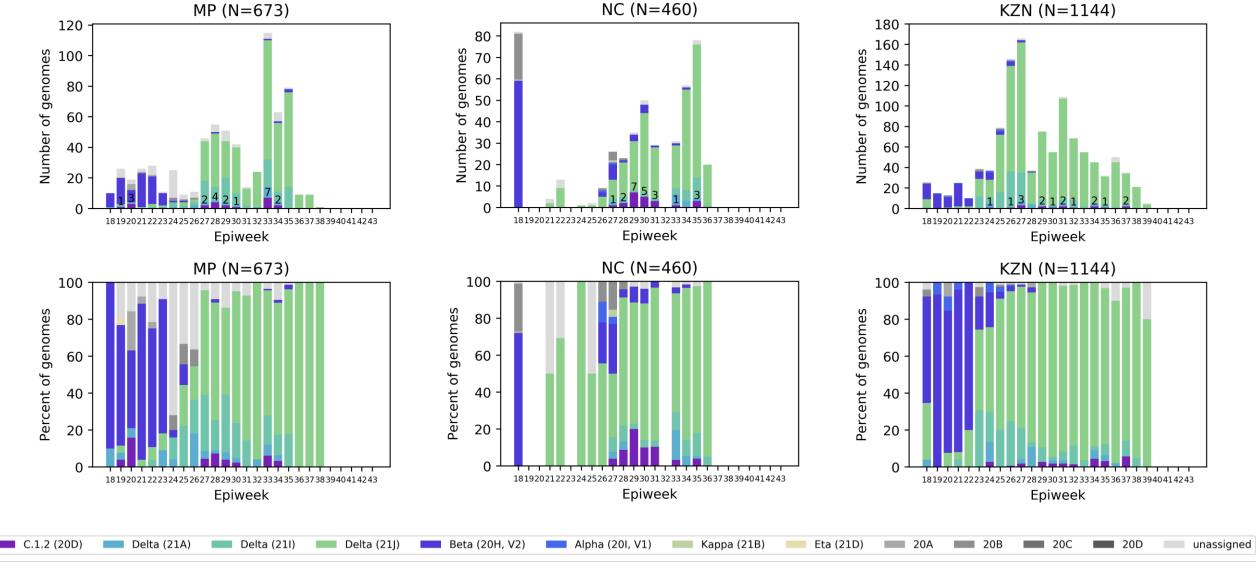
Number of C.1.2 samples indicated above bar, provinces ordered by number of detections



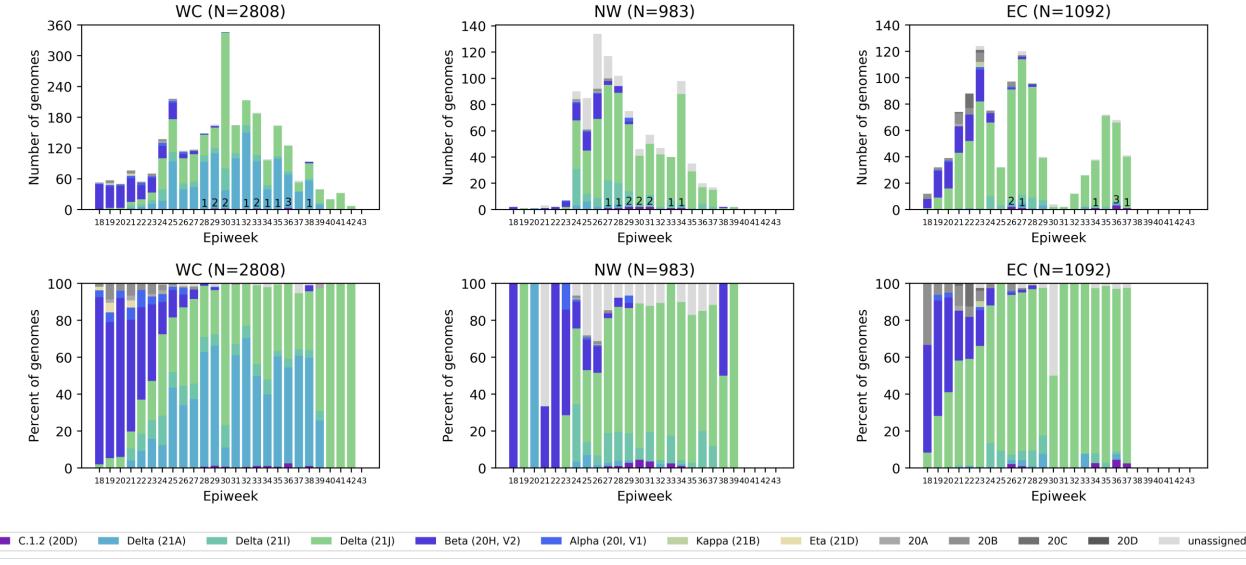
The majority of C.1.2 sequences have been detected in Gauteng (n=115), followed by the Free State (n=28) and then Limpopo (n=22).



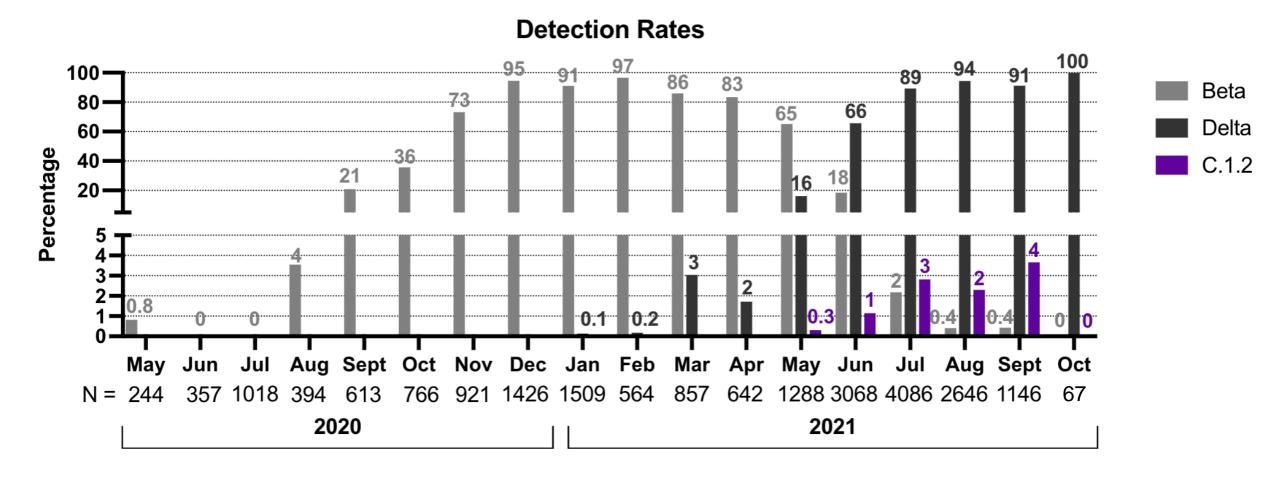
Number of C.1.2 samples indicated above bar, provinces ordered by number of detections



## C.1.2 (n=257 in SA) in May – August 2021 by epiweek



## C.1.2 growth compared to Beta and Delta



C.1.2 continues to be detected at low levels (less than 4% of genomes per month)



# Summary

- Delta continues to dominate in all provinces from specimens collected in September and October
  - Delta has recently been split into three clades.
    - New clades have been assigned due to >20% global circulation of particular sequences for more than 2 months
    - All SA Delta samples have been updated
    - Delta 21J is the dominant clade globally and in SA
    - Based on updated definitions of AY.4.2<sup>1,2</sup>, this sub-lineage has not been detected in the country
- Mutated C.1.2 lineage detected in all provinces of South Africa at less than 4% of genomes
- Lambda and Mu variants not detected in South Africa

























UNIVERSITY OF KWAZULU-NATAL

INYUVESI
YAKWAZULU-NATALI



## UKZN-Inkosi Albert Luthuli Central Hospital





Dr Khanyi Msomi

Dr Kerusha Govender

Dr Pravi Moodley

Dr Aabida Khan

Dr Lili Gounder

Dr Kerri Francois

Dr Cherise Naicker

Dr Joedene Chetty

Dr Neli Ngcaba

Dr Tshepiso Mosito

Mr Malcolm Ellapen

Mr Kubendran Reddy

The COVID-19 Bench team

## University of KwaZulu-Natal & Africa Health Research Institute



#### **KRISP at UKZN:**

Tulio de Oliveira Richard Lessels Houriiyah Tegally Eduan Wilkinson Jennifer Giandhari Sureshnee Pillay Emmanuel James San



#### **AHRI**

Alex Sigal Sandile Cele Willem Hanekom

# University of Stellenbosch & NHLS Tygerberg Virology





Susan Engelbrecht

**Wolfgang Preiser** 

Gert van Zyl

Tongai Maponga

Bronwyn Kleinhans

**Shannon Wilson** 

Karabo Phadu

Tania Stander

Kamela Mahlakwane

Mathilda Claassen

Diagnostic laboratory staff



# University of Cape Town, NHLS & WCG





#### **NHLS-UCT**

Carolyn Williamson Nei-yuan Hsiao Diana Hardie Kruger Marais Stephen Korsman Ziyaad Valley-Omar

#### **WCG-UCT**

Mary-Anne Davies Hannah Hussey Andrew Boulle Masudah Paleker Theuns Jacobs Erna Morden









**CAPE TOWN HVTN** 

## **UCT, IDM and CIDRI-Africa**

Deelan Doolabh
Arash Iranzadeh
Lynn Tyers
Innocent Mudau
Nokuzola Mbhele
Fezokuhle Khumalo
Thabang Serakge
Bruna Galvão
Arghavan Alisoltani

(U. California)

Robert Wilkinson

Darren Martin

Nicola Mulder

Wendy Burgers Ntobeko Ntusi

Rageema Joseph

Sean Wasserman

Linda Boloko



# University of the Free State



#### UFS

Dominique Goedhals
Armand Bester
Martin Myaga
Peter Mwangi
Emmanuel Ogunbayo
Milton Mogotsi
Makgotso Maotoana
Lutfiyya Mohamed



## **NHLS Division of Virology**

Sabeehah Vawda Felicity Burt Thokozani Mkhize Diagnostic laboratory staff



## **National Institute for Communicable Diseases**



## **Centre for Respiratory Diseases & Meningitis**

Jinal Bhiman

Anne von Gottberg

Thabo Mohale

Daniel Amoako

Josie Everatt

Boitshoko Mahlangu

Noxolo Ntuli

Anele Mnguni

Amelia Buys

Cardia Fourie

Noluthando Duma

Linda de Gouveia

Jackie Kleynhans

**Nicole Wolter** 

Sibongile Walaza

Mignon du Plessis

Stefano Tempia

Mvuyo Makhasi

Cheryl Cohen

health

REPUBLIC OF SOUTH AFRICA

## **Centre for HIV and STIs**

**Cathrine Scheepers** 

**Constantinos Kurt Wibmer** 

Thandeka Moyo

**Tandile Hermanus** 

Frances Ayres

Zanele Molaudzi

Bronwen Lambson

**Tandile Hermanus** 

Mashudu Madzivhandila

Prudence Kgagudi

**Brent Oosthuysen** 

**Penny Moore** 

Lynn Morris

## **NICD Groups**

NICD COVID-19 response team

NICD SARS-CoV-2 Sequencing

Group

## **Sequencing Core Facility**

Zamantungwa Khumalo

Annie Chan

Morne du Plessis

Stanford Kwenda

Phillip Senzo Mtshali

Mushal Allam

Florah Mnyameni

**Arshad Ismail** 

















# Zoonotic arbo and respiratory virus program Centre for Viral Zoonoses Department Medical Virology/ NHLS Tshwane Academic division University of Pretoria



## ZARV research program/UP

Marietjie Venter (Head: ZARV)

Adriano Mendes (Postdoc)

Amy Strydom (Postdoc)

Michaela Davis (MSc, intern medical scientist)



#### **NHLS Tshwane**

Prof Simnikiwe Mayaphi (HOD)

#### **Funders**:

GIZ/BMBF: African Network for Improved diagnostics and epidemiology of common and emerging infectious agents (ANDEMIA)
G7 Global Health fund, Robert Koch Institute, Dr Fabian Leendertz

















## Additional support and collaborators

**CAPRISA** 

Nigel Garret

**UKZN - Big Data** 

Ilya Sinayskiy

José Lourenço

Francesco Pettruccione

**University of Oxford** 

Salim Abdool Karim

**NHLS** Koeleka Mlisana Zinhle Makatini **Eugene Elliot** Florette K. Treurnicht Kathleen Subramoney

Oluwakemi Laguda-Akingba **Shareef Abrahams** Greta Hoyland Gloria Selabe Elias Bereda

**Hyrax Biosciences Simon Travers** 

Jeannette Wadula

**Cape Town HVTN Laboratory** Erica Anderson-Nissen Anneta Naidoo

Ndlovu Research **Hugo Tempelman** CJ Umunnakwe

Lancet Allison J. Glass

**Ampath** Terry Marshall Cindy van Deventer **Eddie Silberbauer** 

**Pathcare Vermaak Andries Dreyer Howard Newman** Riaan Writes Marianne Wolfaardt Warren Lowman

FioCruz, Brazil Vagner Fonseca Marta Giovanetti Luiz Carlos Junior Alcantara

**Bridge-the-Gap** Raymond Rott

**Cytespace Africa Laboratories** Christa Viljoen

**ARC-OVI** Lia Rotherham **Africa CDC** 

John Nkengasong Sofonias Tessema

**Netcare:** 

Richard Friedland Craig Murphy Caroline Maslo Liza Sitharam

DSI **Glaudina Loots** 

**SA MRC** Glenda Gray





NET*C*ARE





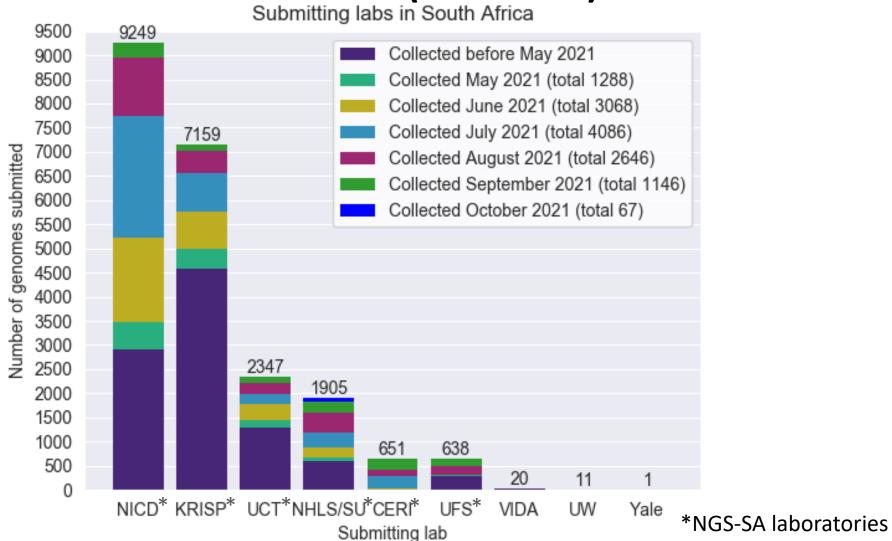








# South African genomes submitted per sequencing lab, 2020 and 2021 (N=21 981)





# Variants of Concern (VOC)

WHO label	Pango lineages+	GISAID clade	Nextstrain clade	Additional amino acid changes monitored*	Earliest documented samples	Date of designation
Alpha	B.1.1.7 <sup>#</sup>	GRY	20I (V1)	+S:484K +S:452R	United Kingdom, Sep-2020	18-Dec-2020
Beta	B.1.351	GH/501Y.V2	20H (V2)	+S:L18F	South Africa, May-2020	18-Dec-2020
Gamma	P.1	GR/501Y.V3	20J (V3)	+S:681H	Brazil, Nov-2020	11-Jan-2021
Delta	B.1.617.2 <sup>§</sup>	G/478K.V1	21A	+S:417N	India, Oct-2020	VOI: 4-Apr-2021 VOC: 11-May-2021

https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/ accessed 22 October 2021

<sup>\*</sup>Notable spike (S) amino acid changes under monitoring, which are currently reported in a minority of sequenced samples <sup>†</sup>Includes all descendant lineages.

<sup>#</sup>Includes all Q.\* lineages in the PANGO nomenclature system.

<sup>§</sup>Includes all AY.\* lineages in the PANGO nomenclature system.

# **Currently designated Variants of Interest (VOI)**

WHO label	Pango* lineages	GISAID clade	Nextstrain clade	Earliest documented samples	Date of designation
Lambda	C.37	GR/452Q.V1	21G	Peru, Dec-2020	14-Jun-2021
Mu	B.1.631	GH	21H	Colombia, Jan-2021	30-Aug-2021

https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/ accessed 22 October 2021

<sup>\*</sup>Includes all descendant lineages.

# Submission of routine specimens for sequencing

- representative of multiple geographic regions (provinces/districts/health facilities) from individuals of
  - all ages
  - over as many time periods during the SARS-CoV-2 epidemic in South Africa
- requested that testing laboratories in both the private and public sectors, submit respiratory samples to their closest NGS-SA sequencing laboratory on a routine basis (ideally every week) as follows, depending on the capacity of the testing laboratory:
  - All positives samples should be sent every week (NGS-SA laboratory will perform random sampling as described below) OR
  - A weekly selection of approximately 10%-20% of randomly selected positive samples should be sent every week. Number of selected samples will depend on the size of laboratory and how many other laboratories are drained by the submitting laboratory.

# Submission of special interest specimens for sequencing

In addition to routine samples mentioned above, please send specimens separately to above and clearly marked if:

- Suspected vaccine breakthrough (≥14 days after vaccine), especially if hospitalised and clinically severe
- Suspected re-infection (≥90 days after previous episode), especially if hospitalised and clinically severe
- Prolonged shedding with high SARS-CoV-2 viral loads (i.e. Ct values less than 30 for more than 1 month post-primary diagnosis) in immunocompromised individuals
- Possible animal-to-human transmission
- Suspected cases of importation from another country, especially countries known to harbour SARS-CoV-2 variants of concern or countries with little available information
- Clusters of "unusual" cases (e.g., in terms of disease presentation, patient groups affected, etc.)