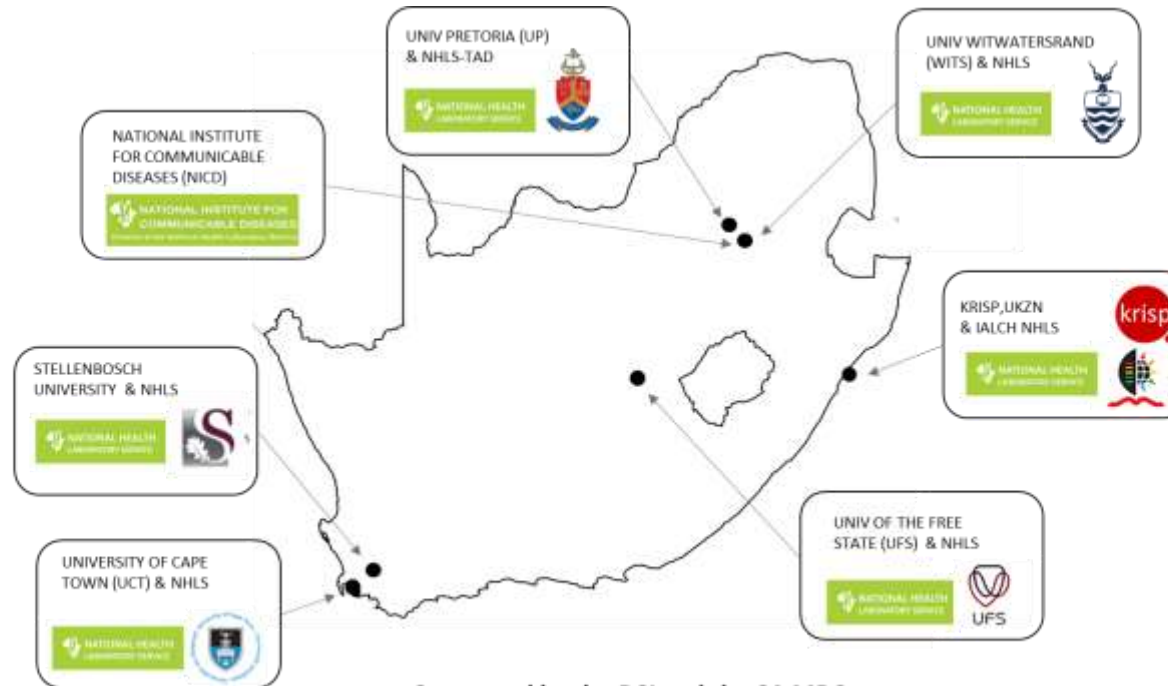


## SARS-CoV-2 Sequencing Update 5 November 2021



Supported by the DSI and the SA MRC

Msomi N. Mlisana K. et al. Lancet Microbe 2020

The genomic data presented here are based on South African SARS-CoV-2 sequence data downloaded from GISAID ([www.gisaid.org](http://www.gisaid.org)) on 4 November at 21h36



Data license: <https://www.gisaid.org/registration/terms-of-use/>

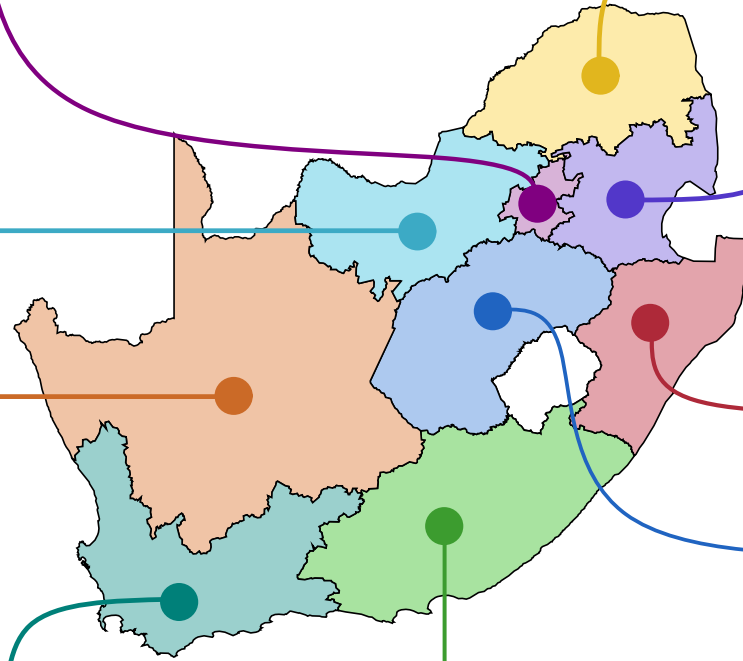
Elbe, S., and Buckland-Merrett, G. (2017) Data, disease and diplomacy: GISAID's innovative contribution to global health. *Global Challenges*, 1:33-46. DOI: 10.1002/gch2.1018 PMCID: 31565258

Shu, Y., McCauley, J. (2017) GISAID: Global initiative on sharing all influenza data – from vision to reality. *EuroSurveillance*, 22(13) DOI: 10.2807/1560-7917.ES.2017.22.13.30494 PMCID: PMC5388101

Case data is based on specimen collection date. Cases from <https://www.nicd.ac.za/diseases-a-z-index/disease-index-covid-19/surveillance-reports/weekly-epidemiological-brief/>

# GENOMIC SURVEILLANCE IN THE THIRD WAVE

epiweeks 18 - 43



## Gauteng

PTP: 0.9%

Genomes Cases  
3 488 (27.6%) 494 744 (37.0%)

Genomes deposited in the last week

■ 1 ■ 3 ■ 1 ■ 6 ■ 103 ■ 5 ■ 14

## Limpopo

PTP: 1.0%

Genomes Cases  
1 036 (8.2%) 58 231 (4.4%)

Genomes deposited in the last week

■ 5 ■ 16 ■ 2

## North West

PTP: 1.5%

Genomes Cases  
1 025 (8.1%) 82 606 (6.2%)

Genomes deposited in the last week

■ 1 ■ 4 ■ 27 ■ 10

## Mpumalanga

PTP: 1.6%

Genomes Cases  
701 (5.5%) 73 540 (5.5%)

Genomes deposited in the last week

■ 1 ■ 2 ■ 23 ■ 1 ■ 1

## Northern Cape

PTP: 5.2%

Genomes Cases  
538 (4.3%) 51 911 (3.9%)

Genomes deposited in the last week

■ 3 ■ 72 ■ 1 ■ 2

## KwaZulu-Natal

PTP: 1.3%

Genomes Cases  
1 145 (9.1%) 178 752 (13.4%)

Genomes deposited in the last week

■ 1

## Western Cape

PTP: 1.7%

Genomes Cases  
2 816 (22.3%) 226 593 (17.0%)

Genomes deposited in the last week

■ 1 ■ 6 ■ 1

## Eastern Cape

PTP: 1.5%

Genomes Cases  
1 140 (9.0%) 96 204 (7.2%)

Genomes deposited in the last week

■ 42 ■ 1 ■ 5

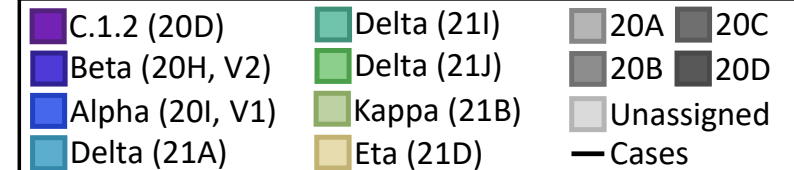
## Free State

PTP: 2.9%

Genomes Cases  
755 (6.0%) 73 718 (5.5%)

Genomes deposited in the last week

■ 4 ■ 1

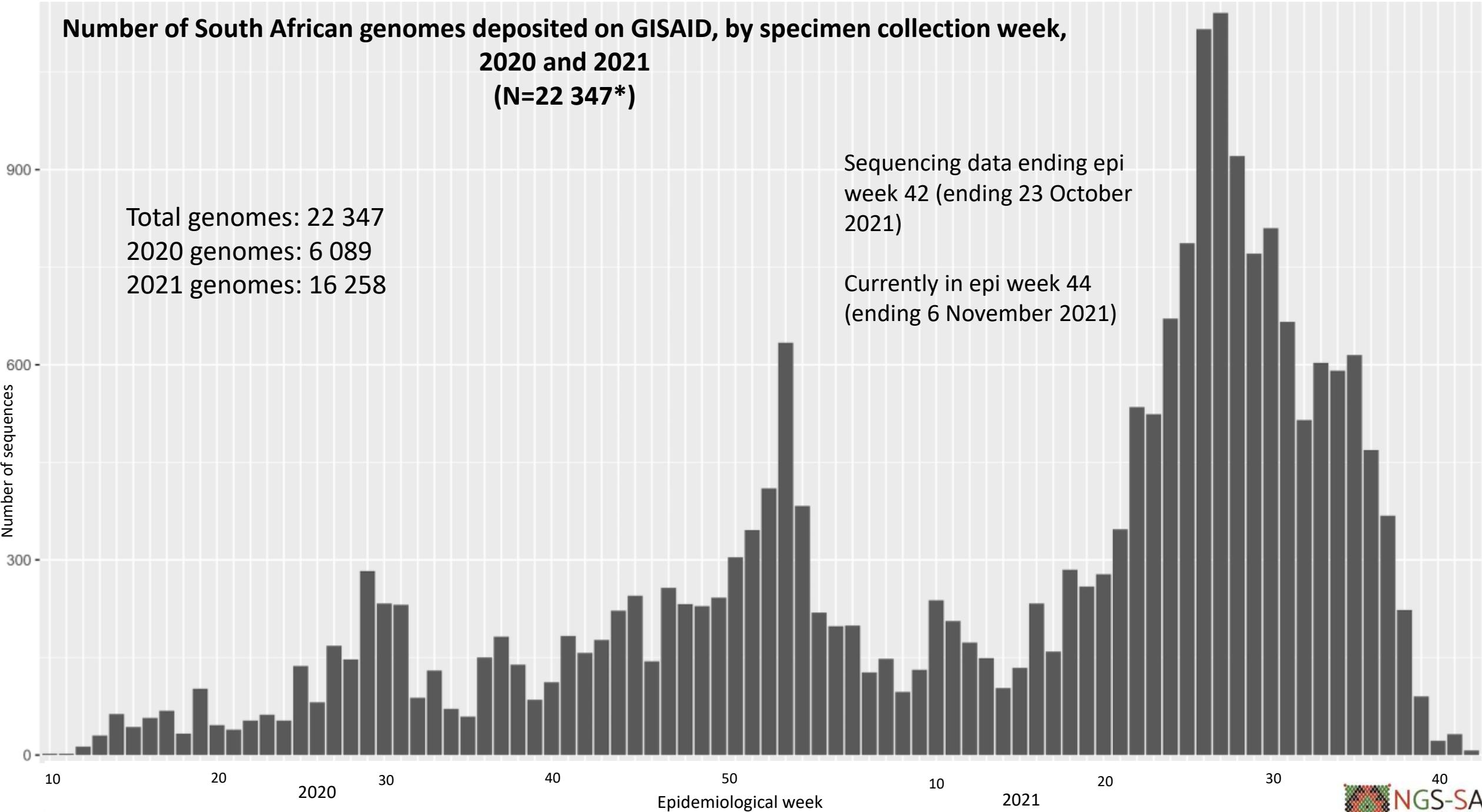


Bar graphs represent genomes sequenced per epiweek, with lines representing cases by collection date (weeks 18 – 43)

Genomes and cases presented as provincial total (percentage of national total) for epiweeks 18 – 43

PTP: percentage testing positive

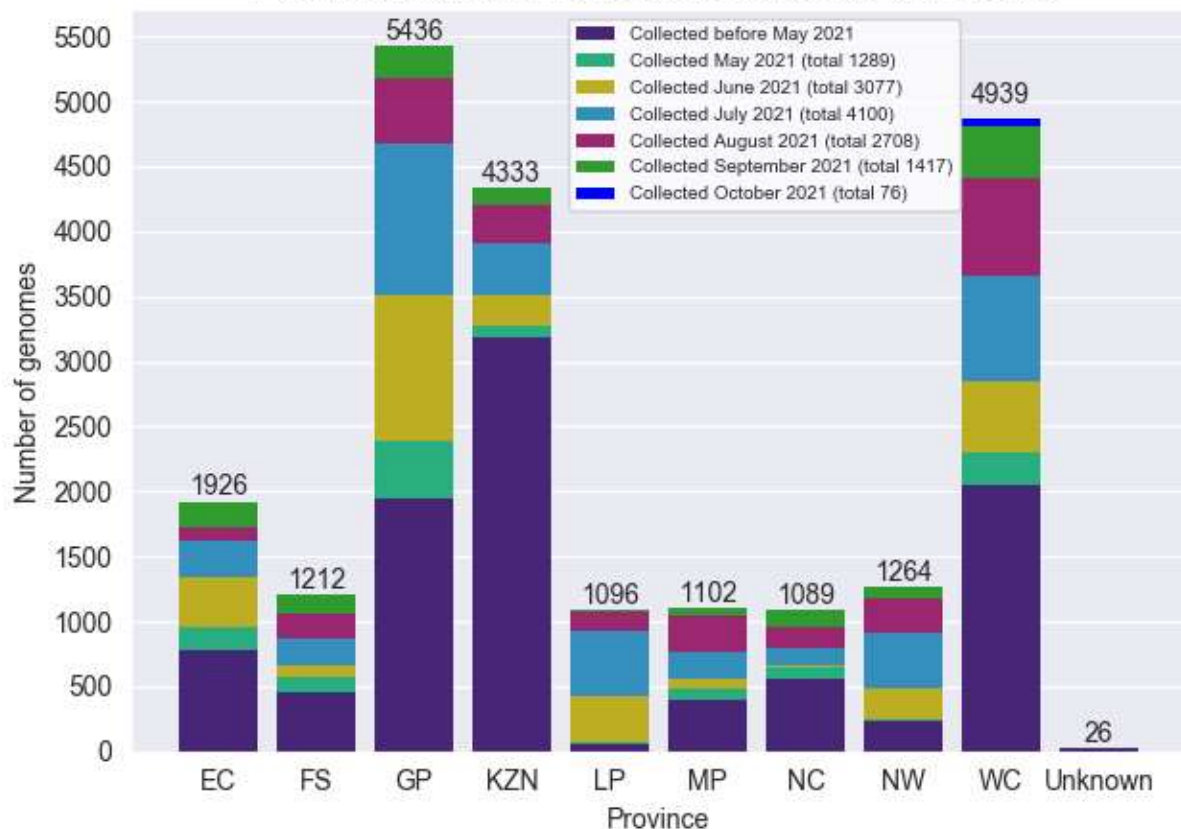
Number of South African genomes deposited on GISAID, by specimen collection week,  
2020 and 2021  
(N=22 347\*)



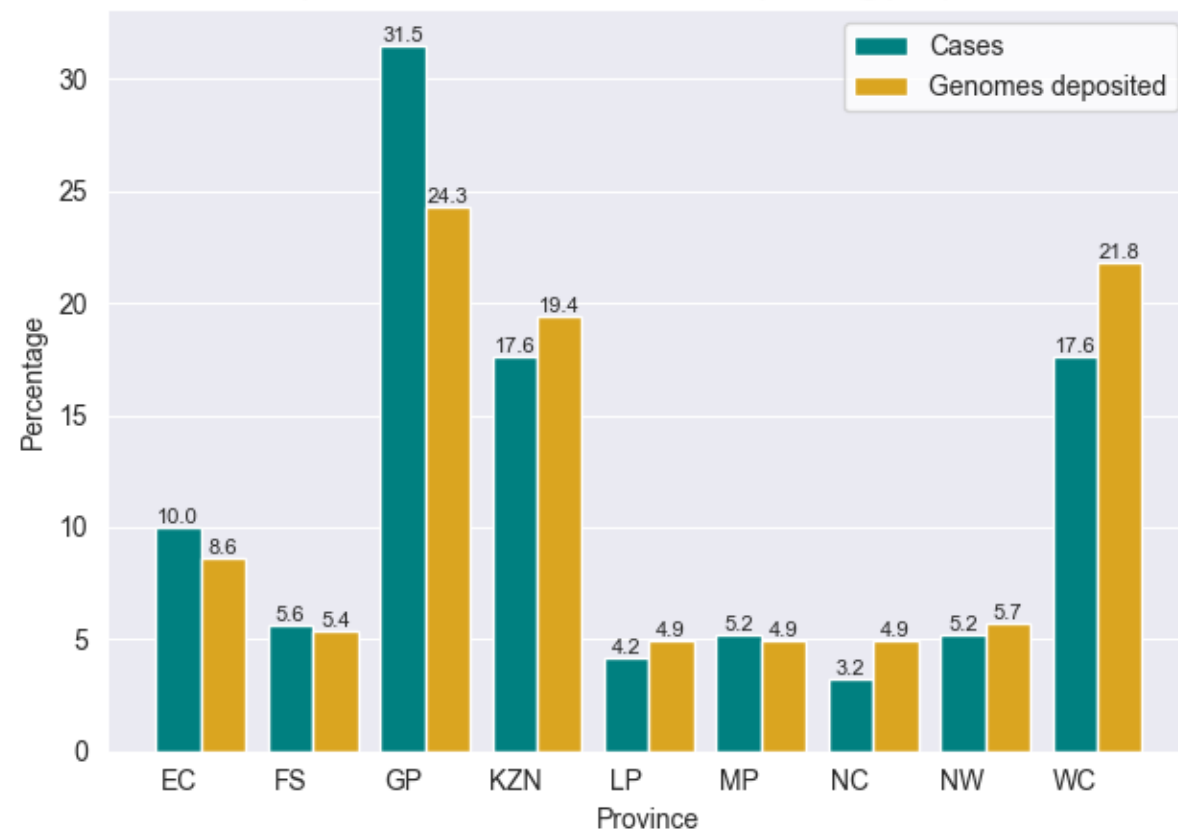
\*This represents the cleaned, de-duplicated dataset of unique sequences. This dataset will be used for all further figures.

# GISAID genomes vs total cases, 2020 and 2021 (N=22 347)

Provincial breakdown of genomes deposited into GISAID



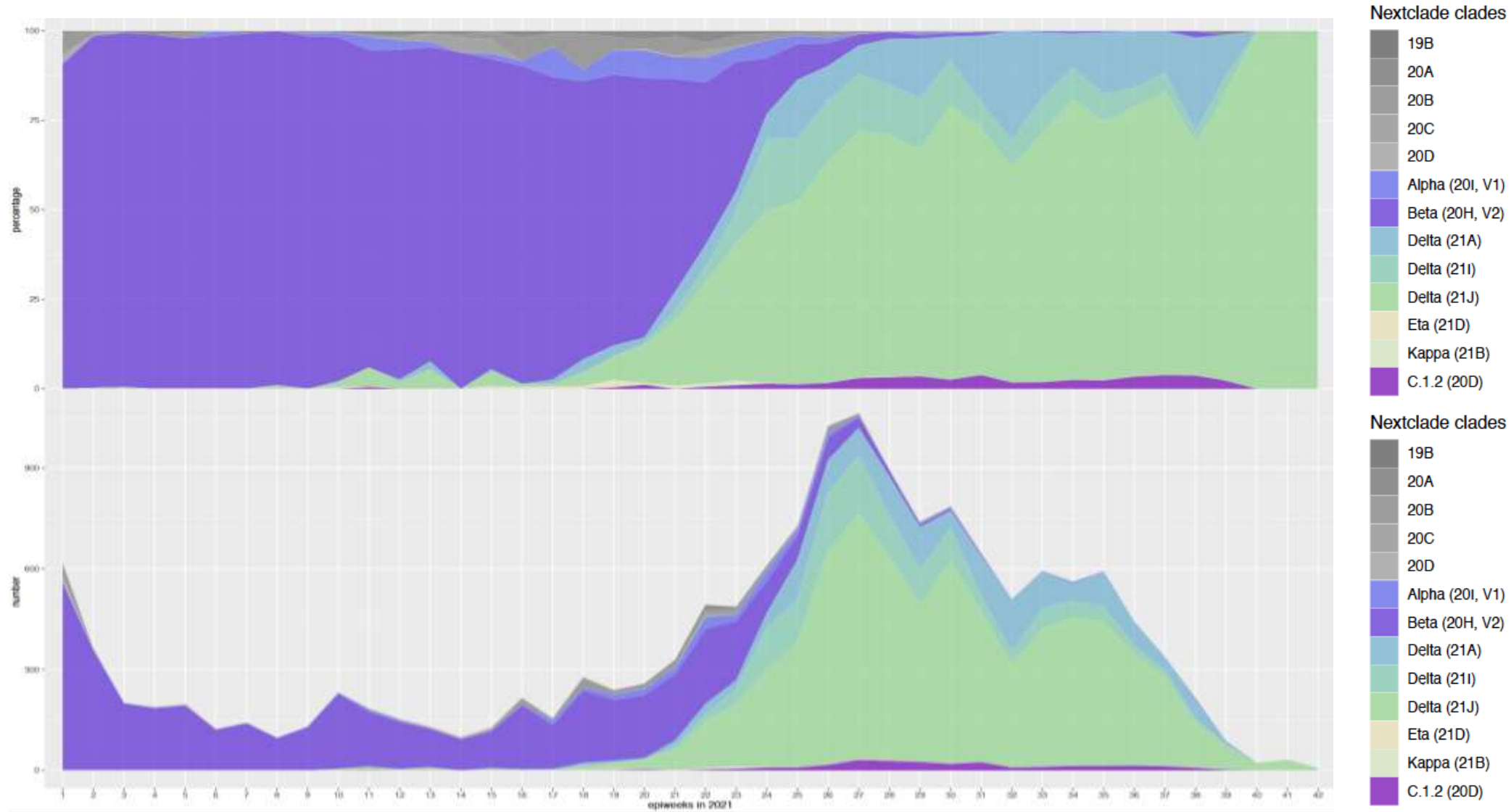
Comparison of total cases versus sequencing per province



All provinces, apart from GP, KZN, NC and WC, have comparable percentage of overall cases and overall sequenced genomes



# Distribution and number of clades in South Africa, 2021 (N= 16 258)



Delta has recently been split into three clades (21A, 21I, 21J).

Sequencing data ending epi week 42 (ending 23 October 2021)

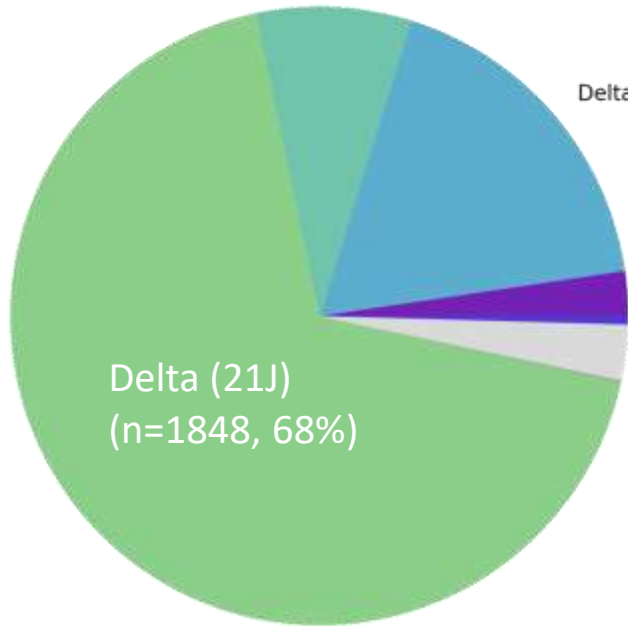
Currently in epi week 44 (ending 6 November 2021)

Delta came to dominate by end June at >65% , in July at >85% and in August and September at >90%  
C.1.2 present at <4% frequency since March

# Prevalence of Variants of Concern (VOC) and Variants of Interest (VOI) in August – October 2021 sequences, South Africa

August (N=2708)

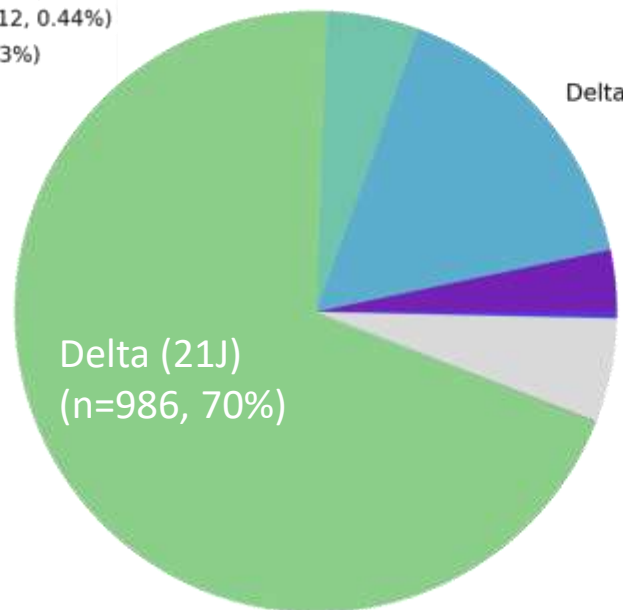
Delta (21I) (n=220, 8%)



Total Delta in Aug: 2551 (94%)

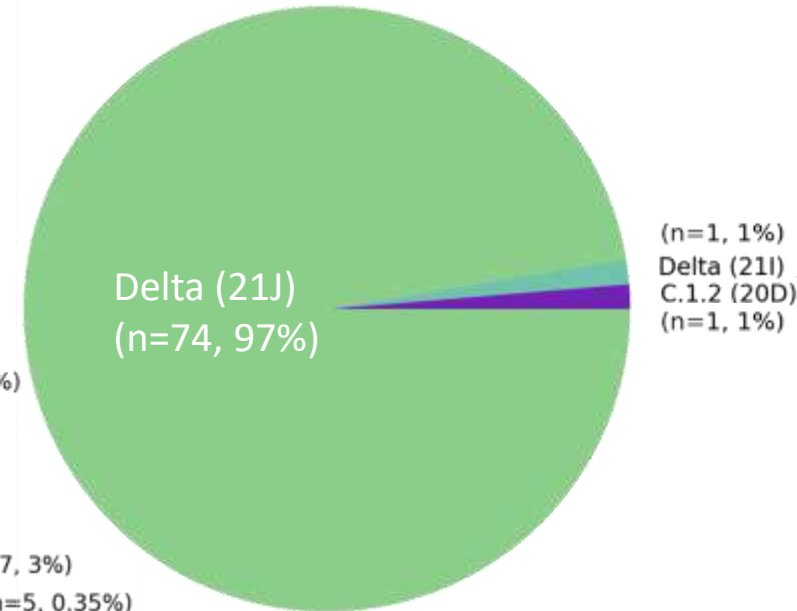
September (N=1417)

Delta (21I) (n=70, 5%)



Total Delta in Sep: 1285 (91%)

October (N=76)

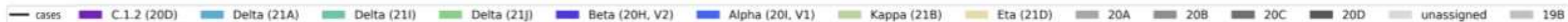
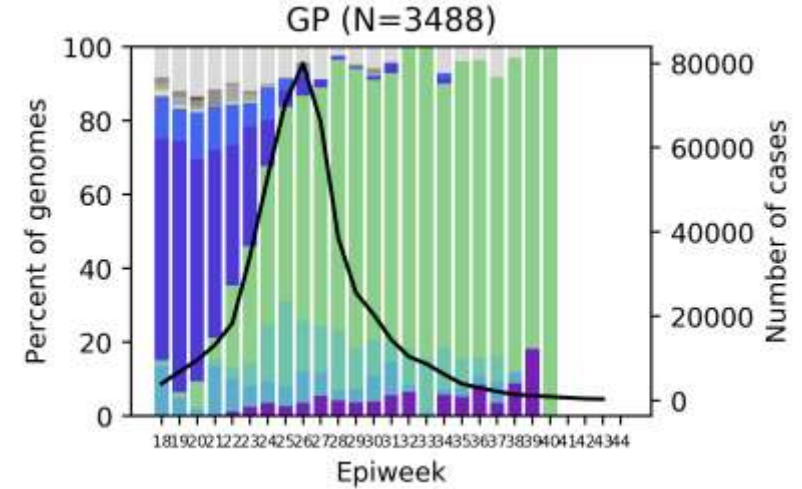
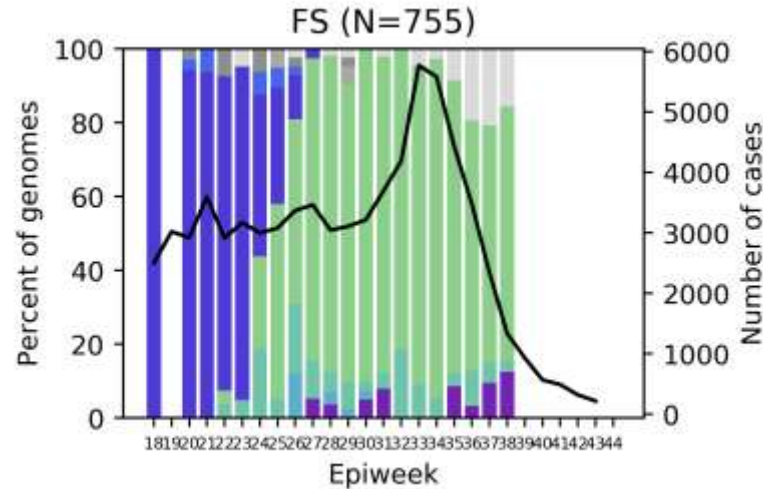
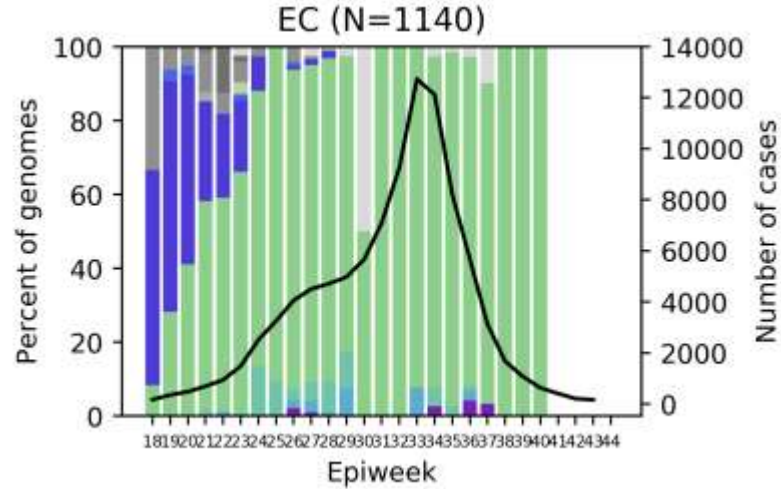
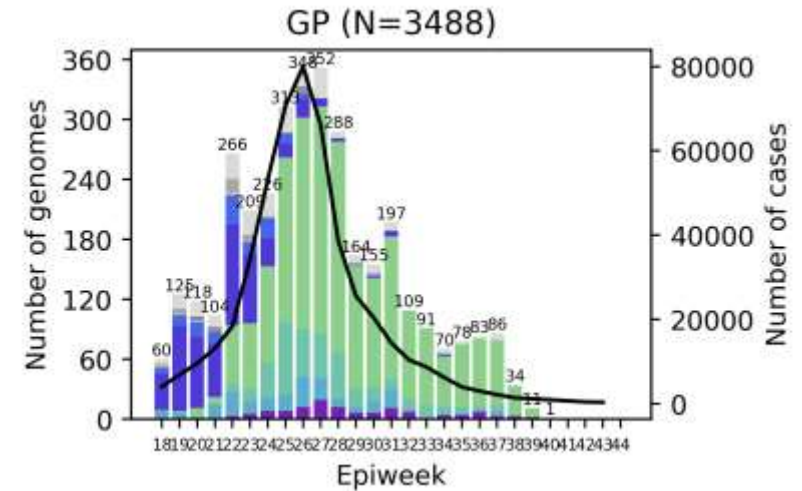
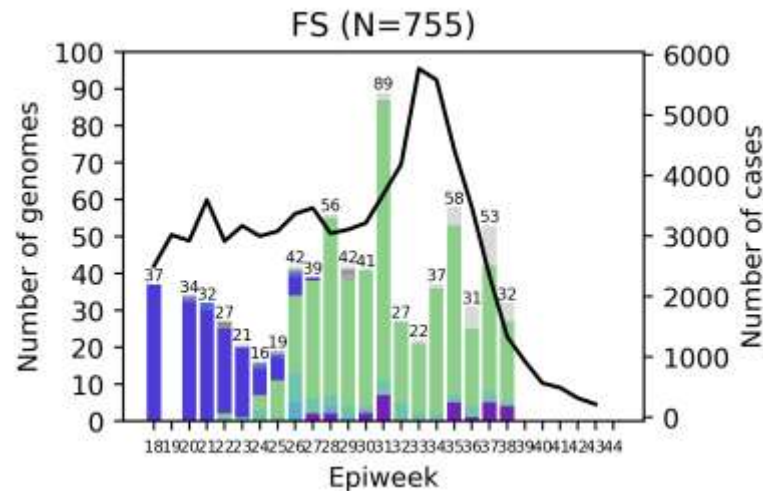
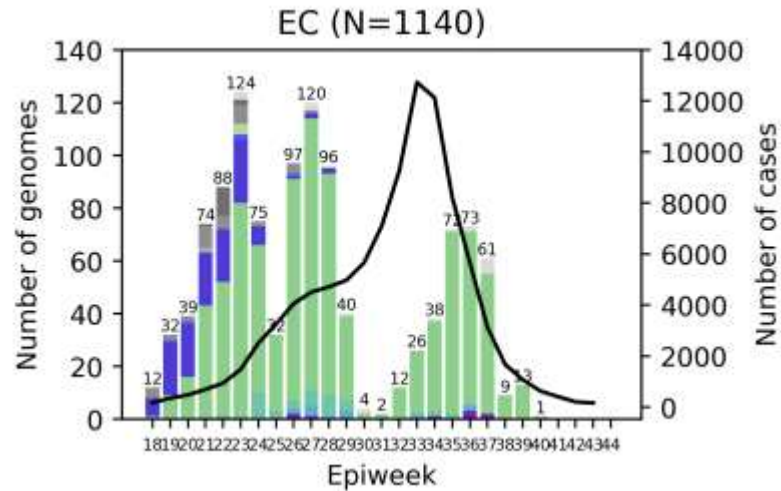


Total Delta in Oct: 75 (99%)

Beta (20H, V2) Alpha (20I, V1) Delta (21A) Delta (21I) Delta (21J) C.1.2 (20D) Kappa (21B) Eta (21D) Other unassigned

The Delta variant dominated August and September in South Africa, and appears to still dominate in October.

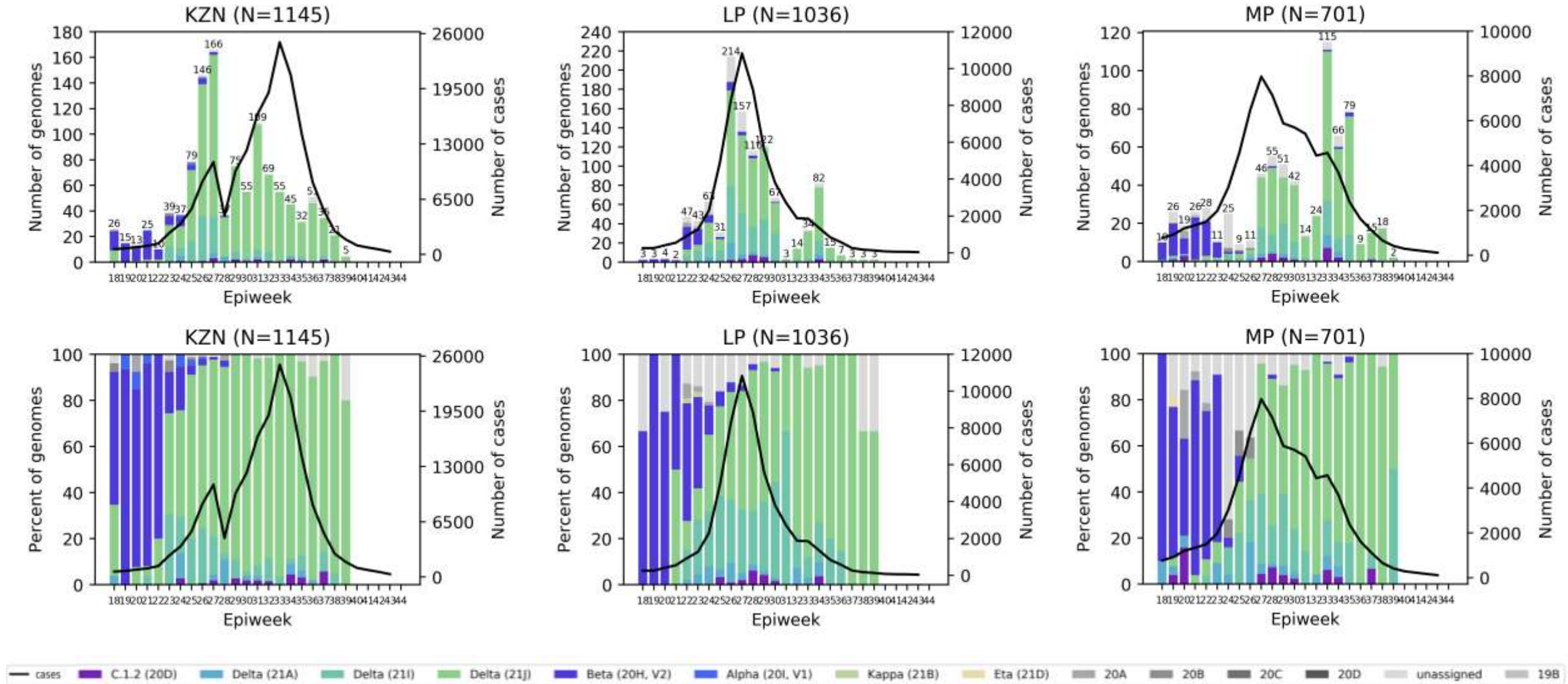
# Genomes sequenced from specimens collected in May to October 2021 (epiweeks 18 – 44) from Eastern Cape, Free State and Gauteng Provinces



Delta dominates the third wave in Eastern Cape, Free State and Gauteng provinces

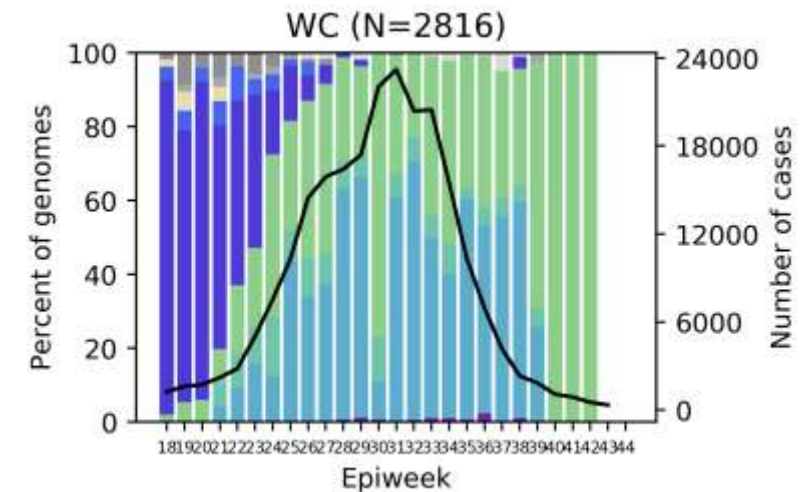
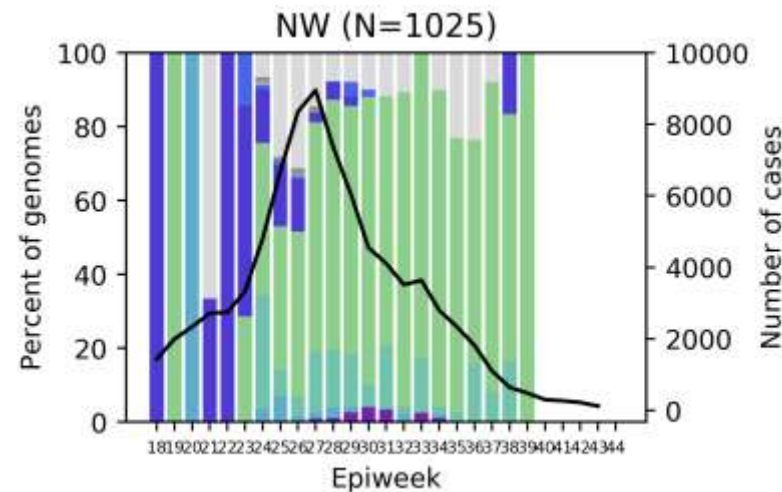
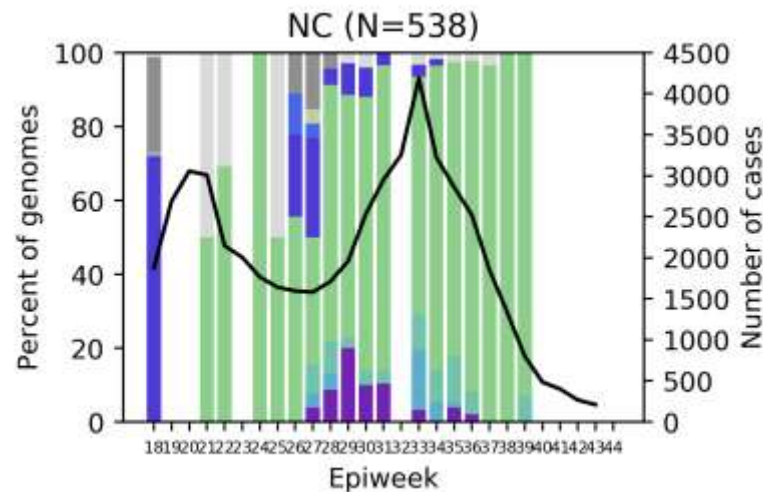
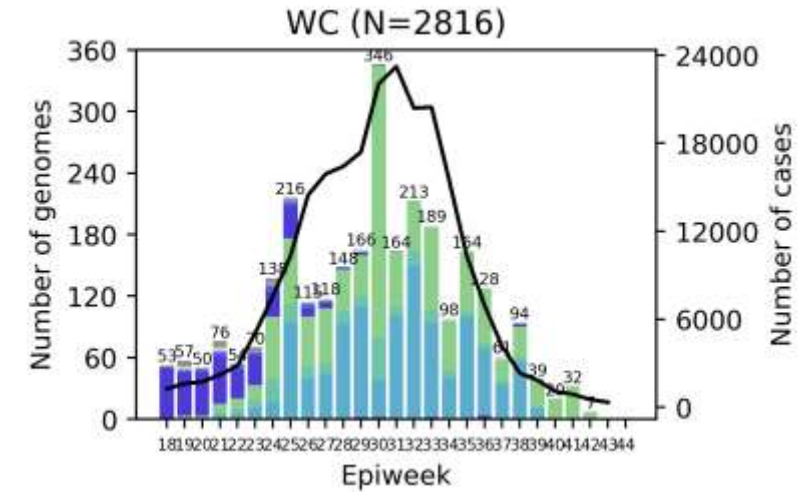
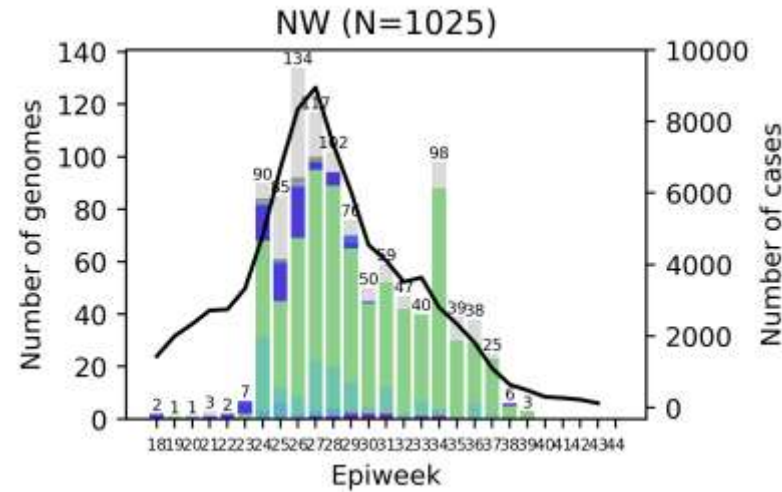
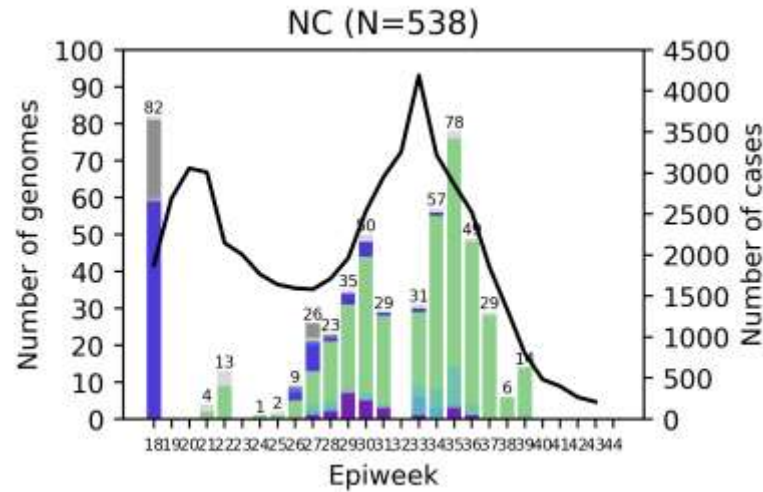


# Genomes sequenced from specimens collected in May to October 2021 (epiweeks 18 – 44) from KwaZulu-Natal, Limpopo and Mpumalanga Provinces



Delta variant dominates the third wave in KwaZulu-Natal, Limpopo and Mpumalanga Provinces

# Genomes sequenced from specimens collected in May to October 2021 (epiweeks 18 – 44) from Northern Cape, North West, and Western Cape Provinces

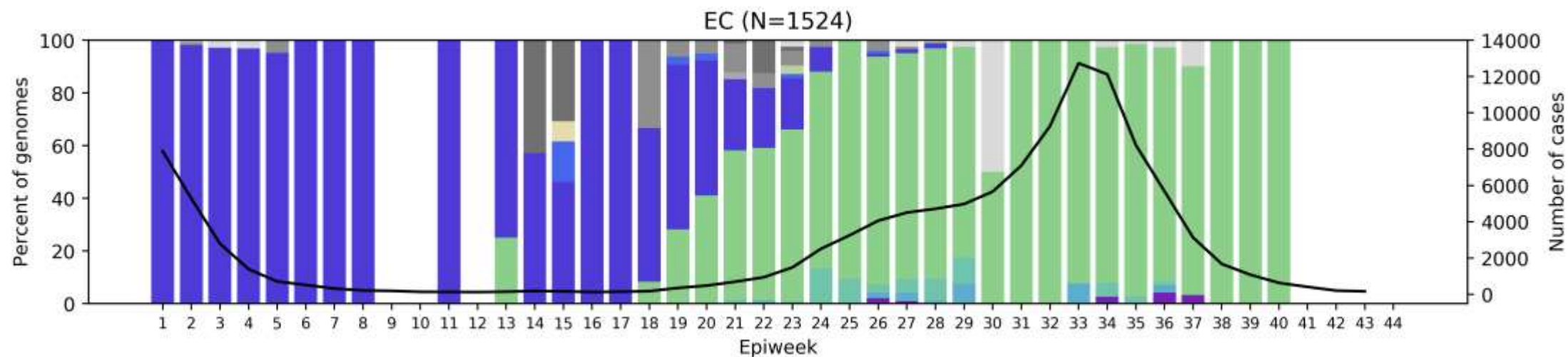
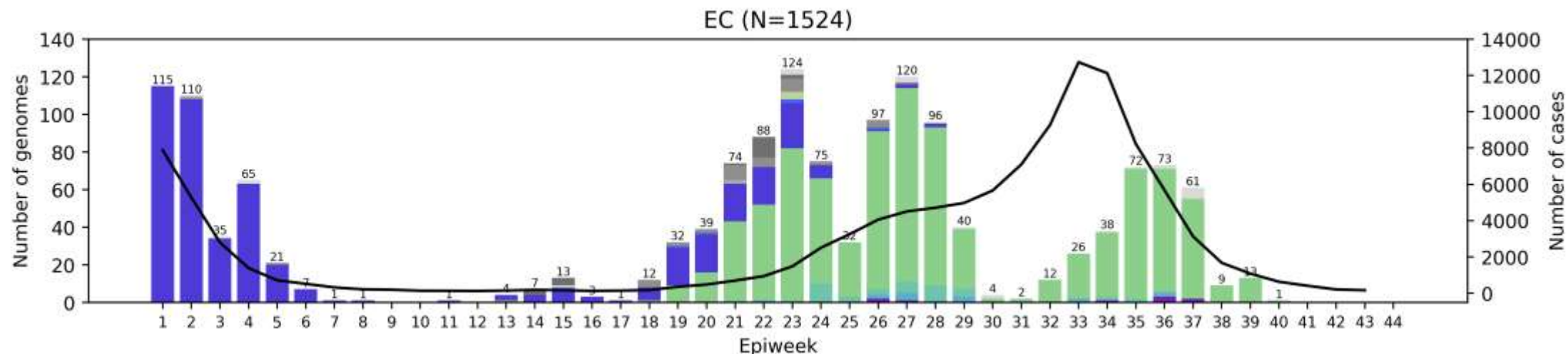


— cases — C.1.2 (20D) — Delta (21A) — Delta (21I) — Delta (21J) — Beta (20H, V2) — Alpha (20I, V1) — Kappa (21B) — Eta (21D) — 20A — 20B — 20C — 20D — unassigned — 19B

Delta dominates the third wave in Northern Cape, North West, and Western Cape provinces

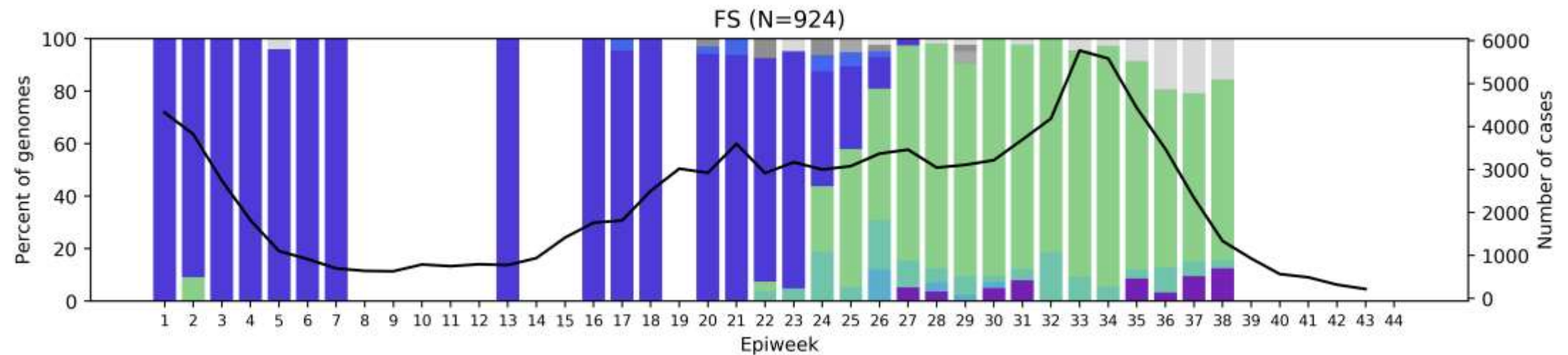
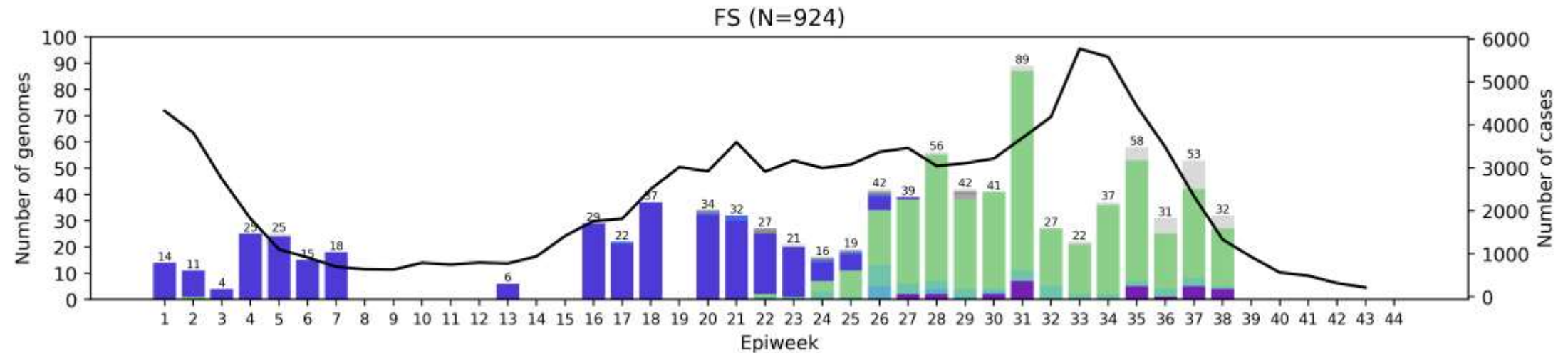


# Eastern Cape Province, 2021, n = 1524



— cases C.1.2 (20D) Delta (21A) Delta (21I) Delta (21J) Beta (20H, V2) Alpha (20I, V1) Kappa (21B) Eta (21D) 20A 20B 20C 20D unassigned 19B

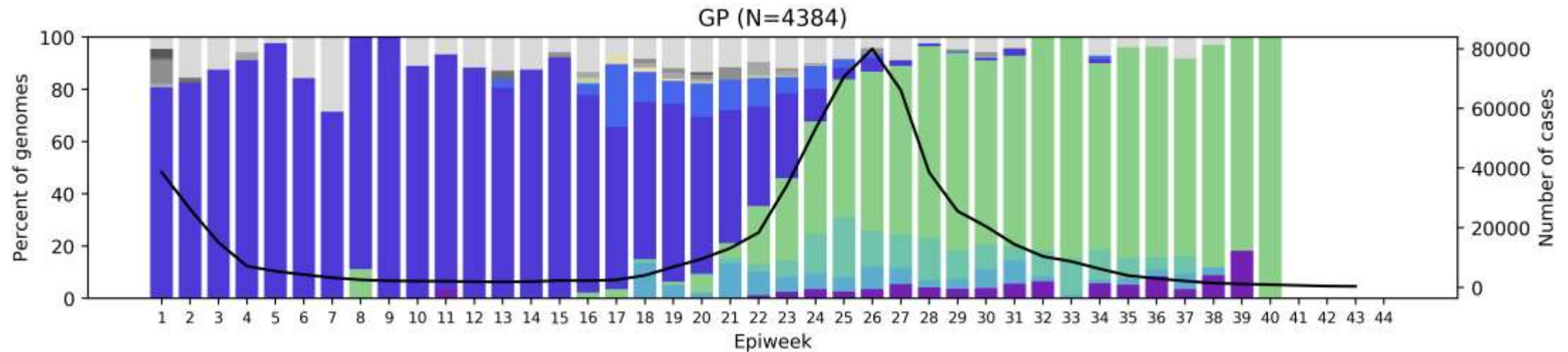
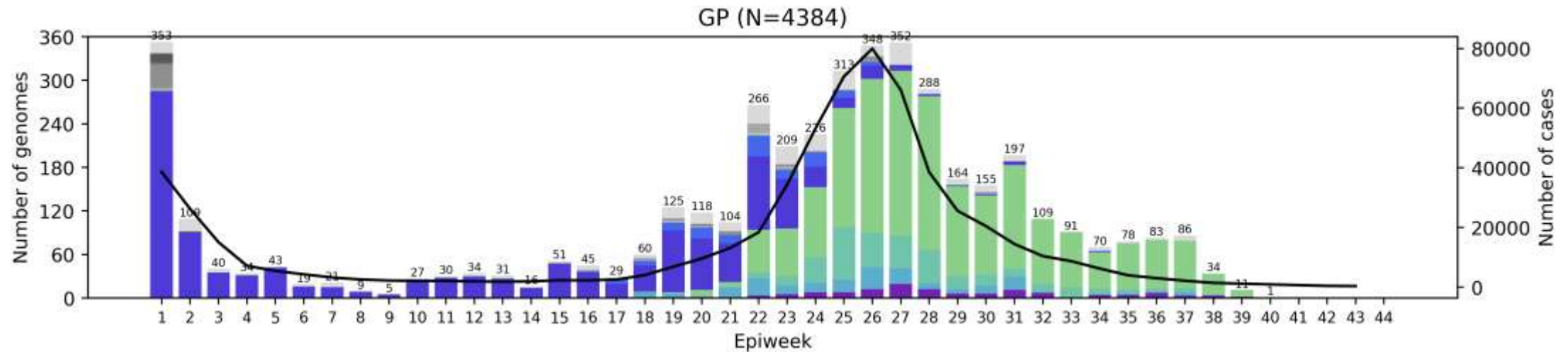
# Free State Province, 2021, n = 924



— cases   C.1.2 (20D)   Delta (21A)   Delta (21I)   Delta (21J)   Beta (20H, V2)   Alpha (20I, V1)   Kappa (21B)   Eta (21D)   20A   20B   20C   20D   unassigned   19B

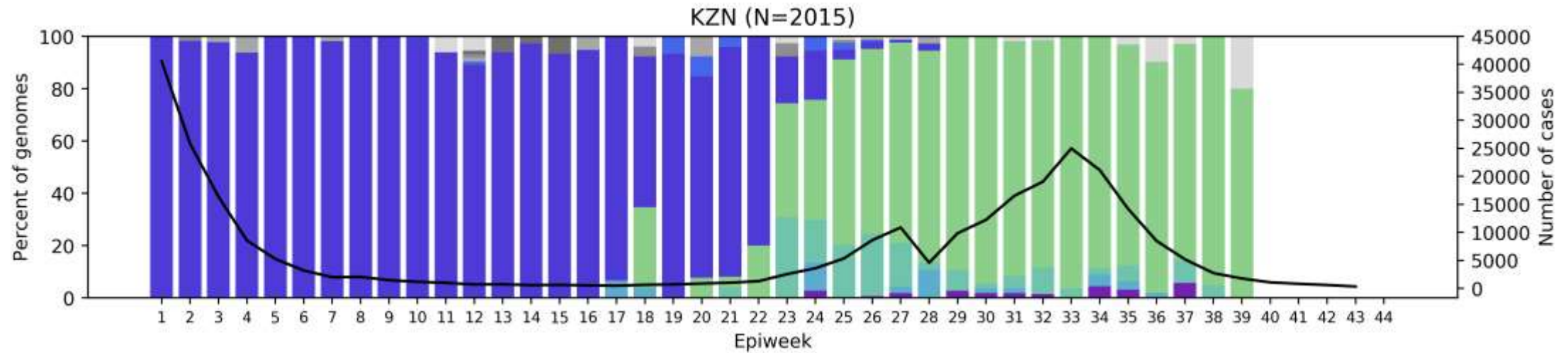
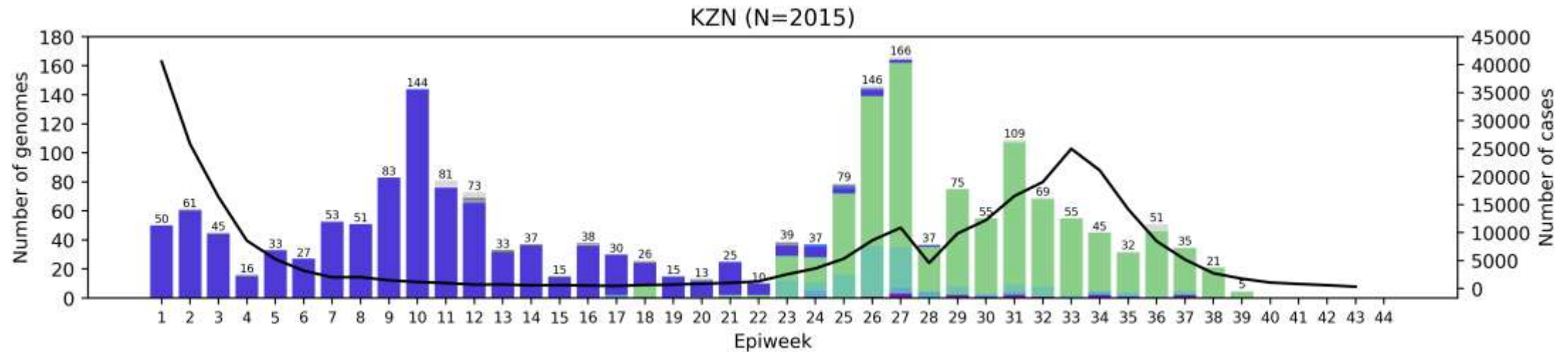


# Gauteng Province, 2021, n = 4384



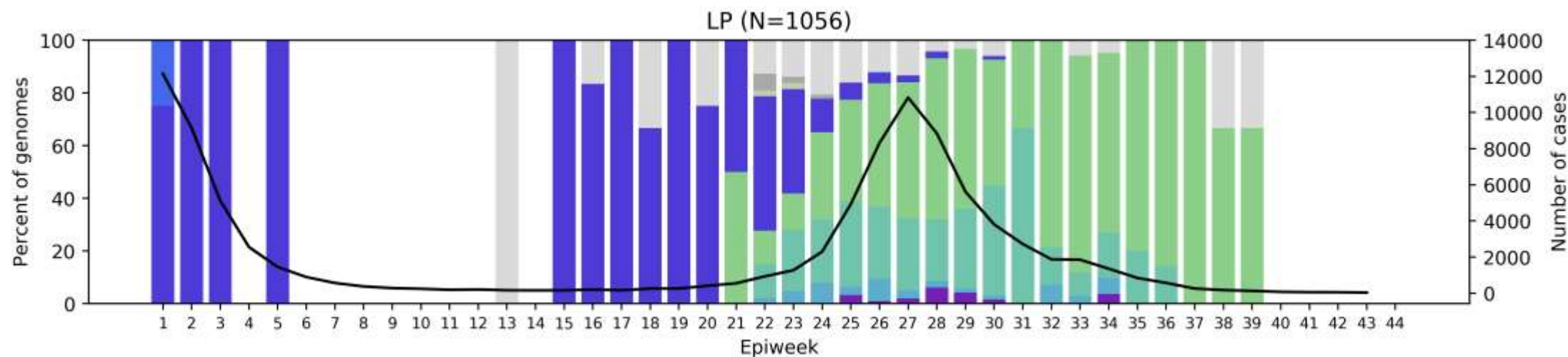
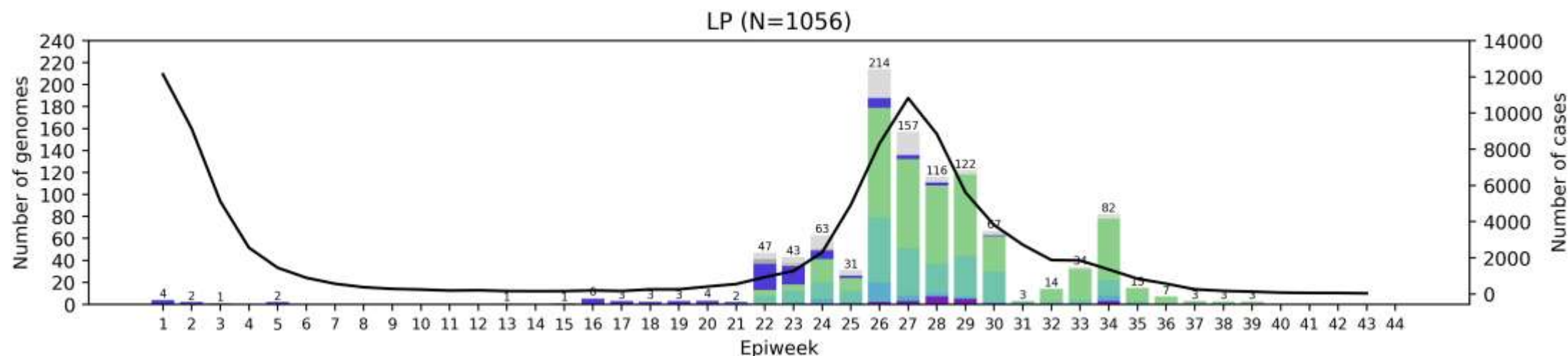
— cases C.1.2 (20D) Delta (21A) Delta (21I) Delta (21J) Beta (20H, V2) Alpha (20I, V1) Kappa (21B) Eta (21D) 20A 20B 20C 20D unassigned 19B

# KwaZulu-Natal Province, 2021, n = 2015



— cases C.1.2 (20D) Delta (21A) Delta (21I) Delta (21J) Beta (20H, V2) Alpha (20I, V1) Kappa (21B) Eta (21D) 20A 20B 20C 20D unassigned 19B

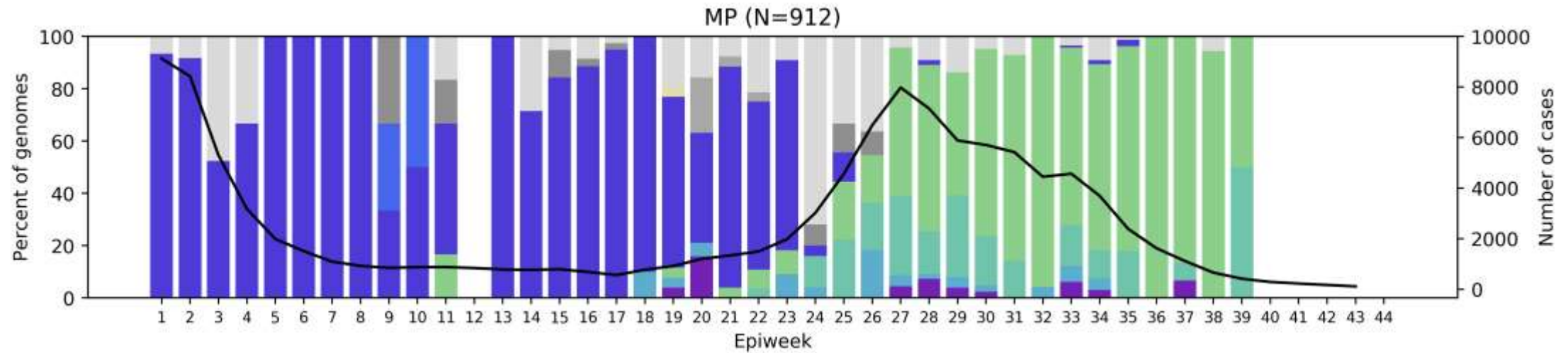
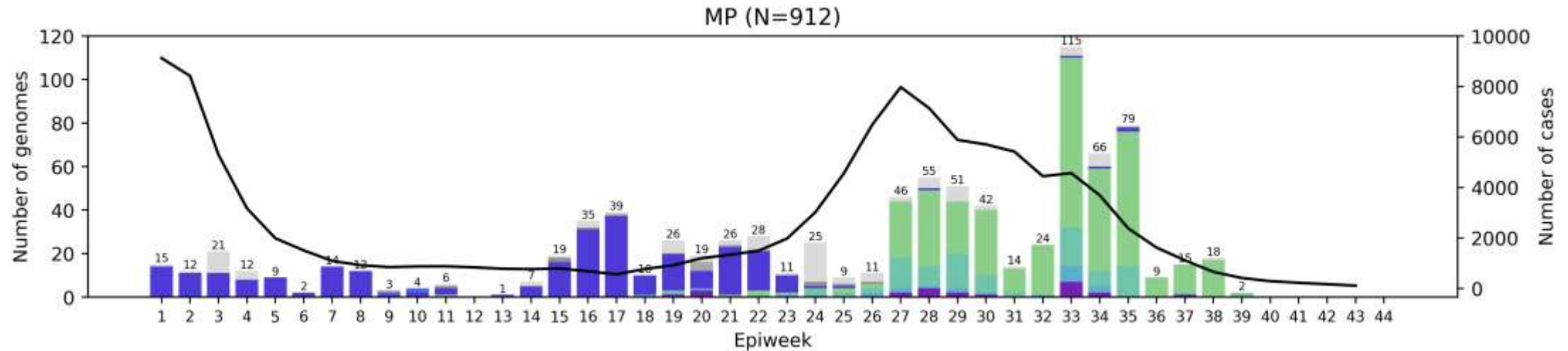
# Limpopo Province, 2021, n = 1056



— cases — C.1.2 (20D) — Delta (21A) — Delta (21I) — Delta (21J) — Beta (20H, V2) — Alpha (20I, V1) — Kappa (21B) — Eta (21D) — 20A — 20B — 20C — 20D — unassigned — 19B



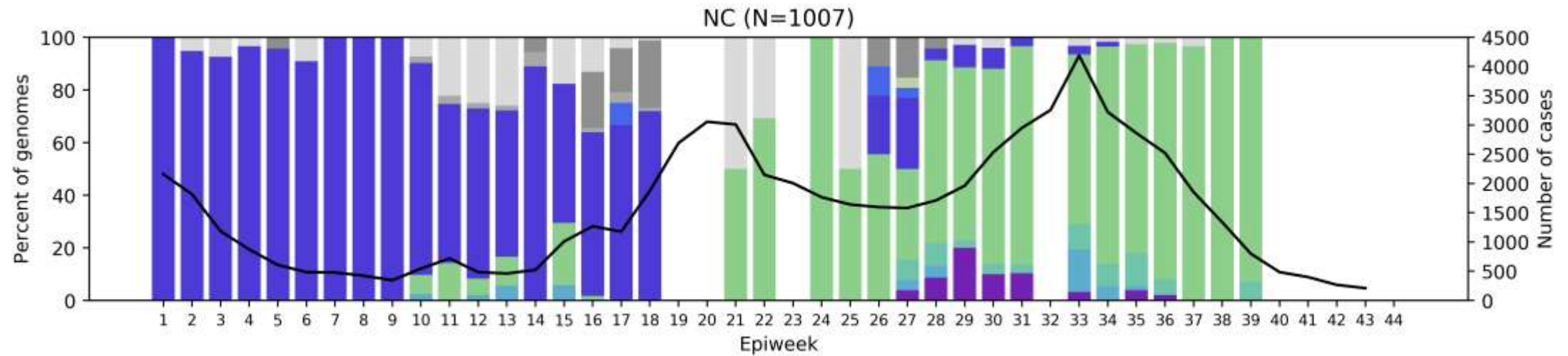
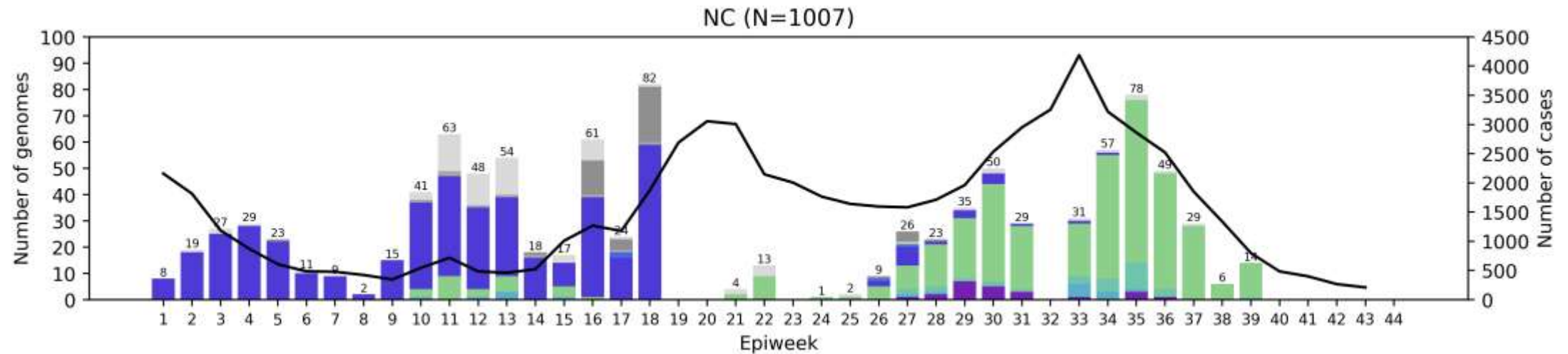
# Mpumalanga Province, 2021, n = 912



— cases C.1.2 (20D) Delta (21A) Delta (21I) Delta (21J) Beta (20H, V2) Alpha (20I, V1) Kappa (21B) Eta (21D) 20A 20B 20C 20D unassigned 19B

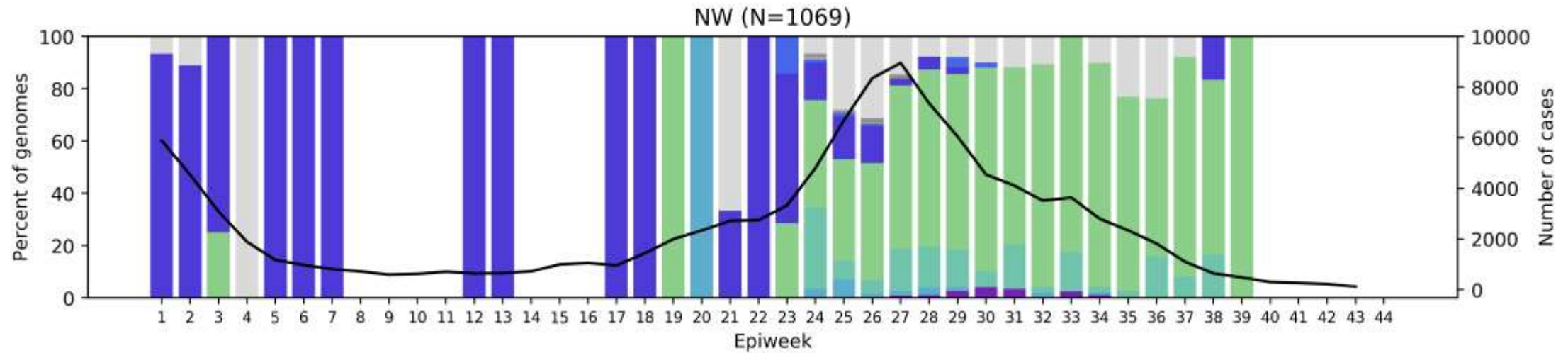
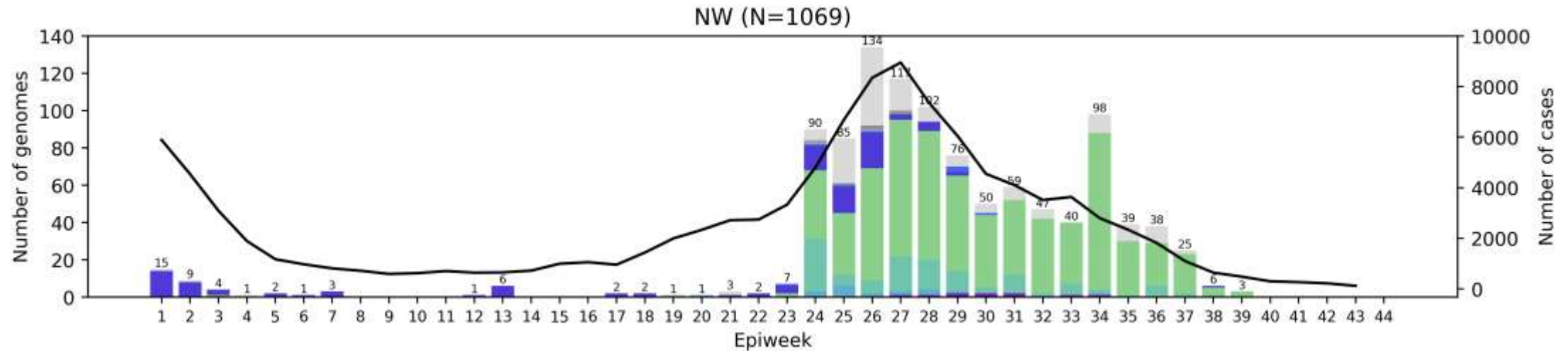


# Northern Cape Province, 2021, n = 929



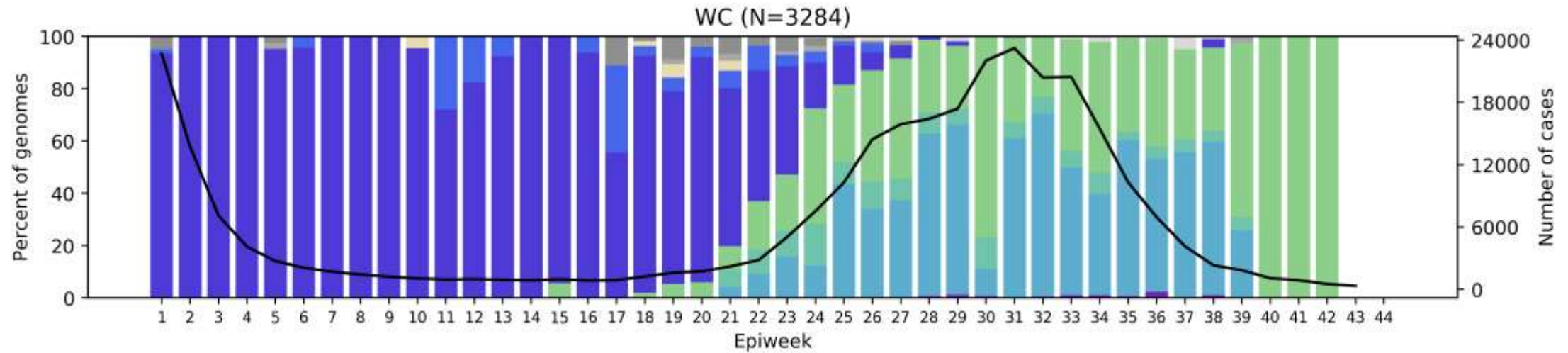
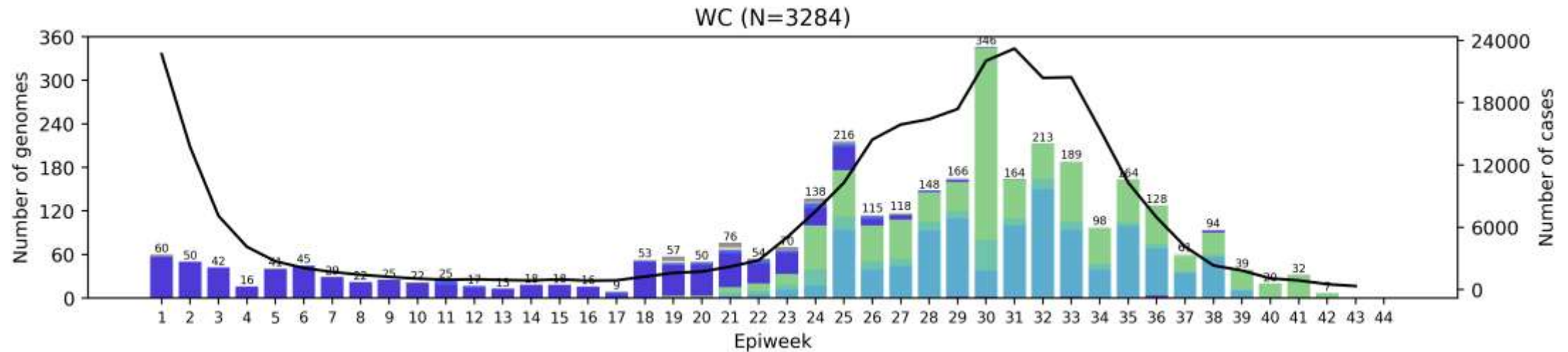
— cases C.1.2 (20D) Delta (21A) Delta (21I) Delta (21J) Beta (20H, V2) Alpha (20I, V1) Kappa (21B) Eta (21D) 20A 20B 20C 20D unassigned 19B

# North West Province, 2021, n = 970



— cases — C.1.2 (20D) — Delta (21A) — Delta (21I) — Delta (21J) — Beta (20H, V2) — Alpha (20I, V1) — Kappa (21B) — Eta (21D) — 20A — 20B — 20C — 20D — unassigned — 19B

# Western Cape Province, 2021, n = 3284

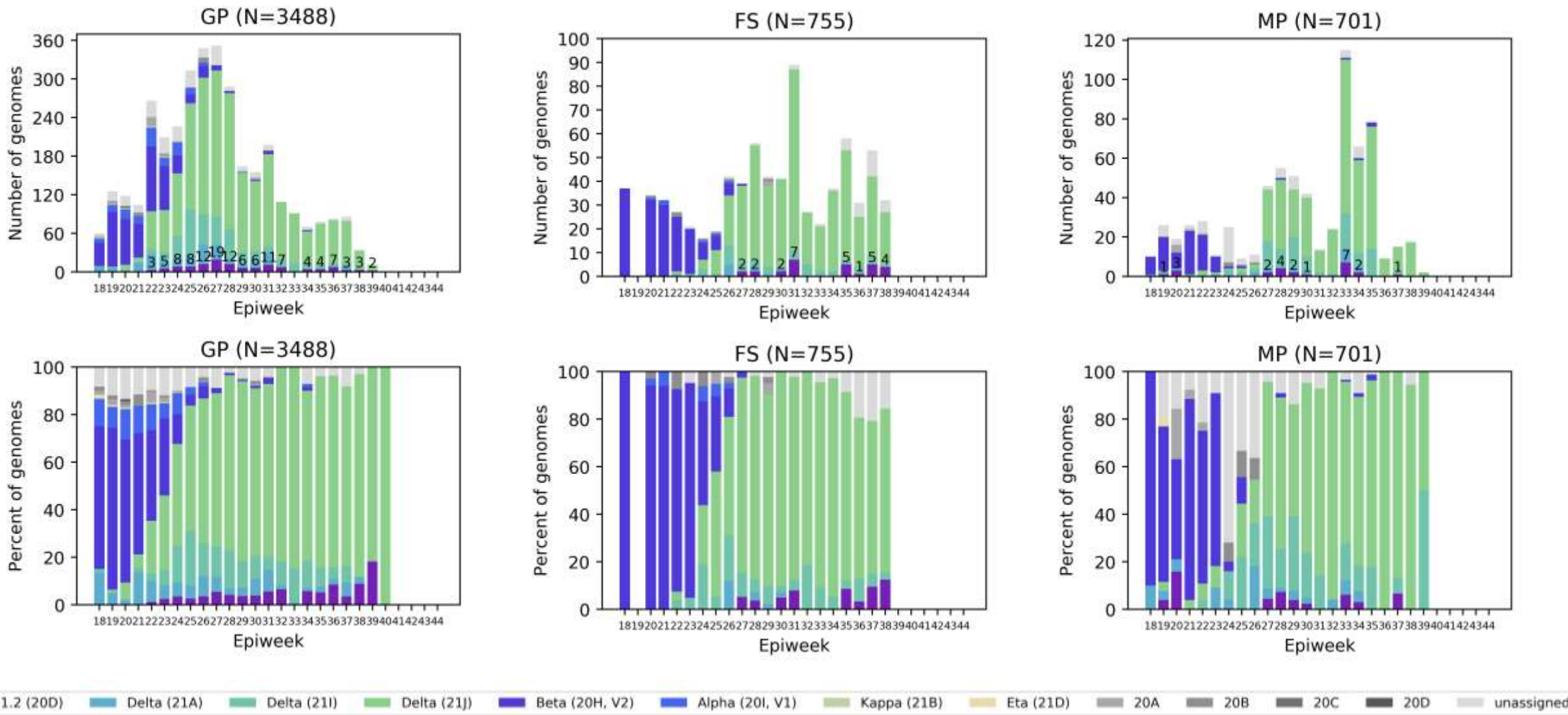


— cases   C.1.2 (20D)   Delta (21A)   Delta (21I)   Delta (21J)   Beta (20H, V2)   Alpha (20I, V1)   Kappa (21B)   Eta (21D)   20A   20B   20C   20D   unassigned   19B



# C.1.2 (n=265 in SA) in May – October 2021 by epiweek

Number of C.1.2 samples indicated above bar, provinces ordered by number of detections



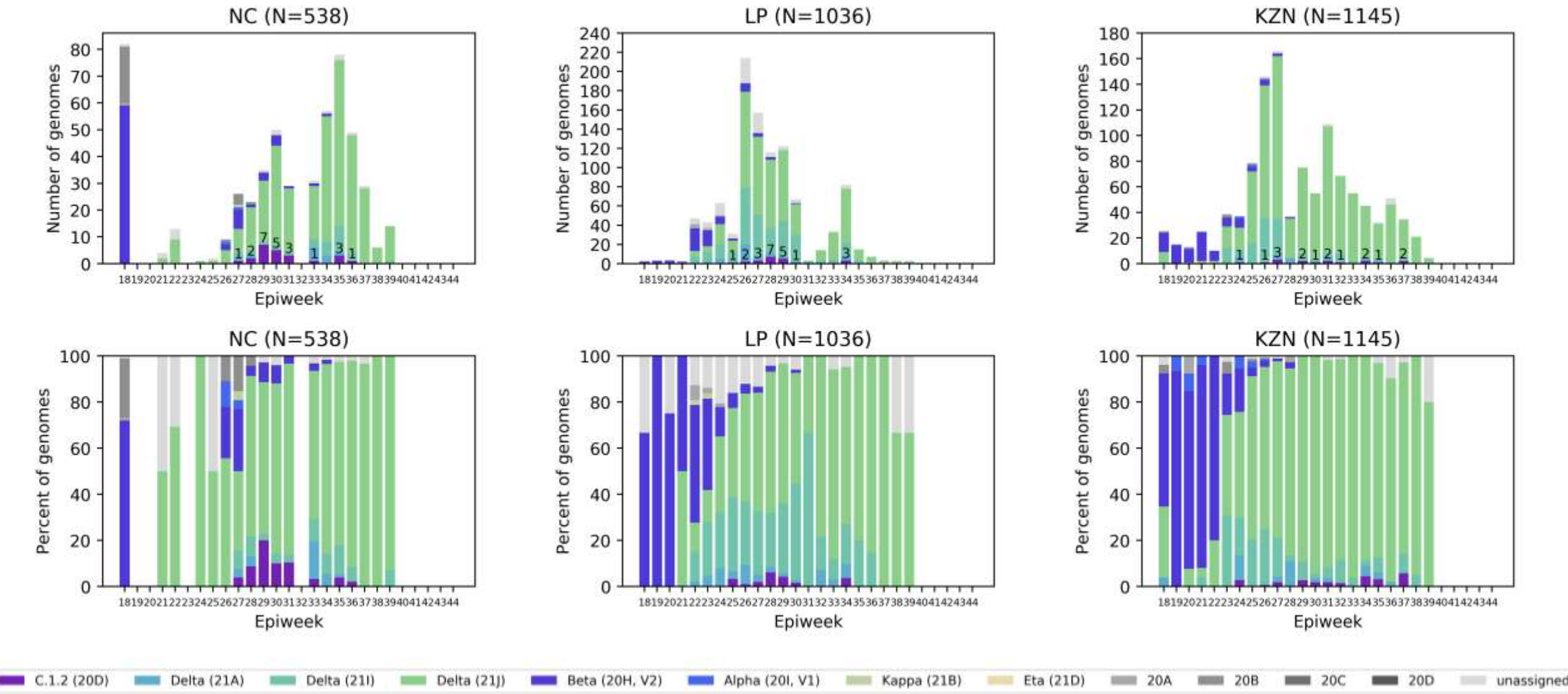
The majority of C.1.2 sequences have been detected in Gauteng (n=120), followed by the Free State (n=28) and then Mpumalanga (n=23).





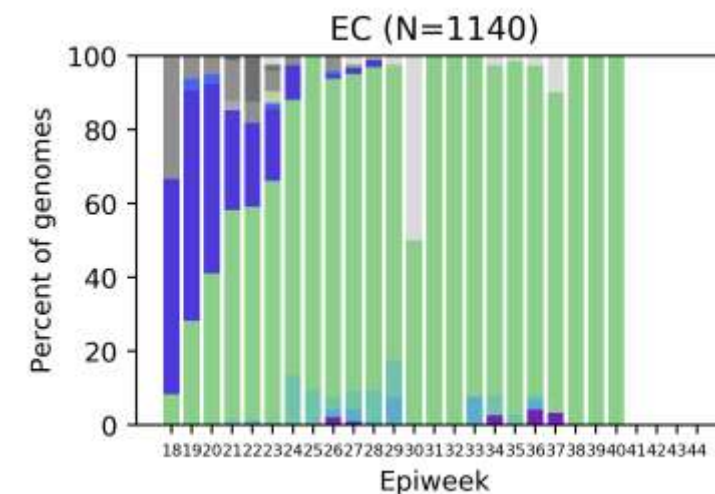
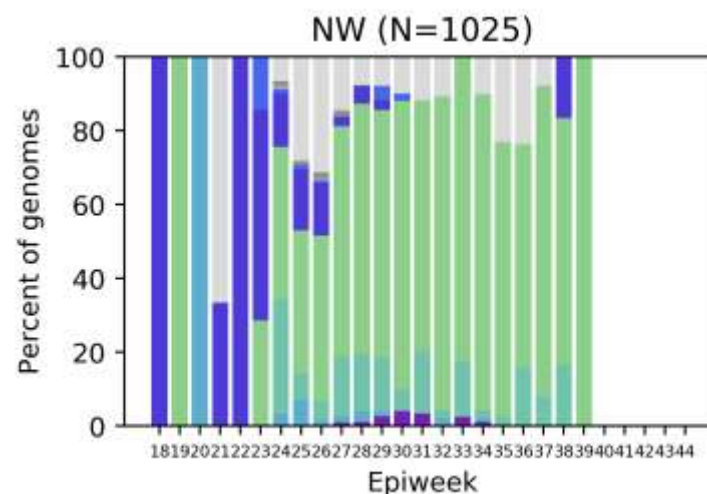
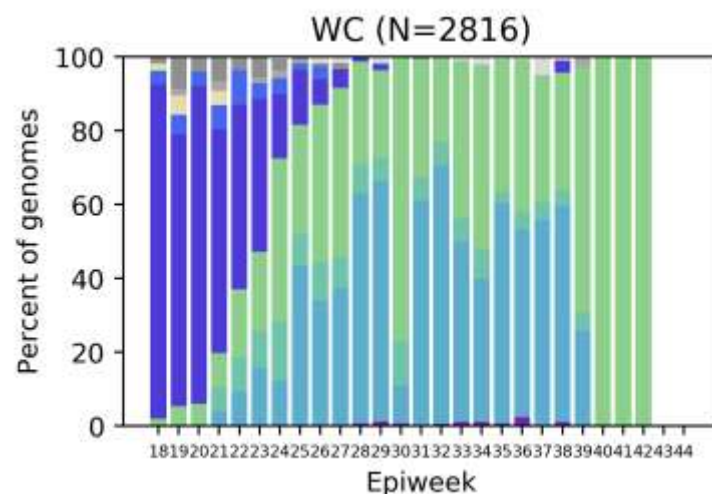
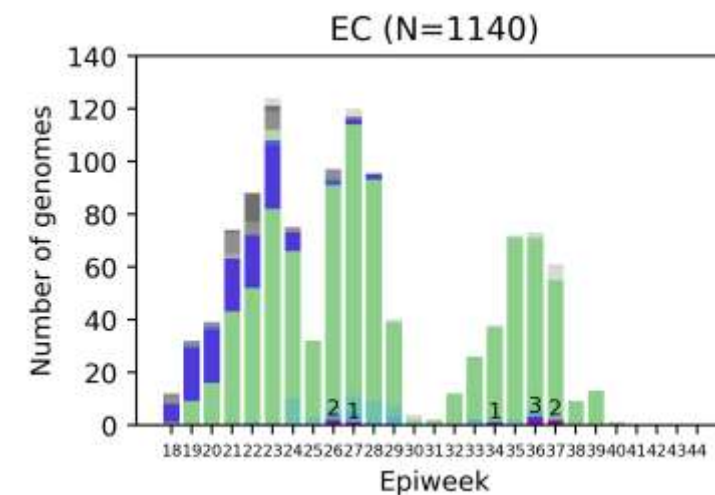
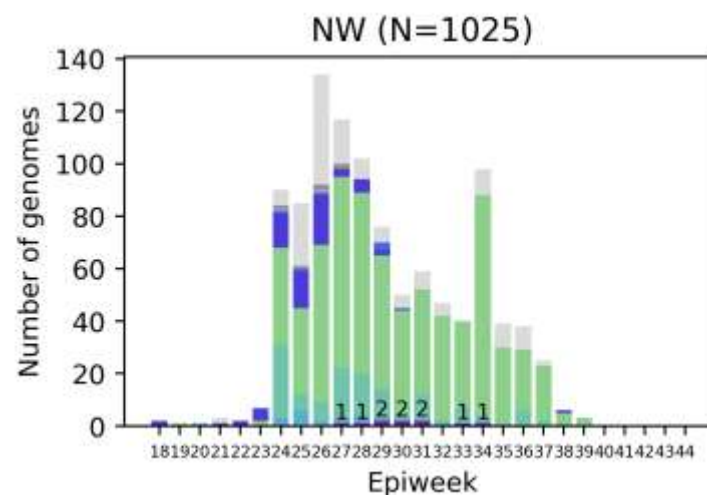
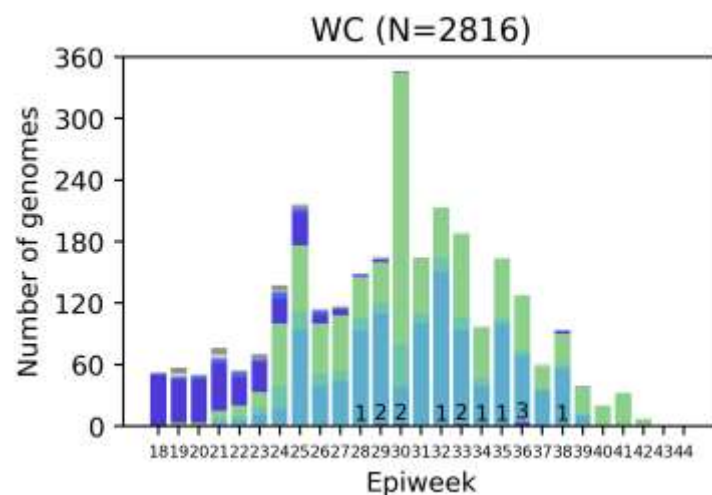
# C.1.2 (n=265 in SA) in May – October 2021 by epiweek

Number of C.1.2 samples indicated above bar, provinces ordered by number of detections



23 C.1.2 sequences have been detected in the Northern Cape, 22 in the Limpopo, and 16 in KwaZulu-Natal.

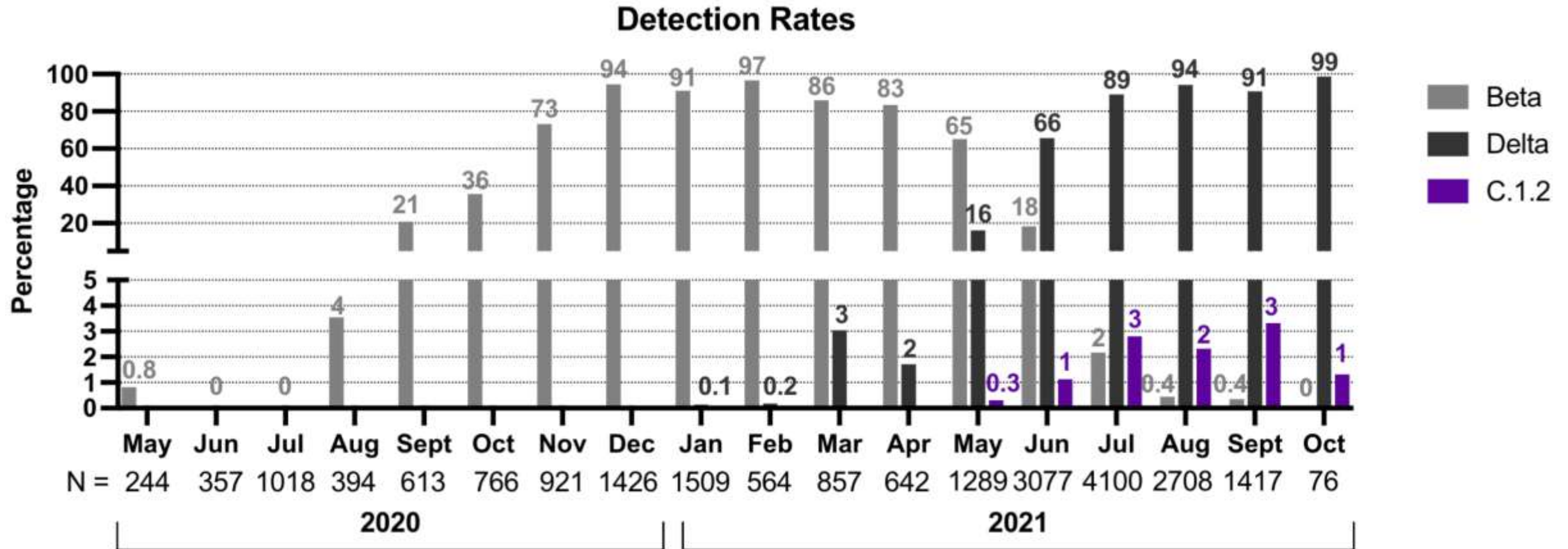
Number of C.1.2 samples indicated above bar, provinces ordered by number of detections



C.1.2 (20D)
  Delta (21A)
  Delta (21I)
  Delta (21J)
  Beta (20H, V2)
  Alpha (20I, V1)
  Kappa (21B)
  Eta (21D)
  20A
  20B
  20C
  20D
  unassigned

**The Western Cape has 14 sequences, the North West has 10, and the Eastern Cape has 9 detections of C.1.2.**

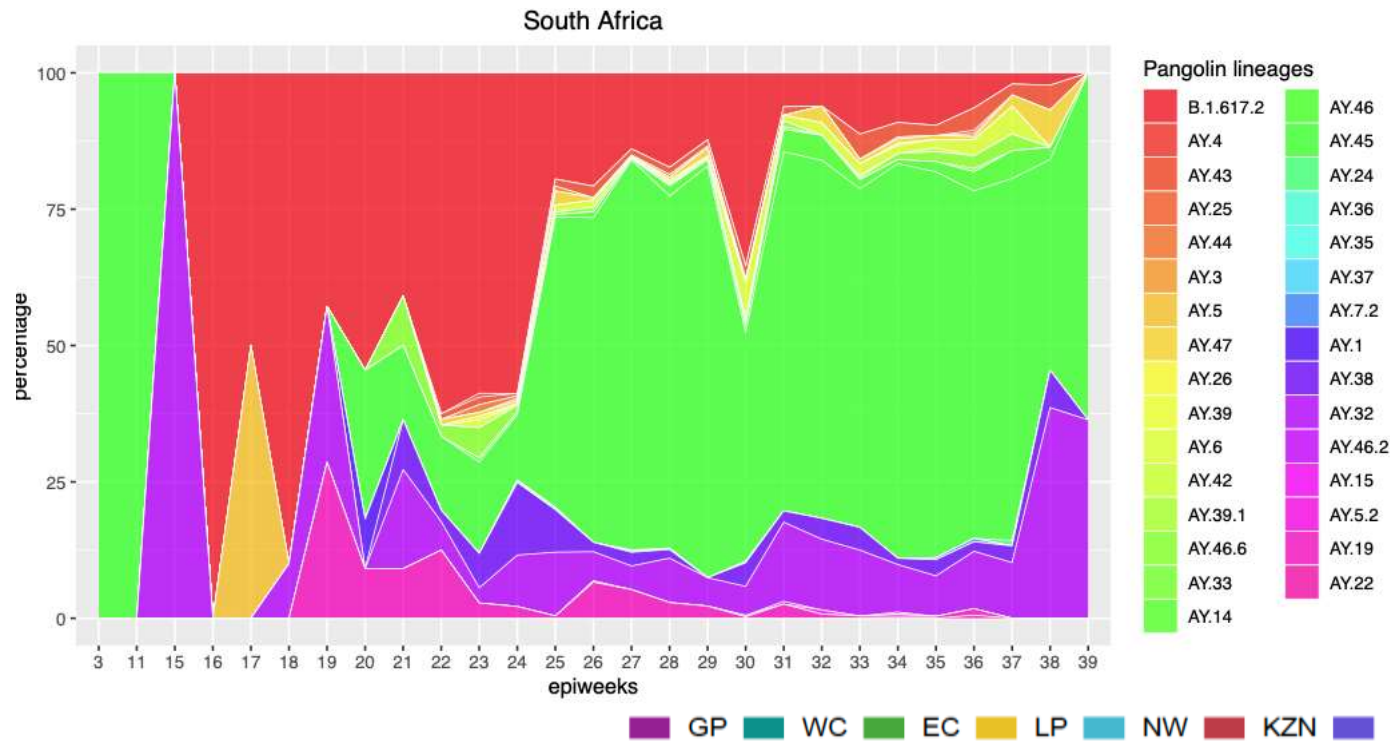
## C.1.2 growth compared to Beta and Delta



C.1.2 continues to be detected at low levels (less than 4% of genomes per month)

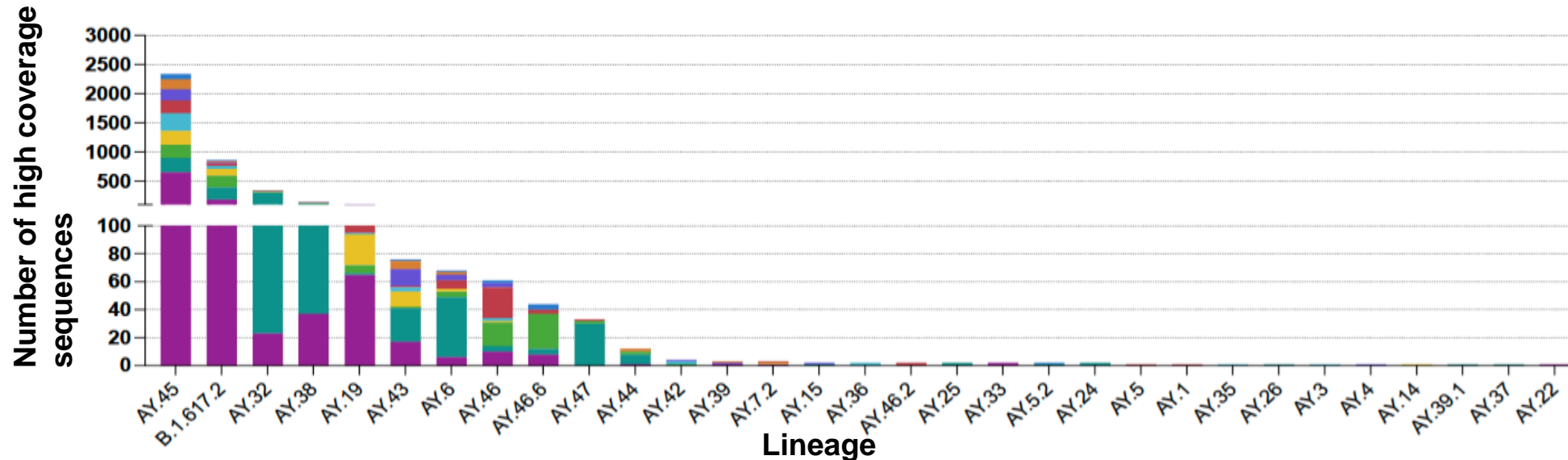


# Delta sub-lineages\* in South Africa



**Delta in South Africa is dominated by the AY.45, B.1.617.2, AY.32, AY.38 and AY.19 sub-lineages, although the dominance of sub-lineages varies with province. None of these are known to have a fitness advantage.**

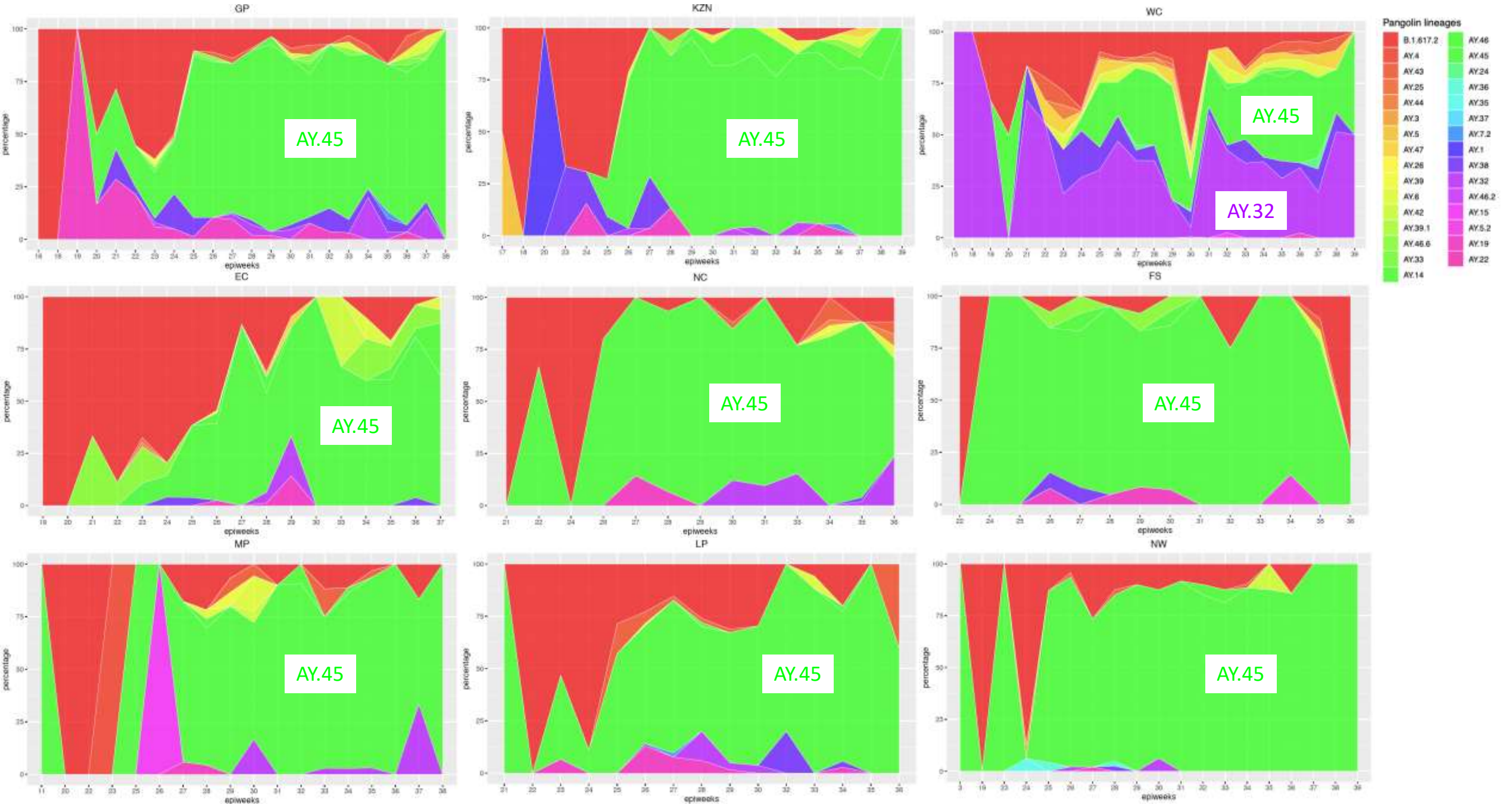
\*Data sourced from the GISAID variant surveillance file, high coverage sequences only, lineages assigned using lineages version 2021-10-18





Delta by province (epiweeks 16 – 38)

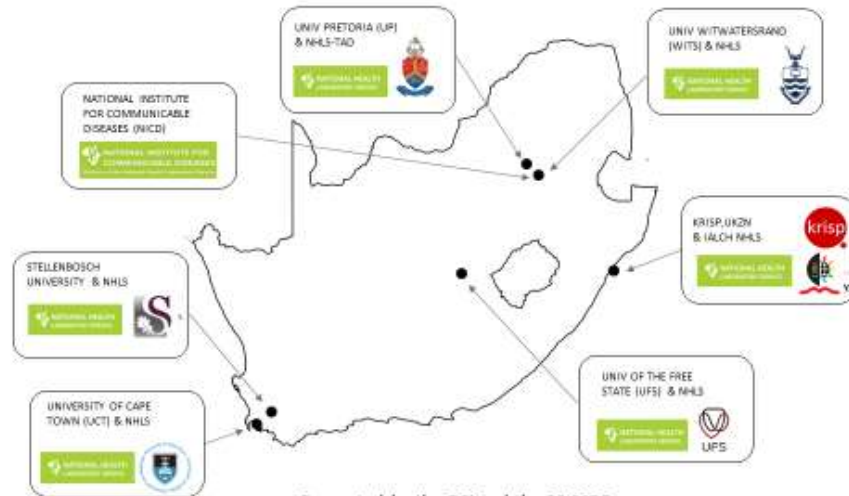
The Western Cape has a different distribution of AY lineages, with AY.32 more dominant than elsewhere



# Summary

- Delta continues to dominate in all provinces from specimens collected in September and October
  - Delta has recently been split into three clades.
    - New clades have been assigned due to >20% global circulation of particular sequences for more than 2 months
    - All SA Delta samples have been updated
    - Delta 21J is the dominant clade globally and in SA
    - The Delta sub-lineages vary by province
    - Based on updated definitions of AY.4.2<sup>1,2</sup>, this sub-lineage has not been detected in the country
- Mutated C.1.2 lineage detected in all provinces of South Africa at less than 4% of genomes
- Lambda and Mu variants not detected in South Africa

1. <https://github.com/cov-lineages/constellations/blob/main/constellations/definitions/cAY.4.2.json>  
2. [https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment\\_data/file/1028113/Technical\\_Briefing\\_26.pdf](https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/1028113/Technical_Briefing_26.pdf)



UNIVERSITEIT VAN PRETORIA  
UNIVERSITY OF PRETORIA  
YUNIBESITHI YA PRETORIA



UNIVERSITEIT  
YUNIVESITHI  
STELLENBOSCH  
UNIVERSITY



science & innovation  
Department:  
Science and Innovation  
REPUBLIC OF SOUTH AFRICA



WITS  
UNIVERSITY



UNIVERSITY OF  
KWAZULU-NATAL<sup>TM</sup>  
INYUVESI  
YAKWAZULU-NATALI





# UKZN-Inkosi Albert Luthuli Central Hospital



Dr Khanyi Msomi  
Dr Kerusha Govender  
Dr Pravi Moodley  
Dr Aabida Khan  
Dr Lili Gounder  
Dr Kerri Francois  
Dr Cherise Naicker  
Dr Joedene Chetty  
Dr Neli Ngcaba  
Dr Tshepiso Mosito  
Mr Malcolm Ellapen  
Mr Kubendran Reddy  
The COVID-19 Bench team

# University of KwaZulu-Natal & Africa Health Research Institute



## KRISP at UKZN:

Tulio de Oliveira  
Richard Lessels  
Houriiyah Tegally  
Eduan Wilkinson  
Jennifer Giandhari  
Sureshnee Pillay  
Emmanuel James San



## AHRI

Alex Sigal  
Sandile Cele  
Willem Hanekom

# University of Stellenbosch & NHLS Tygerberg Virology



Susan Engelbrecht  
Wolfgang Preiser  
Gert van Zyl  
Tongai Maponga  
Bronwyn Kleinhans  
Shannon Wilson  
Karabo Phadu  
Tania Stander  
Kamela Mahlakwane  
Mathilda Claassen  
Diagnostic laboratory staff

# University of Cape Town, NHLS & WCG



## NHLS-UCT

Carolyn Williamson  
Nei-yuan Hsiao  
Diana Hardie  
Kruger Marais  
Stephen Korsman  
Ziyaad Valley-Omar

## WCG-UCT

Mary-Anne Davies  
Hannah Hussey  
Andrew Boulle  
Masudah Paleker  
Theuns Jacobs  
Erna Morden



EDCTP



## UCT, IDM and CIDRI-Africa

Deelan Doolabh  
Arash Iranzadeh  
Lynn Tyers  
Innocent Mudau  
Nokuzola Mbhele  
Fezokuhle Khumalo  
Thabang Serakge  
Bruna Galvão  
Arghavan Alisoltani  
(U. California)

Robert Wilkinson  
Darren Martin  
Nicola Mulder  
Wendy Burgers  
Ntobeko Ntusi  
Rageema Joseph  
Sean Wasserman  
Linda Boloko



CAPE TOWN HYTN  
IMMUNOLOGY LABORATORY



science & innovation  
Department  
Science and Innovation  
REPUBLIC OF SOUTH AFRICA

# University of the Free State



## UFS

Dominique Goedhals  
Armand Bester  
Martin Myaga  
Peter Mwangi  
Emmanuel Ogunbayo  
Milton Mogotsi  
Makgotso Maotoana  
Lutfiyya Mohamed



## NHLS Division of Virology

Sabeegah Vawda  
Felicity Burt  
Thokozani Mkhize  
Diagnostic laboratory staff



# National Institute for Communicable Diseases



## Centre for Respiratory Diseases & Meningitis

Jinal Bhiman  
Anne von Gottberg  
Thabo Mohale  
Daniel Amoako  
Josie Everatt  
Boitshoko Mahlangu  
Noxolo Ntuli  
Anele Mnguni  
Amelia Buys  
Cardia Fourie  
Noluthando Duma  
Linda de Gouveia  
Jackie Kleynhans  
Nicole Wolter  
Sibongile Walaza  
Mignon du Plessis  
Stefano Tempia  
Mvuyo Makhasi  
Cheryl Cohen

## Centre for HIV and STIs

Cathrine Scheepers  
Constantinos Kurt Wibmer  
Thandeka Moyo  
Tandile Hermanus  
Frances Ayres  
Zanele Molaudzi  
Bronwen Lambson  
Tandile Hermanus  
Mashudu Madzivhandila  
Prudence Kgagudi  
Brent Oosthuysen  
Penny Moore  
Lynn Morris

## NICD Groups

NICD COVID-19 response team  
NICD SARS-CoV-2 Sequencing Group

## Sequencing Core Facility

Zamantungwa Khumalo  
Annie Chan  
Morne du Plessis  
Stanford Kwenda  
Phillip Senzo Mtshali  
Mushal Allam  
Florah Mnyameni  
Arshad Ismail



# Zoonotic arbo and respiratory virus program

## Centre for Viral Zoonoses Department Medical Virology/ NHLS Tshwane Academic division University of Pretoria



## ZARV research program/UP

Marietjie Venter (Head: ZARV)  
Adriano Mendes (Postdoc)  
Amy Strydom (Postdoc)  
Michaela Davis (MSc, intern medical scientist)



## NHLS Tshwane

Prof Simnikiwe Mayaphi (HOD)

## Funders:

GIZ/BMBF: African Network for Improved diagnostics and epidemiology of common and emerging infectious agents (ANDEMIA)  
G7 Global Health fund, Robert Koch Institute, Dr Fabian Leendertz







# Additional support and collaborators



## NHLS

Koeleka Mlisana  
Zinhle Makatini  
Eugene Elliot  
Florette K. Treurnicht  
Kathleen Subramoney  
Oluwakemi Laguda-Akingba  
Shareef Abrahams  
Greta Hoyland  
Gloria Selabe  
Elias Bereda  
Jeannette Wadula

## Hyrax Biosciences

Simon Travers

## Cape Town HVTN Laboratory

Erica Anderson-Nissen  
Anneta Naidoo

## Ndlovu Research

Hugo Tempelman  
CJ Umunnakwe

## Lancet

Allison J. Glass

## Ampath

Terry Marshall  
Cindy van Deventer  
Eddie Silberbauer

## Pathcare Vermaak

Andries Dreyer  
Howard Newman  
Riaan Writes  
Marianne Wolfaardt  
Warren Lowman

## Bridge-the-Gap

Raymond Rott

## Cytespace Africa Laboratories

Christa Viljoen

## ARC-OVI

Lia Rotherham

## CAPRISA

Salim Abdool Karim  
Nigel Garret

## UKZN - Big Data

Francesco Pettruccione  
Ilya Sinayskiy

## University of Oxford

José Lourenço

## FioCruz, Brazil

Vagner Fonseca  
Marta Giovanetti  
Luiz Carlos Junior Alcantara

## Africa CDC

John Nkengasong  
Sofonias Tessema

## Netcare:

Richard Friedland  
Craig Murphy  
Caroline Maslo  
Liza Sitharam

## DSI

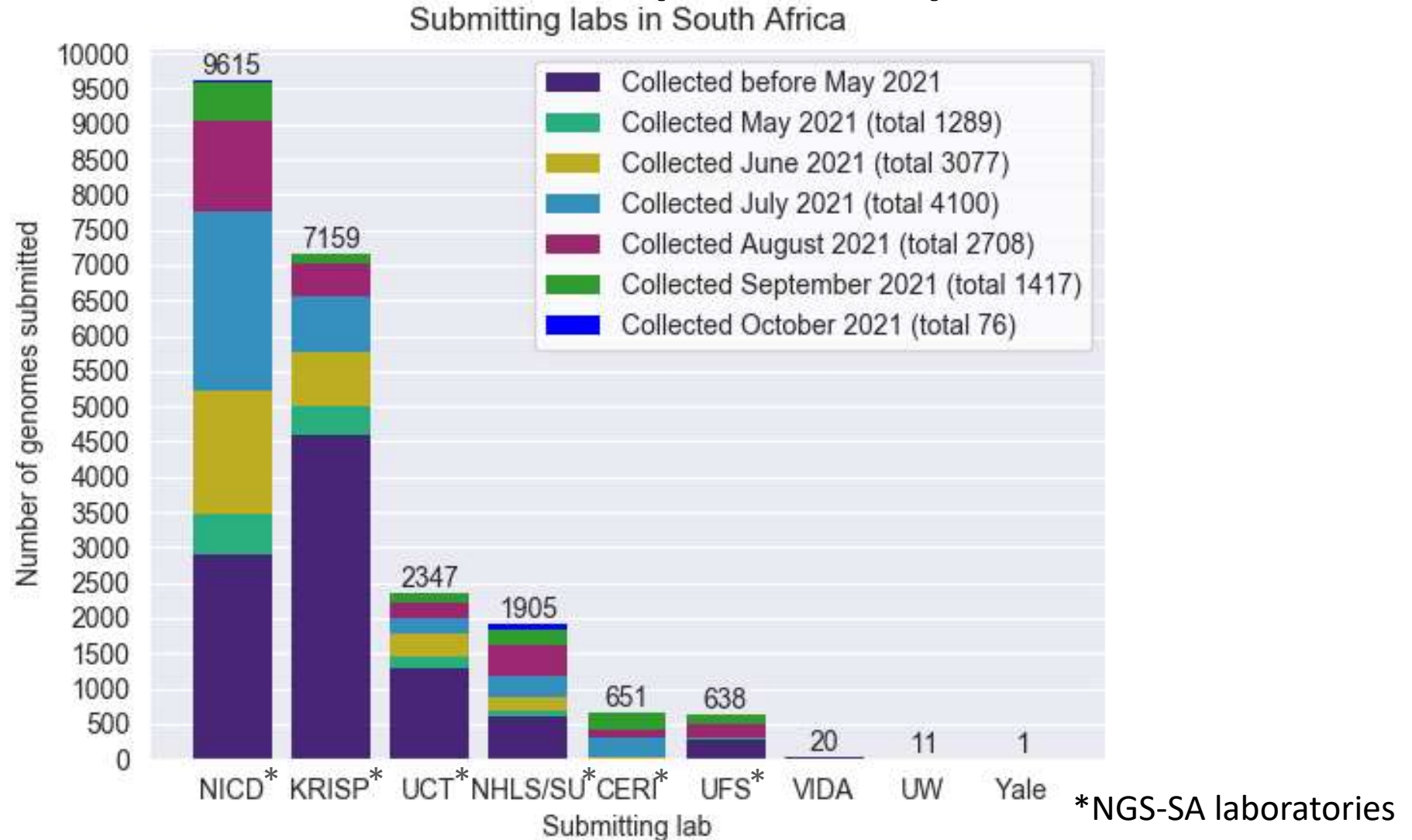
Glaudina Loots

## SA MRC

Glenda Gray



# South African genomes submitted per sequencing lab, 2020 and 2021 (N=22 347)



Multiple labs from NGS-SA are contributing to the sequencing effort. Sequencing efforts increased with the third wave.

# Variants of Concern (VOC)

WHO label	Pango lineages <sup>+</sup>	GISAID clade	Nextstrain clade	Additional amino acid changes monitored*	Earliest documented samples	Date of designation
Alpha	B.1.1.7 <sup>#</sup>	GRY	20I (V1)	+S:484K +S:452R	United Kingdom, Sep-2020	18-Dec-2020
Beta	B.1.351	GH/501Y.V2	20H (V2)	+S:L18F	South Africa, May-2020	18-Dec-2020
Gamma	P.1	GR/501Y.V3	20J (V3)	+S:681H	Brazil, Nov-2020	11-Jan-2021
Delta	B.1.617.2 <sup>§</sup>	G/478K.V1	21A	+S:417N	India, Oct-2020	VOI: 4-Apr-2021 VOC: 11-May-2021

<https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/> accessed 5 November 2021

\*Notable spike (S) amino acid changes under monitoring, which are currently reported in a minority of sequenced samples

<sup>+</sup>Includes all descendant lineages.

<sup>#</sup>Includes all Q.\* lineages in the PANGO nomenclature system.

<sup>§</sup>Includes all AY.\* lineages in the PANGO nomenclature system.



# Currently designated Variants of Interest (VOI)

WHO label	Pango* lineages	GISAID clade	Nextstrain clade	Earliest documented samples	Date of designation
Lambda	C.37	GR/452Q.V1	21G	Peru, Dec-2020	14-Jun-2021
Mu	B.1.631	GH	21H	Colombia, Jan-2021	30-Aug-2021

<https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/> accessed 5 November 2021

\* Includes all descendant lineages.

# Submission of routine specimens for sequencing

- representative of multiple geographic regions (provinces/districts/health facilities) from individuals of
  - all ages
  - over as many time periods during the SARS-CoV-2 epidemic in South Africa
- requested that testing laboratories in both the private and public sectors, submit respiratory samples to their closest NGS-SA sequencing laboratory on a routine basis (ideally every week) as follows, depending on the capacity of the testing laboratory:
  - All positives samples should be sent every week (NGS-SA laboratory will perform random sampling as described below) **OR**
  - A weekly selection of approximately 10%-20% of randomly selected positive samples should be sent every week. Number of selected samples will depend on the size of laboratory and how many other laboratories are drained by the submitting laboratory.

# Submission of special interest specimens for sequencing

In addition to routine samples mentioned above, please send specimens separately to above and clearly marked if:

- Suspected vaccine breakthrough ( $\geq 14$  days after vaccine), especially if hospitalised and clinically severe
- Suspected re-infection ( $\geq 90$  days after previous episode), especially if hospitalised and clinically severe
- Prolonged shedding with high SARS-CoV-2 viral loads (i.e. Ct values less than 30 for more than 1 month post-primary diagnosis) in immunocompromised individuals
- Possible animal-to-human transmission
- Suspected cases of importation from another country, especially countries known to harbour SARS-CoV-2 variants of concern or countries with little available information
- Clusters of “unusual” cases (e.g., in terms of disease presentation, patient groups affected, etc.)