

## SARS-CoV-2 Sequencing Update 4 March 2022



Supported by the DSI and the SA MRC

Msomi N, Mlisana K, et al. Lancet Microbe 2020

The genomic data presented here are based on South African SARS-CoV-2 sequence data downloaded from GISAID ([www.gisaid.org](https://www.gisaid.org)) on 4 March 2022 at 14h16



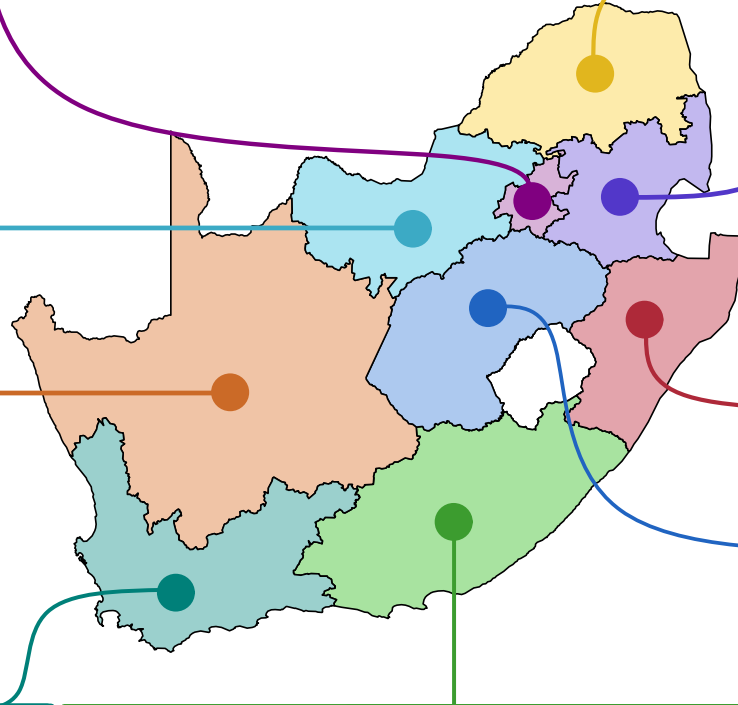
Data license: <https://www.gisaid.org/registration/terms-of-use/>

Elbe, S., and Buckland-Merrett, G. (2017) Data, disease and diplomacy: GISAID's innovative contribution to global health. *Global Challenges*, 1:33-46. DOI: 10.1002/gch2.1018 PMID: 31565258

Shu, Y., McCauley, J. (2017) GISAID: Global initiative on sharing all influenza data – from vision to reality. *EuroSurveillance*, 22(13) DOI: 10.2807/1560-7917.ES.2017.22.13.30494 PMID: PMC5388101

# SARS-CoV-2

## GENOMIC SURVEILLANCE epiweeks 39 (2021) - 8 (2022)



### Gauteng ↓ PTP: 8.6%

Genomes Cases  
2 396 (26.2%) 268 481 (35.6%)

Genomes deposited in the last week  
77 8 116 1

### North West ↓ PTP: 10.5%

Genomes Cases  
520 (5.7%) 39 283 (5.2%)

Genomes deposited in the last week  
78 6 19 1 1

### Northern Cape ↓ PTP: 9.0%

Genomes Cases  
548 (6.0%) 17 748 (2.4%)

Genomes deposited in the last week  
34 3 17 4 5

### Western Cape ↓ PTP: 12.6%

Genomes Cases  
1 741 (19.0%) 127 748 (16.9%)

Genomes deposited in the last week  
57 10 80 3 1 1 1 1 15 107 1 1 6

### Eastern Cape ↓ PTP: 5.8%

Genomes Cases  
821 (9.0%) 53 182 (7.0%)

Genomes deposited in the last week  
30 2 8 2 14 57 3

### Limpopo ↓ PTP: 8.4%

Genomes Cases  
691 (7.6%) 31 591 (4.2%)

Genomes deposited in the last week  
102 13 33 2

### Mpumalanga ↓ PTP: 14.0%

Genomes Cases  
795 (8.7%) 39 462 (5.2%)

Genomes deposited in the last week  
36 3 51 1

### KwaZulu-Natal ↑ PTP: 7.8%

Genomes Cases  
1 208 (13.2%) 139 016 (18.4%)

Genomes deposited in the last week  
1

### Free State ↓ PTP: 8.2%

Genomes Cases  
423 (4.6%) 38 448 (5.1%)

Genomes deposited in the last week  
11 1 27

Omicron (BA.1) Beta (20H, V2) 20A 20C  
 Omicron (BA.1.1) Alpha (20I, V1) 20B 20D  
 Omicron (BA.2) Delta (21A) C.1.2 Unassigned  
 Omicron (BA.3) Delta (21I) Delta (21J) Cases

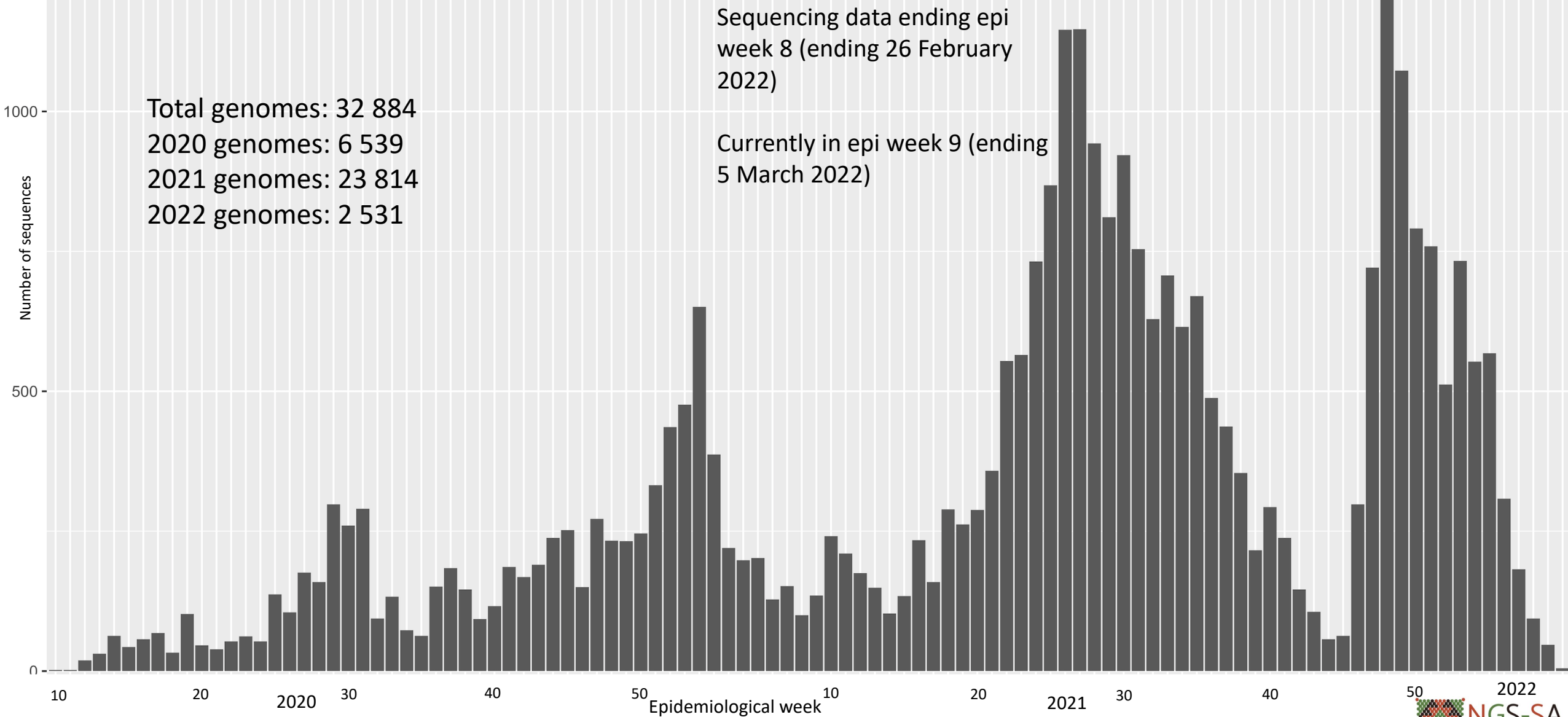
**1 072 genomes deposited in the past week**

Bar graphs represent genomes sequenced per epiweek, with lines representing cases by collection date (weeks 39 [2021] – 8 [2022])

Genomes and cases presented as provincial total (percentage of national total) for epiweeks 39 (2021) – 8 (2022)

PTP: percentage testing positive in week 8 (20 Feb 2022 – 26 Feb 2022); the arrow indicates direction of change since the previous week (13 Feb 2022 – 19 Feb 2022)

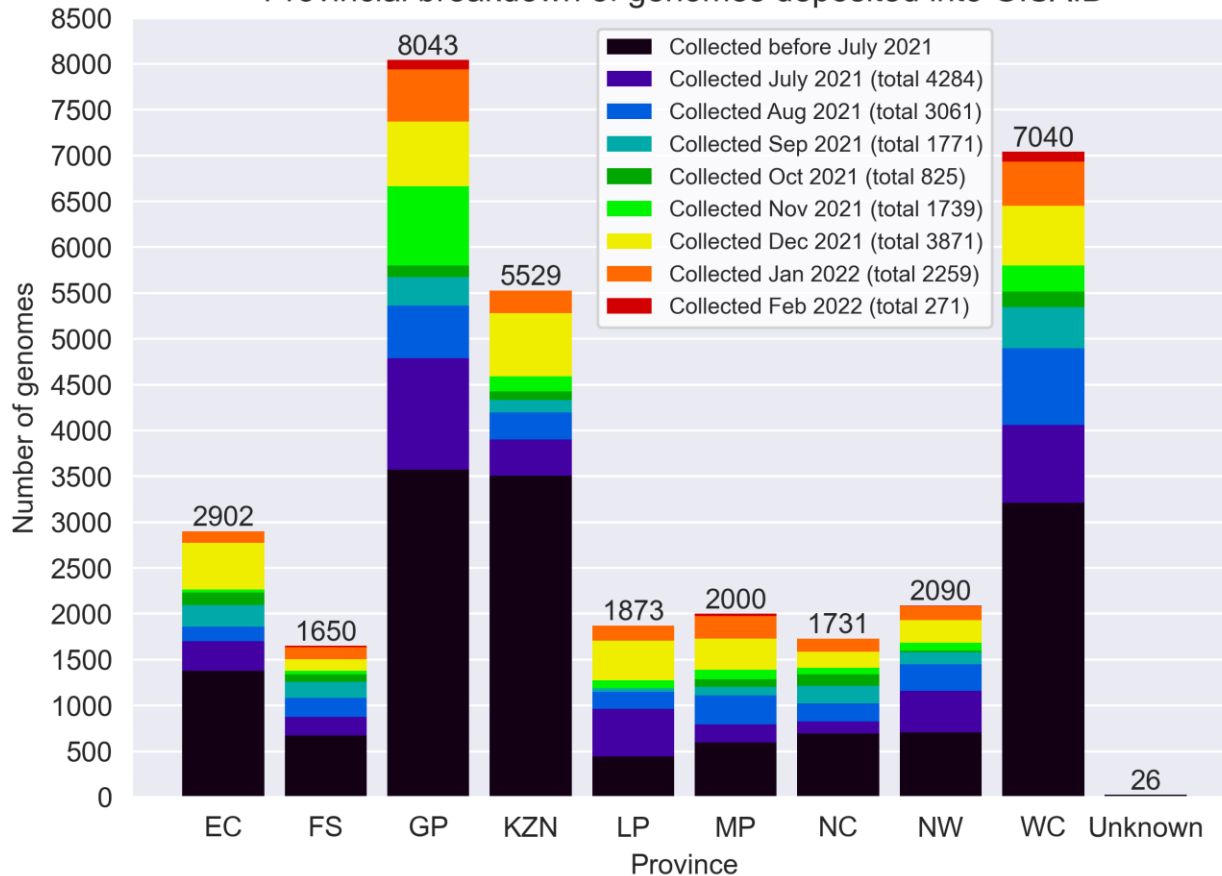
# Number of South African genomes deposited on GISAID, by specimen collection week, 2020 – 2022 (N=32 884\*)



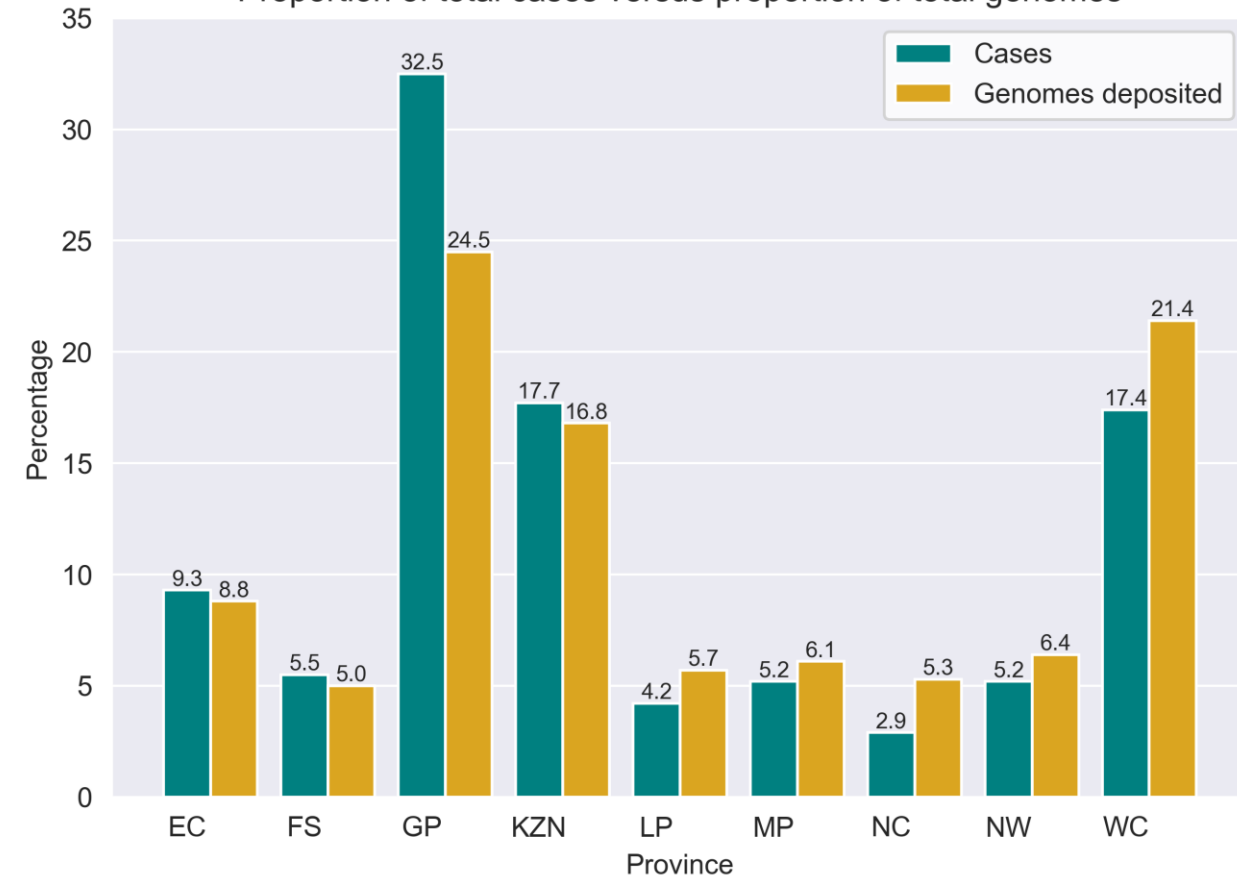
\*This represents the cleaned, de-duplicated dataset of unique **National and Pneumonia Surveillance** sequences. This dataset will be used for all further figures.

# GISAID genomes vs total cases, 2020 – 2022 (N=32 884)

Provincial breakdown of genomes deposited into GISAID

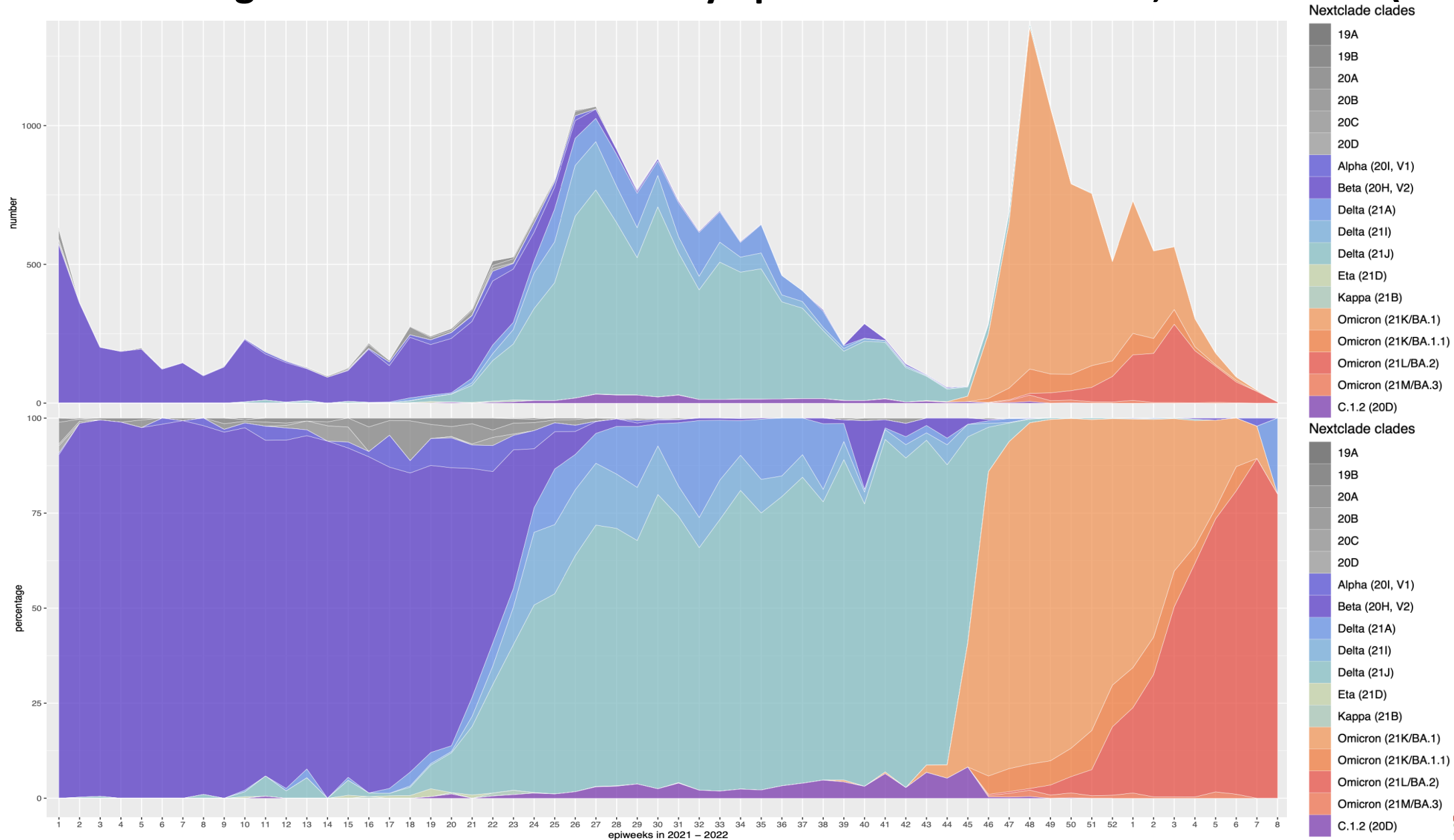


Proportion of total cases versus proportion of total genomes



All provinces, apart from GP, LP, NC and WC, have comparable percentages of overall cases and overall sequenced genomes.

# Percentage and number of clades by epiweek in South Africa, 2021 - 2022 (N=26 345)



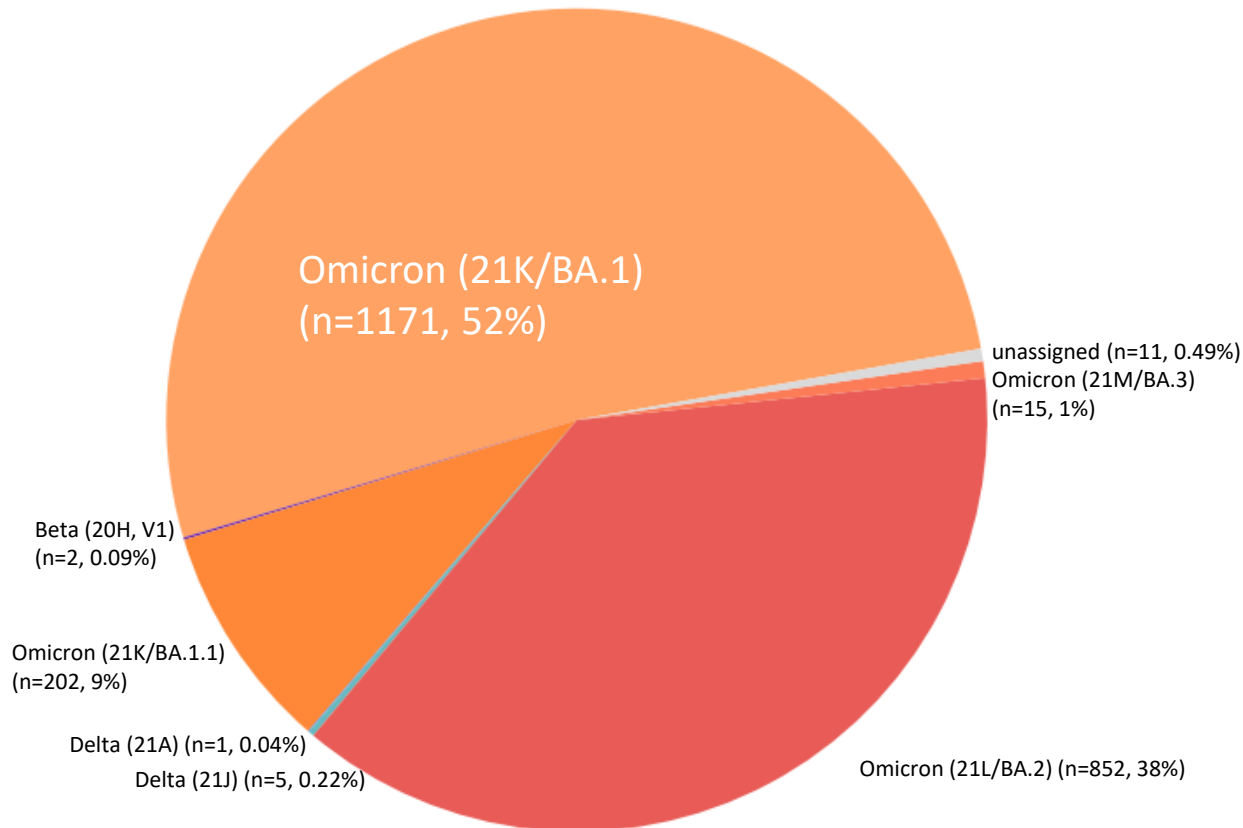
Sequencing data  
ending epi week 8  
(ending 26  
February 2022)

Currently in epi  
week 9 (ending 5  
March 2022)

Delta dominated in South Africa until October at >80%. Omicron has dominated from November onwards.

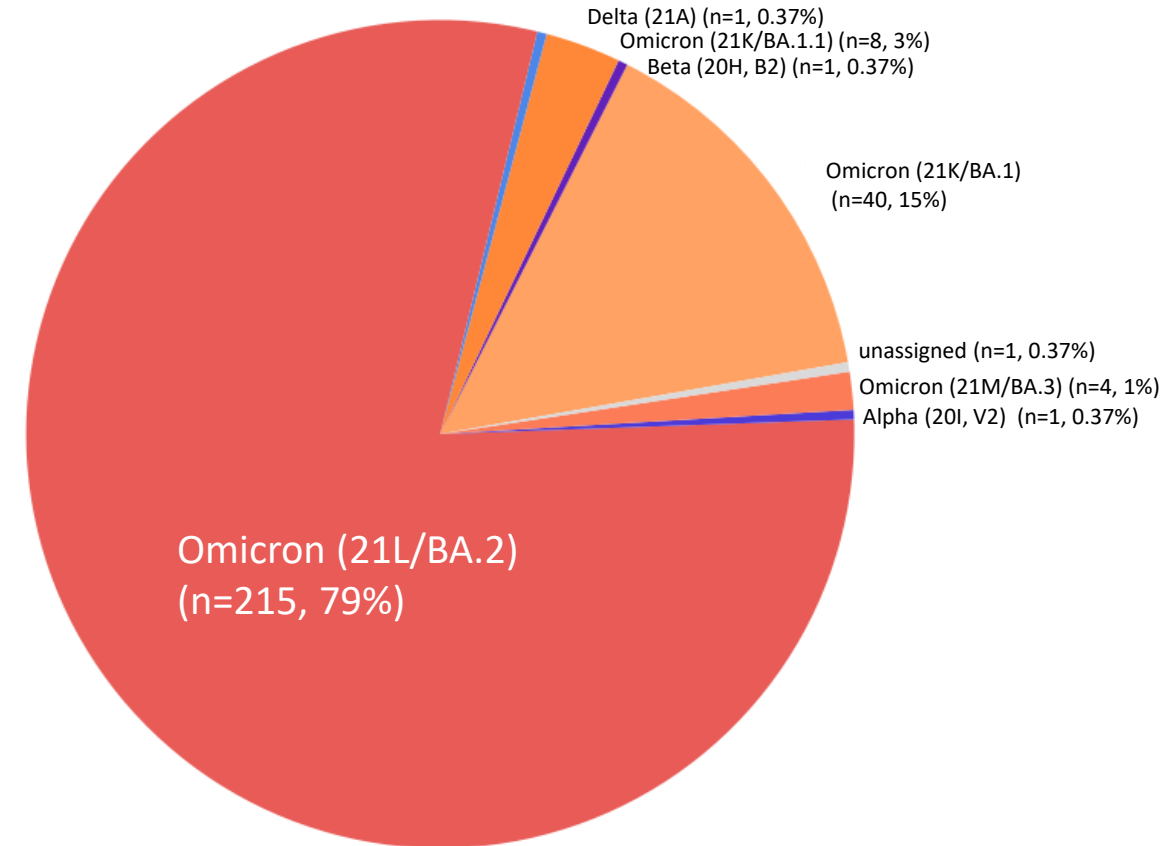
# Prevalence of Variants of Concern (VOC) and Variants of Interest (VOI) in Jan 2021 – Feb 2022

January (N=2259)



Total Omicron in Jan: 2240 (99.2%)

February (N=271)



Total Omicron in Feb: 267 (98.5%)

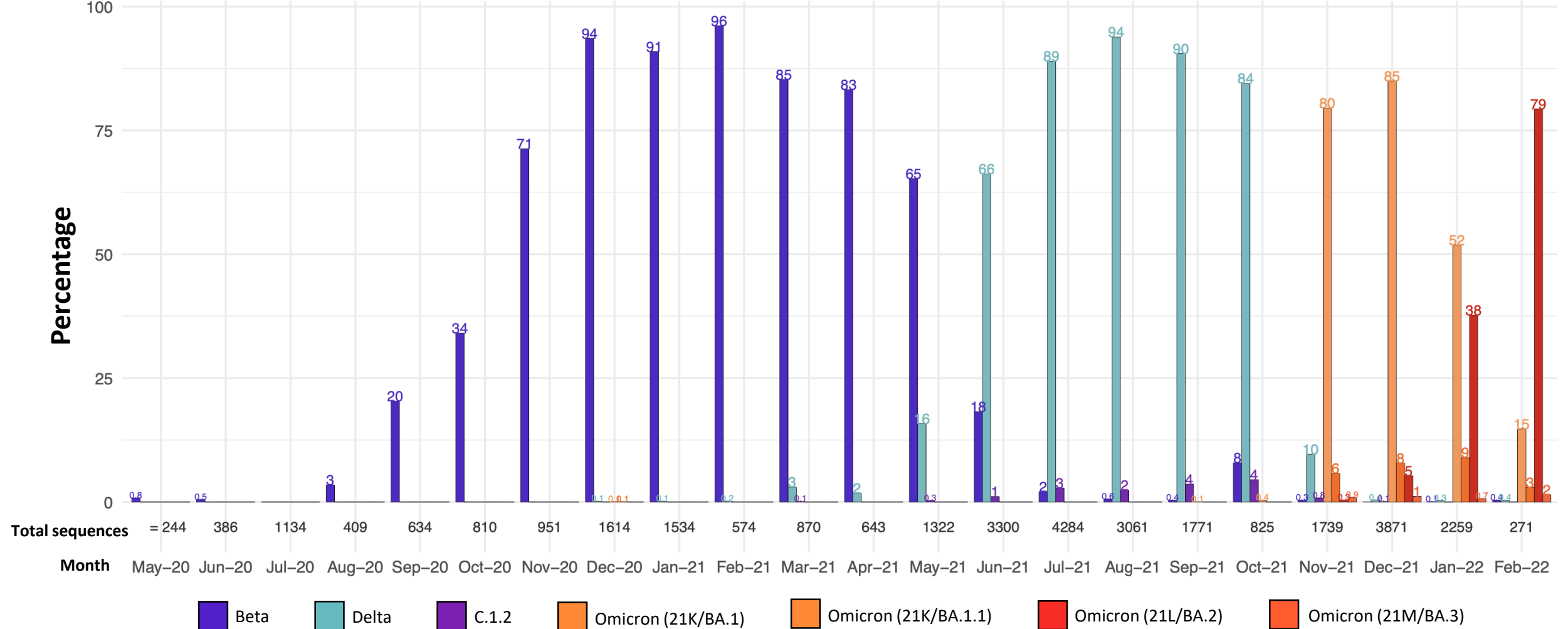


Omicron dominated in January (99%, 2240/2259), and continues to dominate in February (98%, 267/271) with sub-lineage BA.2 increasing in prevalence. Very low levels (<1% each) of Alpha, Beta and Delta have also been detected.



# Detection Rates: Beta, Delta, C.1.2 and Omicron

Detection rates of variants being monitored in South Africa\*

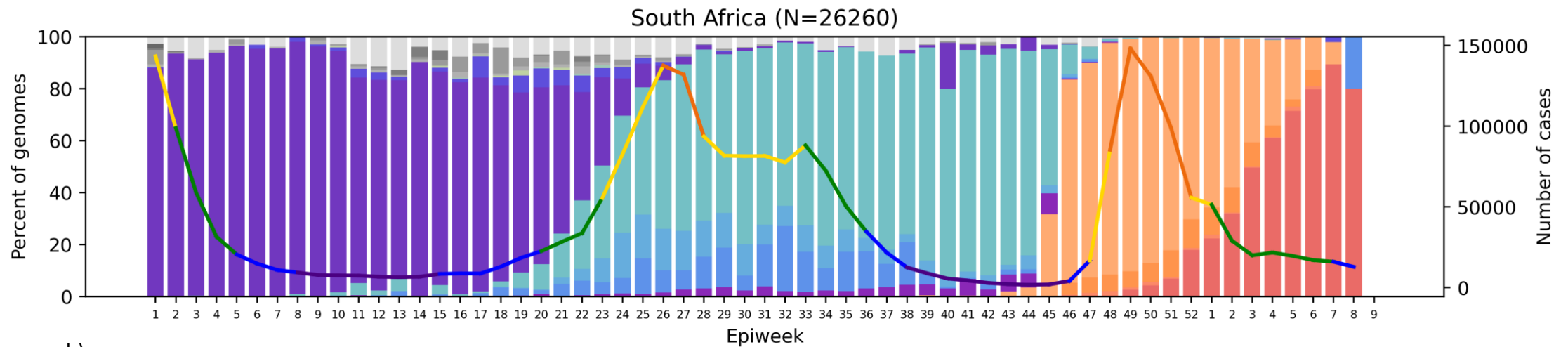
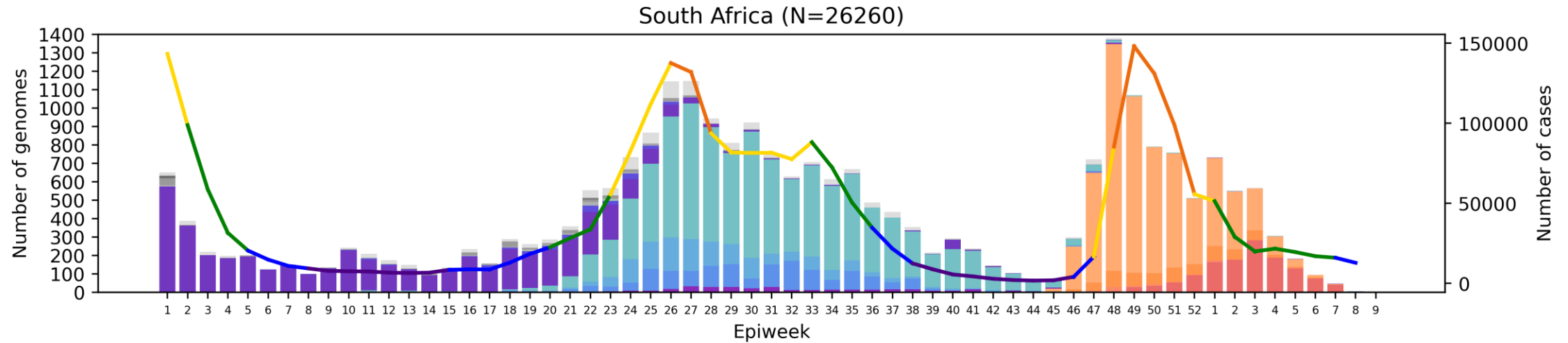


\*Bars represent percentage prevalence of variant for the month; total sequences collected for the month are given below

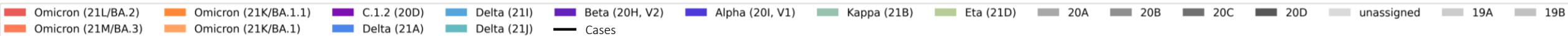
**Omicron has been dominant since November (>80% in November, >99% in December and January). BA.2 increased in frequency in January, making up 38% of genomes. BA.2 dominates in February (79%).**  
**BA.3 continues to be present at low levels.**



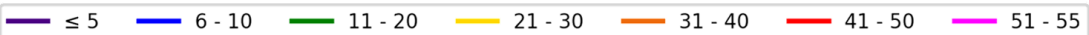
# South Africa, 2021-2022, n = 26260\*



Clade key (bar graph)

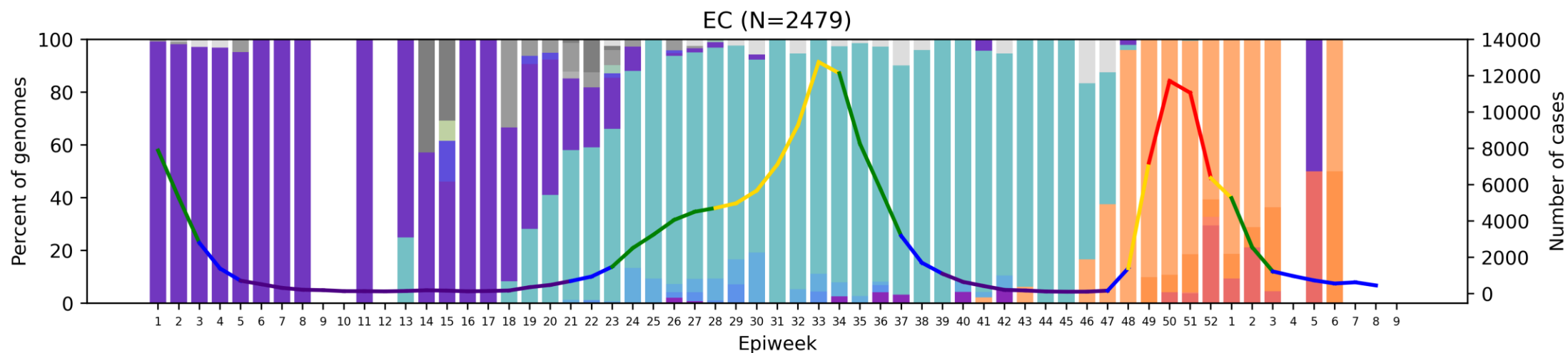
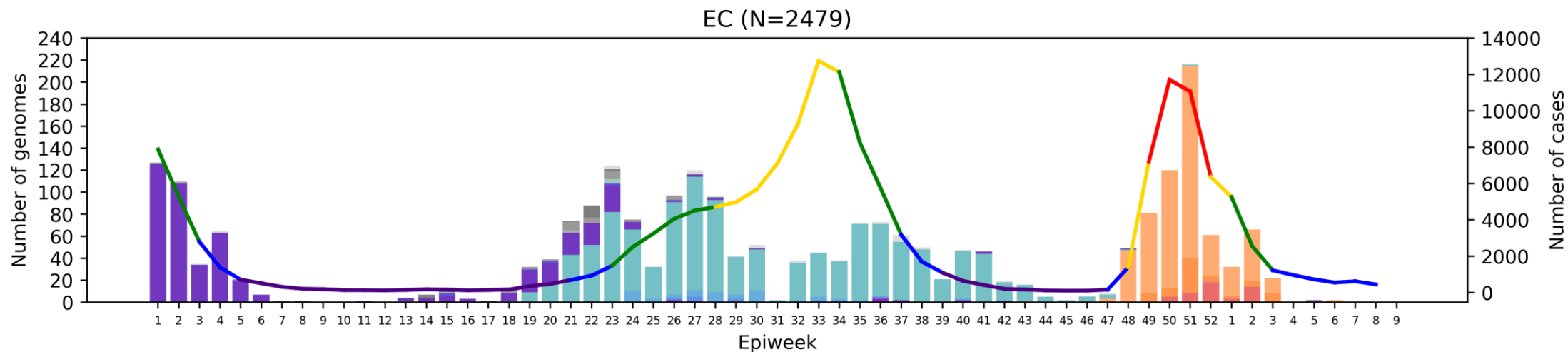


Weekly percentage testing positive key (line graph)

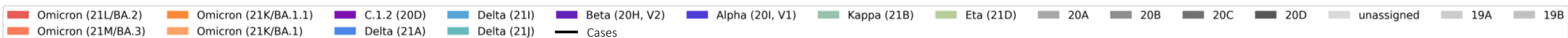


\*Excludes sequences missing collection dates, as well as those collected January 1<sup>st</sup> and 2<sup>nd</sup> 2021 as they are part of epiweek 53 of 2020.

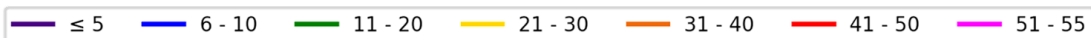
# Eastern Cape Province, 2021-2022, n = 2479



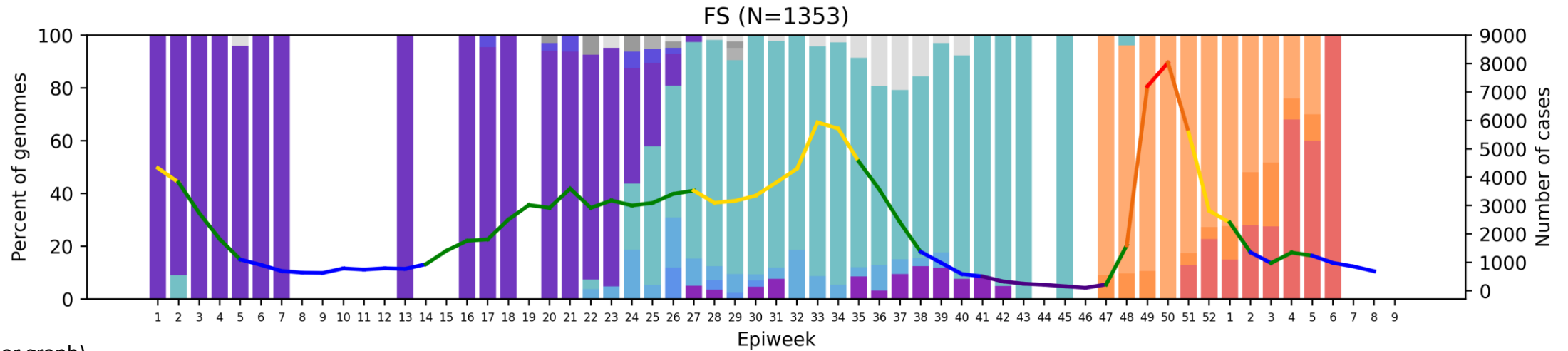
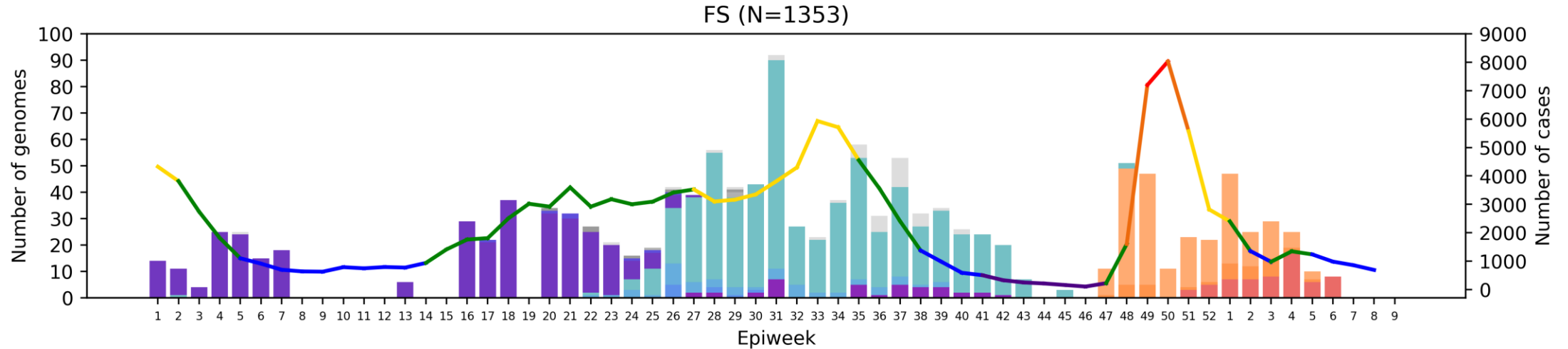
Clade key (bar graph)



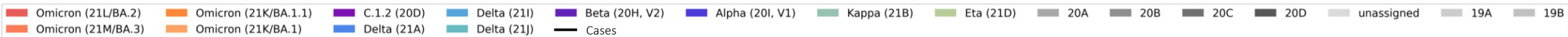
Weekly percentage testing positive key (line graph)



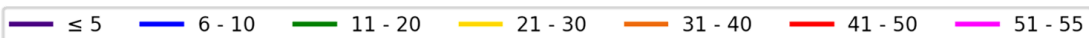
# Free State Province, 2021-2022, n = 1353



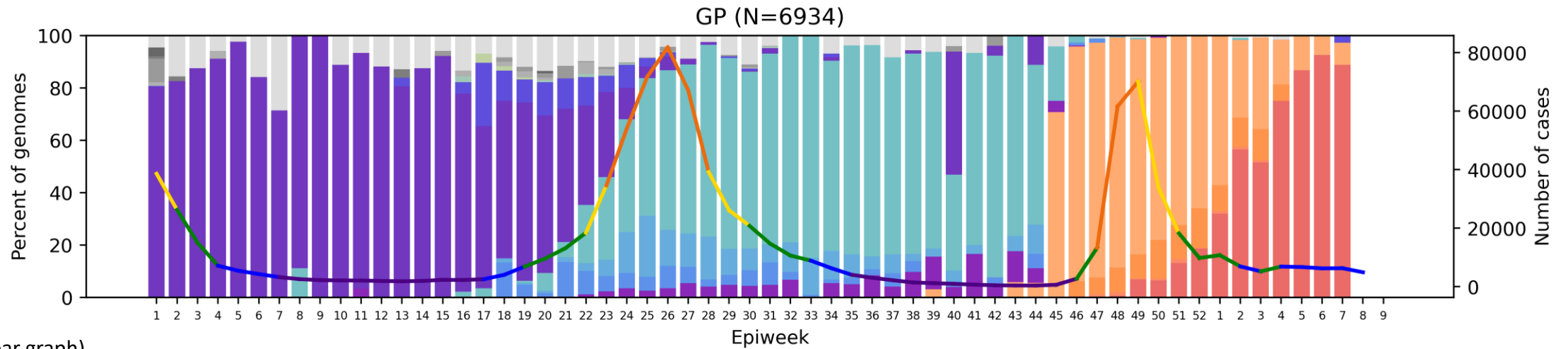
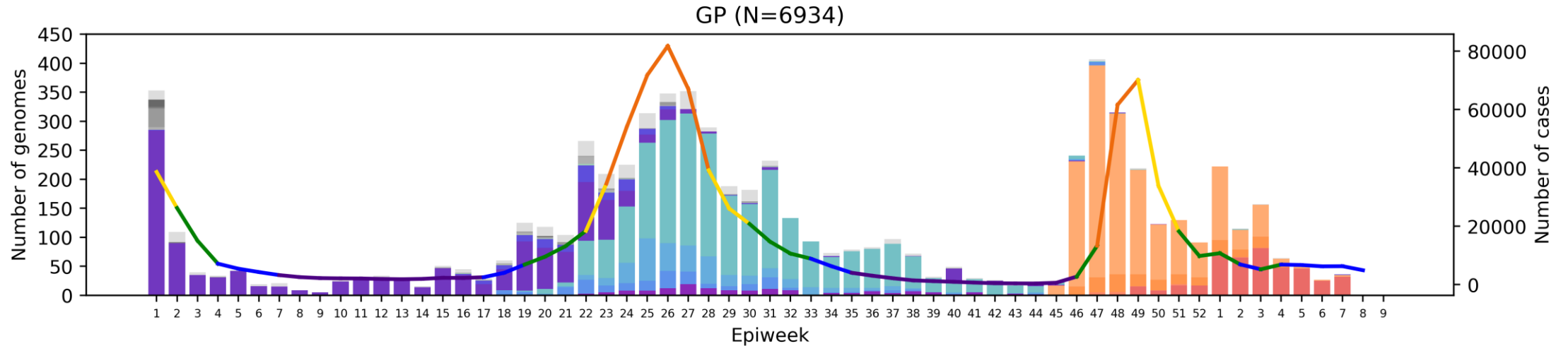
Clade key (bar graph)



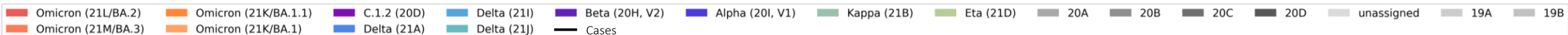
Weekly percentage testing positive key (line graph)



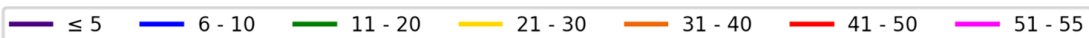
# Gauteng Province, 2021-2022, n = 6934



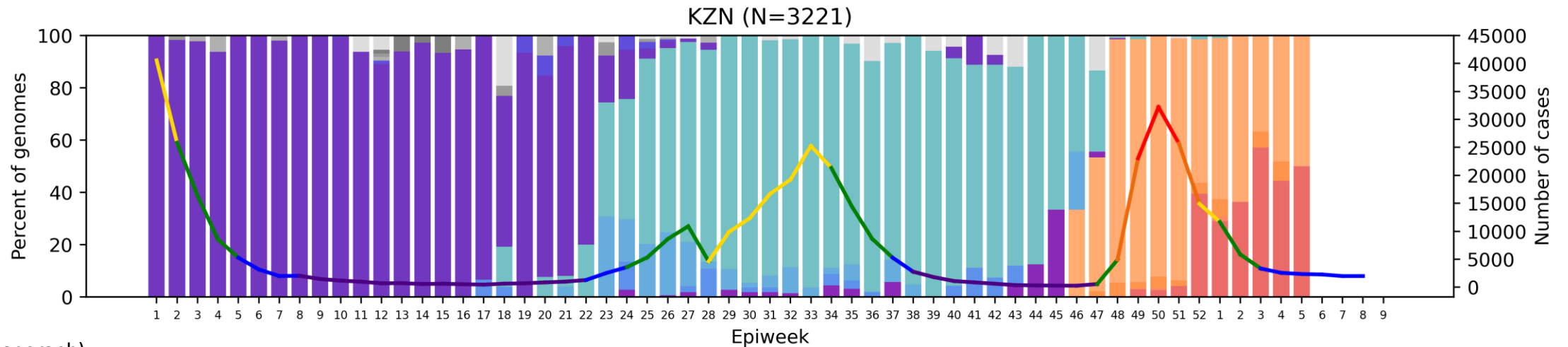
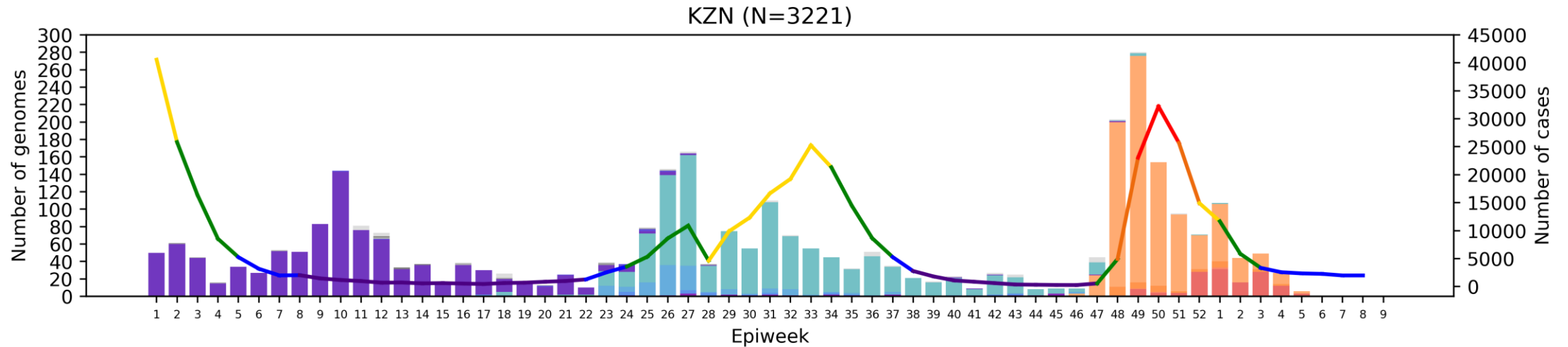
Clade key (bar graph)



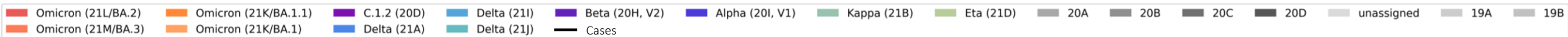
Weekly percentage testing positive key (line graph)



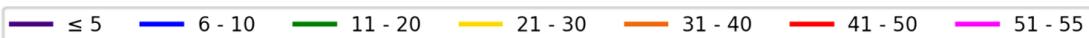
# KwaZulu-Natal Province, 2021-2022, n = 3221



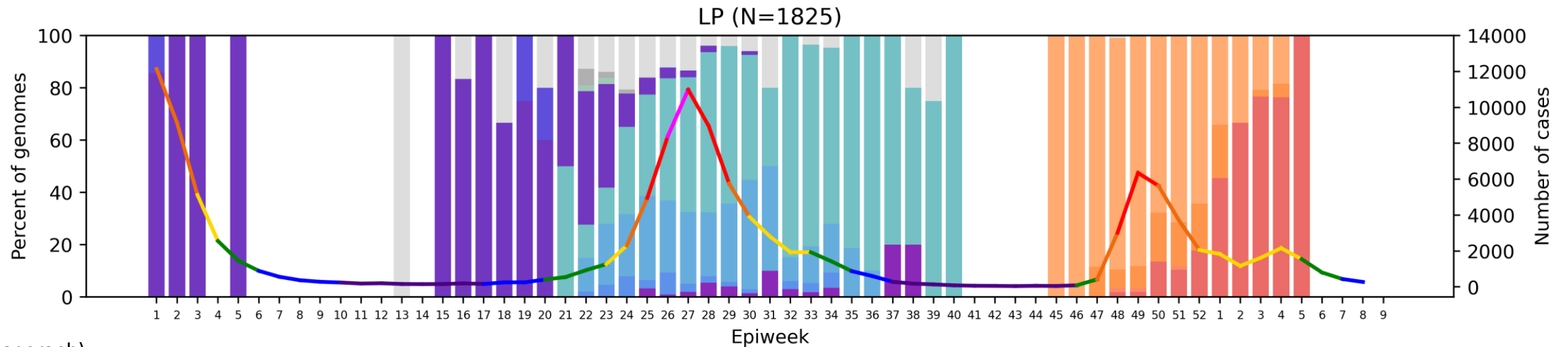
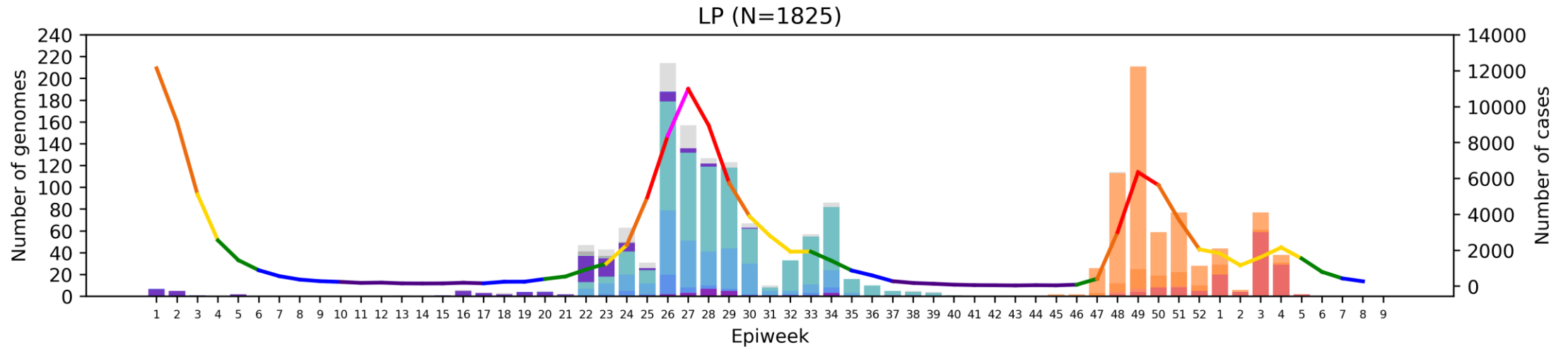
Clade key (bar graph)



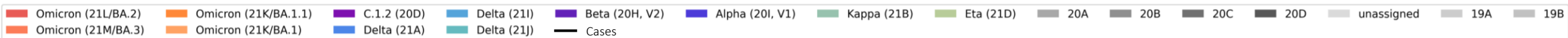
Weekly percentage testing positive key (line graph)



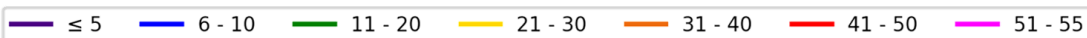
# Limpopo Province, 2021-2022, n = 1825



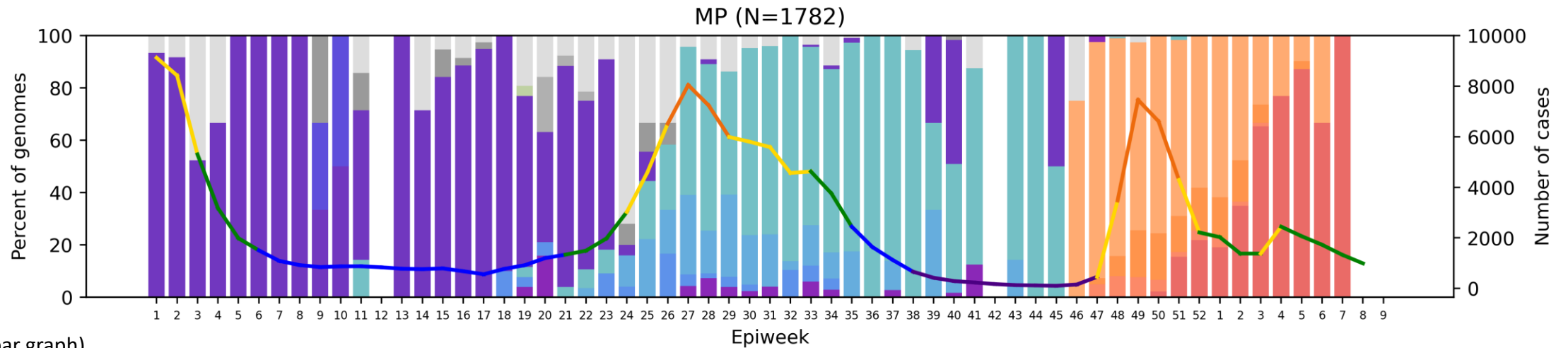
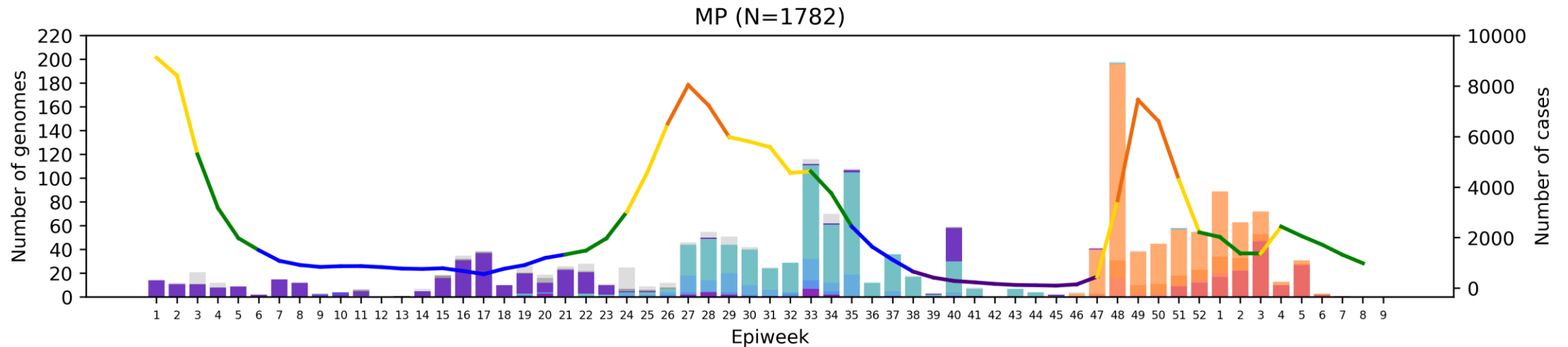
Clade key (bar graph)



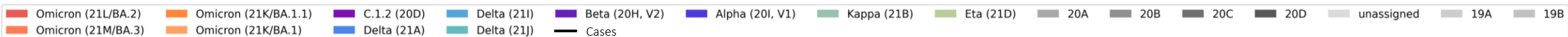
Weekly percentage testing positive key (line graph)



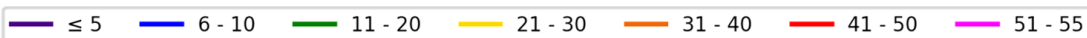
# Mpumalanga Province, 2021-2022, n = 1782



Clade key (bar graph)

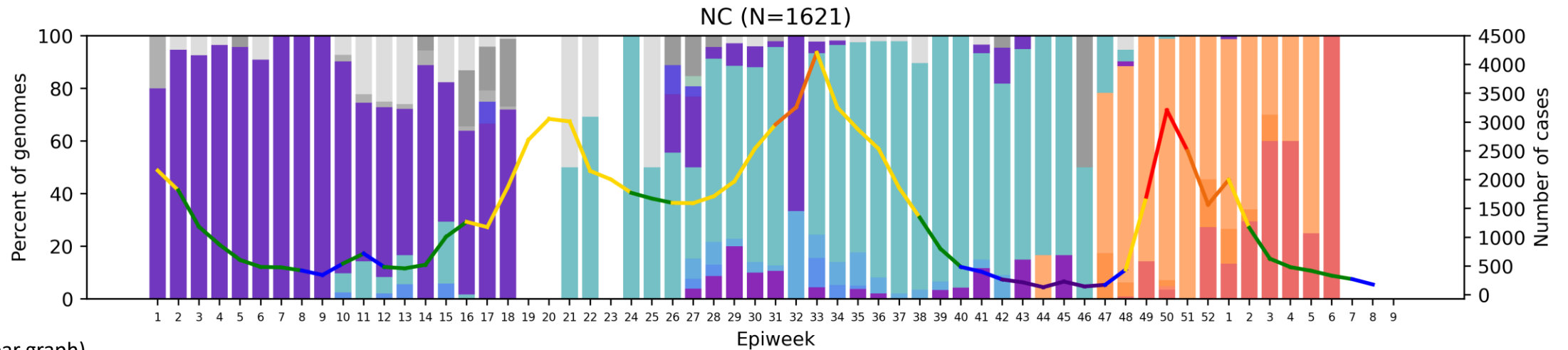
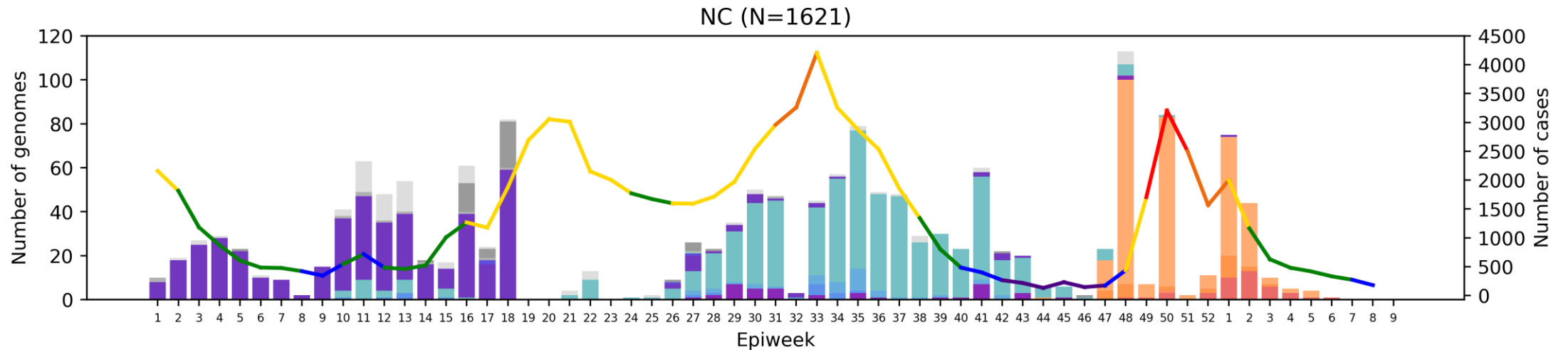


Weekly percentage testing positive key (line graph)

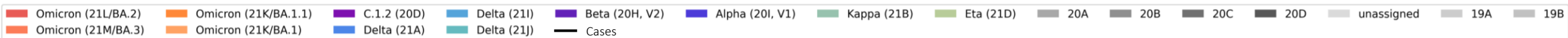




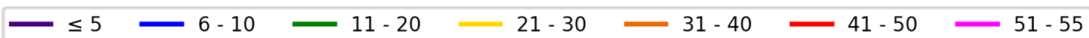
# Northern Cape Province, 2021-2022, n = 1621



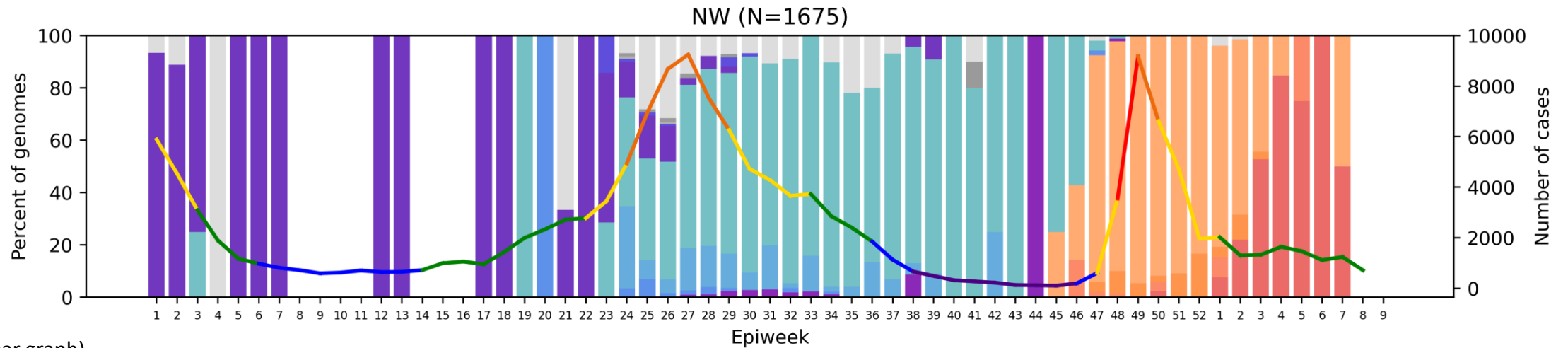
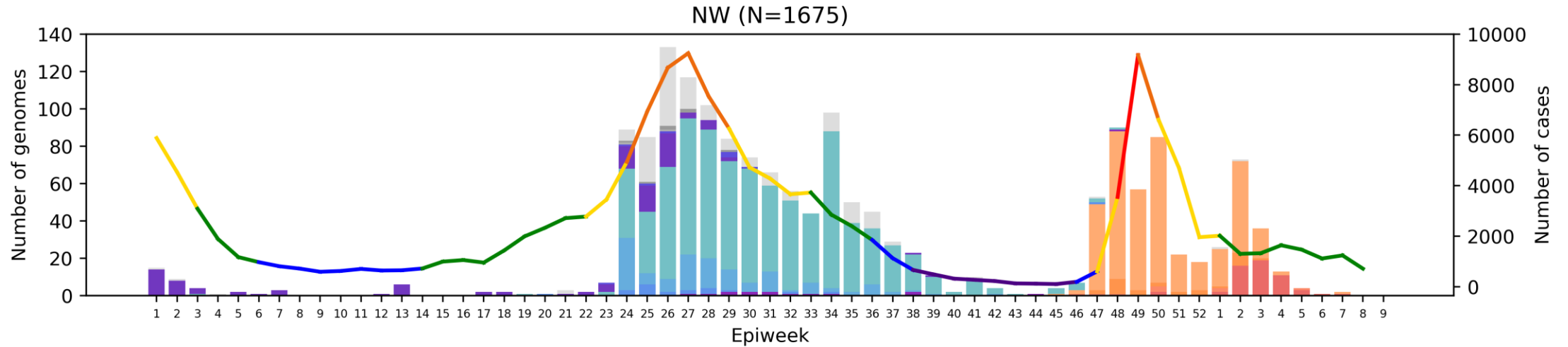
Clade key (bar graph)



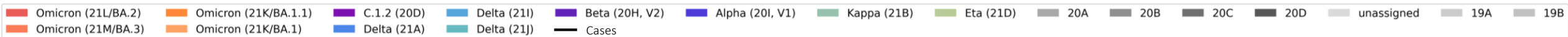
Weekly percentage testing positive key (line graph)



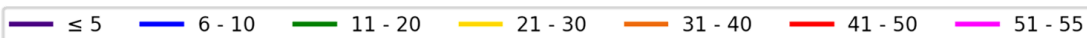
# North West Province, 2021, n = 1675



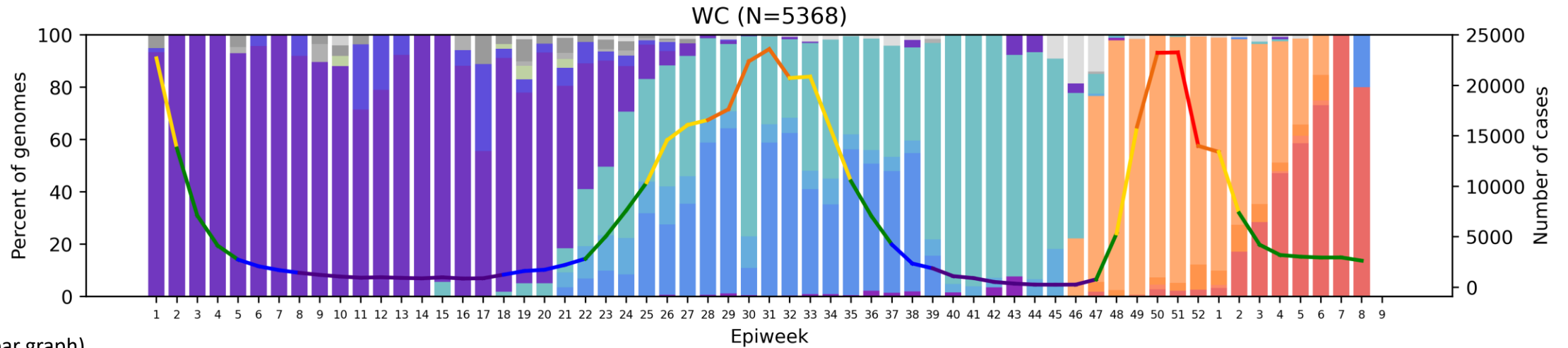
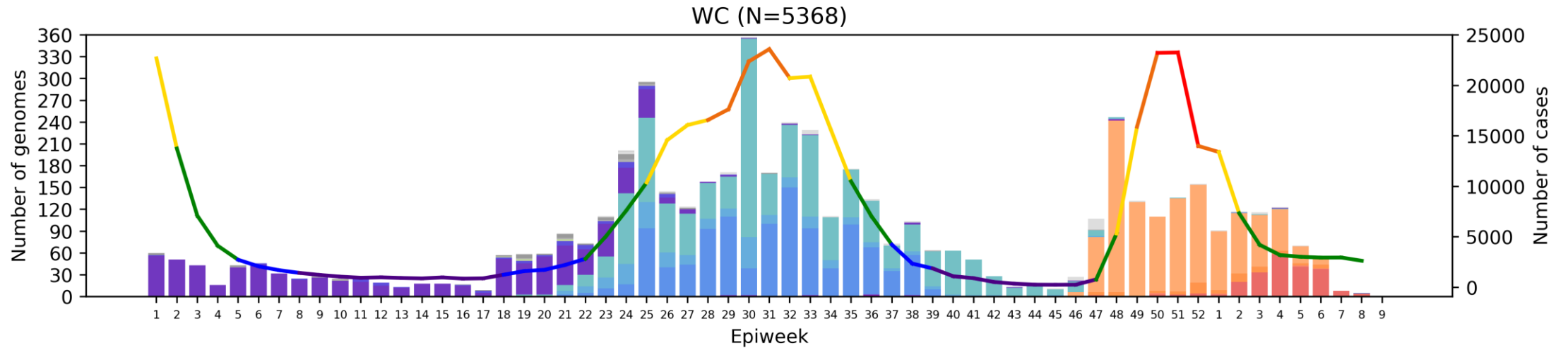
Clade key (bar graph)



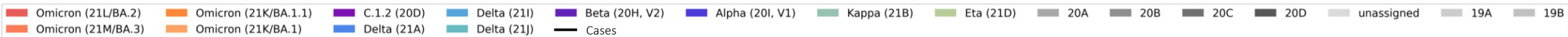
Weekly percentage testing positive key (line graph)



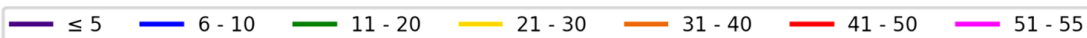
# Western Cape Province, 2021-2022, n = 5368



Clade key (bar graph)



Weekly percentage testing positive key (line graph)



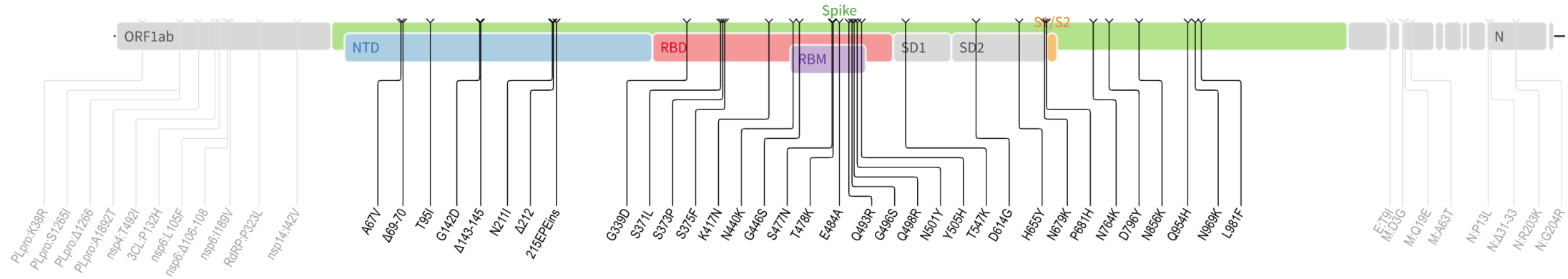
# Summary

- **Variant of Concern Omicron**

- Detected in at least 153 countries and dominating globally
- Split into different lineages based on different mutational profiles. Predominant lineages currently BA.1 (21K), BA.1.1 (21K, BA.1+spike R346K), BA.2 (21L), BA.3 (21M). More lineages are being assigned but defining mutations are not yet available.
- South Africa (detected in all provinces):
  - Dominated December, January and February sequencing data at >99% of genomes
  - While BA.1 was the predominant sub-lineage in December (85%) and January (52%), the proportion of BA.2 increased from 5% in December and 38% in January to 79% in February
  - BA.3 continues to be detected at low levels
- Low frequency of previously circulating variants such as Delta and Beta still detected in recent data

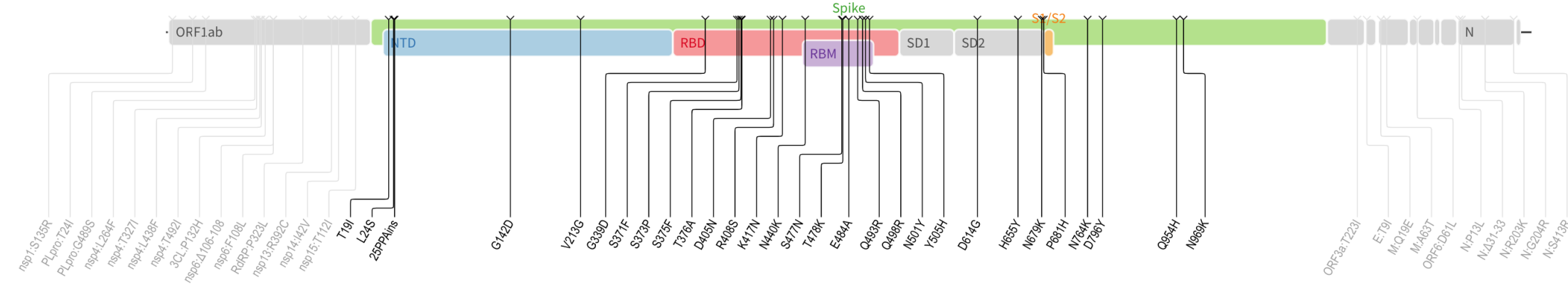
# Omicron sub-lineage spike mutation profiles

**BA.1  
21K**

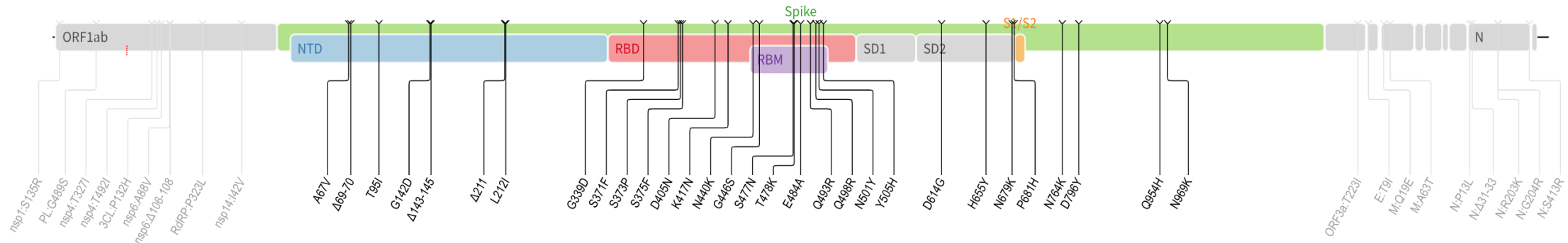


**BA.2  
21L**

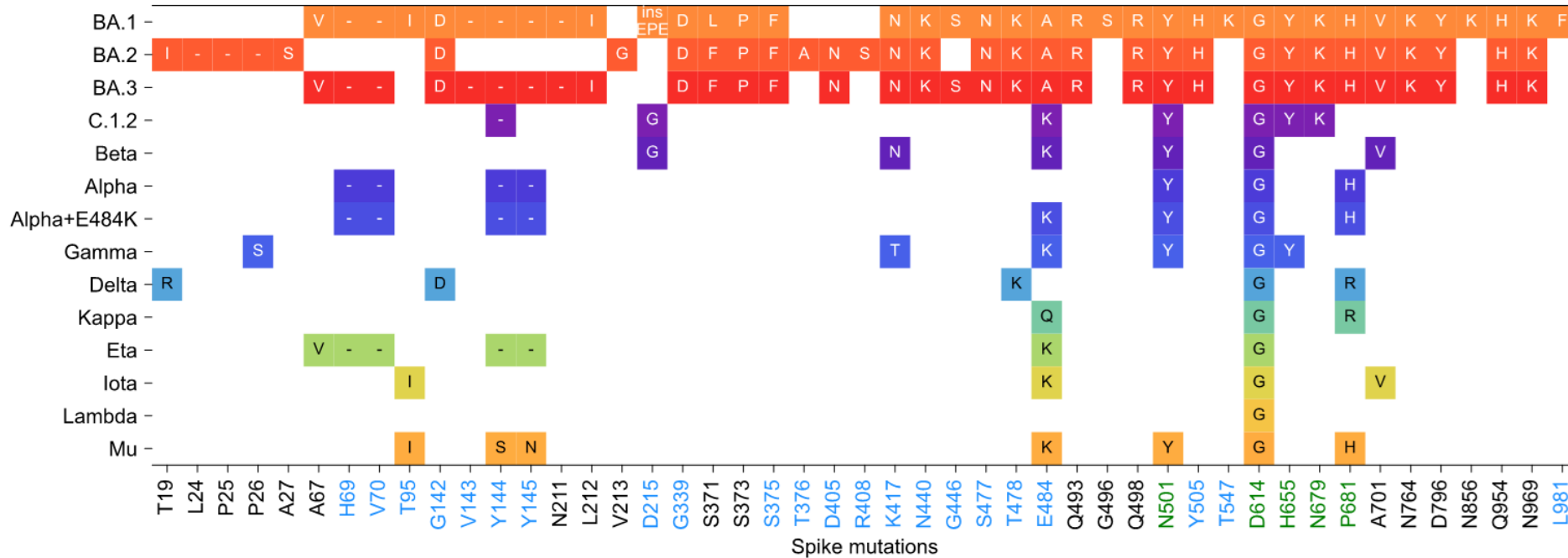
Lacks 69-70del  
Not detectable by  
S-Gene Target  
Failure



**BA.3  
21M**



# Omicron spike mutations compared to other VOC/VOIs



Only lineage-defining mutations are pictured here. Low prevalence mutations can be seen on the following slide.

## Mutation impact key

- Unknown or unconfirmed impact
- Known/predicted immune escape
- Enhanced infectivity

- Multiple changes within the two immunogenic regions in S1 (NTD and RBD)
  - including a three amino acid insertion
- Accumulation of mutations surrounding the furin cleavage site
  - Including combination of N679K and P681H
- Effect of most spike S2 subunit changes have not been defined, but may be linked to immune escape





Supported by the DSI and the SA MRC



UNIVERSITEIT VAN PRETORIA  
UNIVERSITY OF PRETORIA  
YUNIBESITHI YA PRETORIA



UNIVERSITEIT  
YUNIBESITHI  
STELLENBOSCH  
UNIVERSITY



UNIVERSITY OF CAPE TOWN  
IYUNIBESITHI YASEKAPA • UNIVERSITEIT VAN KAAPSTAD



science & innovation  
Department:  
Science and Innovation  
REPUBLIC OF SOUTH AFRICA



WITS  
UNIVERSITY



UNIVERSITY OF  
KWAZULU-NATAL  
INYUVESI  
YAKWAZULU-NATALI



EDCTP

This project (RIA2020EF-3030) is part of the EDCTP2 programme supported by the European Union





## University of Stellenbosch & NHLS Tygerberg Virology



Susan Engelbrecht  
Wolfgang Preiser  
Gert van Zyl  
Tongai Maponga  
Bronwyn Kleinhans  
Shannon Wilson  
Karabo Phadu  
Tania Stander  
Kamela Mahlakwane  
Mathilda Claassen  
Diagnostic laboratory staff

## UKZN-Inkosi Albert Luthuli Central Hospital



Dr Khanyi Msomi  
Dr Kerusha Govender  
Dr Pravi Moodley  
Dr Aabida Khan  
Dr Lili Gounder  
Dr Kerri Francois  
Dr Cherise Naicker  
Dr Joedene Chetty

Dr Neli Ngcaba  
Dr Tshepiso Mosito  
Mr Malcolm Ellapen  
Mr Kubendran Reddy  
The COVID-19 Bench team

## University of KwaZulu-Natal & Africa Health Research Institute



**KRISP at UKZN:**  
Tulio de Oliveira  
Richard Lessels  
Houriiyah Tegally  
Eduan Wilkinson  
Jennifer Giandhari  
Sureshnee Pillay  
Emmanuel James San



**AHRI**  
Alex Sigal  
Sandile Cele  
Willem Hanekom

## University of Cape Town, NHLS & Western Cape Government



**NHLS-UCT**  
Carolyn Williamson  
Nei-yuan Hsiao  
Diana Hardie  
Kruger Marais  
Stephen Korsman  
Ziyaad Valley-Omar

**WCG-UCT**  
Mary-Anne Davies  
Hannah Hussey  
Andrew Boule  
Masudah Paleker  
Theuns Jacobs  
Erna Morden

**NHLS Greenpoint**  
Annabel Enoch



### UCT, IDM and CIDRI-Africa

Deelan Doolabh  
Arash Iranzadeh  
Lynn Tyers  
Innocent Mudau  
Nokuzola Mbhele  
Fezokuhle Khumalo  
Thabang Serake  
Bruna Galvão  
Arghavan Alisoltani  
(U. California)

Robert Wilkinson  
Darren Martin  
Nicola Mulder  
Wendy Burgers  
Ntobeko Ntusi  
Rageema Joseph  
Sean Wasserman  
Linda Boloko



## Zoonotic arbo and respiratory virus program Centre for Viral Zoonoses Department Medical Virology/ NHLS Tshwane Academic division University of Pretoria



**ZARV research program/UP**  
Marietjie Venter (Head: ZARV)  
Adriano Mendes (Postdoc)  
Amy Strydom (Postdoc)  
Michaela Davis (MSc, intern medical scientist)  
Carien van Niekerk



**NHLS Tshwane**  
Prof Simnikiwe Mayaphi (HOD)

**Funders:**  
GIZ/BMBF: African Network for Improved  
diagnostics and epidemiology of common and  
emerging infectious agents (ANDEMIA)  
G7 Global Health fund, Robert Koch Institute, Dr  
Fabian Leendertz

## National Institute for Communicable Diseases



### Centre for Respiratory Diseases & Meningitis

Anne von Gottberg  
Thabo Mohale  
Daniel Amoako  
Josie Everatt  
Boitshoko Mahlangu  
Noxolo Ntuli  
Anele Mnguni  
Amelia Buys  
Cardia Fourie  
Noluthando Duma  
Linda de Gouveia  
Jackie Kleynhans  
Nicole Wolter  
Sibongile Walaza  
Mignon du Plessis  
Stefano Tempia  
Mvuyo Makhasi  
Cheryl Cohen

### Centre for HIV and STIs

Jinal Bhiman  
Cathrine Scheepers  
Constantinos Kurt Wibmer  
Thandeka Moyo  
Tandile Hermanus  
Frances Ayres  
Zanele Molaudzi  
Bronwen Lambson  
Tandile Hermanus  
Mashudu Madzivhandila  
Prudence Kgagudi  
Brent Oosthuysen  
Penny Moore  
Lynn Morris

### NICD Groups

NICD COVID-19 response team  
NICD SARS-CoV-2 Sequencing  
Group

### Sequencing Core Facility

Zamantungwa Khumalo  
Annie Chan  
Morne du Plessis  
Stanford Kwenda  
Phillip Senzo Mtshali  
Mushal Allam  
Florah Mnyameni  
Arshad Ismail



## University of the Free State



**UFS**  
Dominique Goedhals  
Armand Bester  
Martin Myaga  
Peter Mwangi  
Emmanuel Ogunbayo  
Milton Mogotsi  
Makgotso Maotoana  
Lutfiyya Mohamed



**NHLS Division of Virology**  
Sabeehah Vawda  
Felicity Burt  
Thokozani Mkhize  
Diagnostic laboratory staff



# Additional support and collaborators



## NHLS

Koeleka Mlisana  
Zinhle Makatini  
Eugene Elliot  
Florette K. Treurnicht  
Kathleen Subramoney  
Oluwakemi Laguda-Akingba  
Shareef Abrahams  
Greta Hoyland  
Gloria Selabe  
Elias Bereda  
Jeannette Wadula

## Hyrax Biosciences

Simon Travers

## Cape Town HVTN Laboratory

Erica Anderson-Nissen  
Anneta Naidoo

## Ndlovu Research

Hugo Tempelman  
CJ Umunnakwe

## Lancet

Allison J. Glass  
Raquel Viana

## Ampath

Terry Marshall  
Cindy van Deventer  
Eddie Silberbauer

## Pathcare Vermaak

Andries Dreyer  
Howard Newman  
Riaan Writes  
Marianne Wolfaardt  
Warren Lowman

## Bridge-the-Gap

Raymond Rott

## Cytespace Africa Laboratories

Christa Viljoen

## ARC-OVI

Lia Rotherham

## CAPRISA

Salim Abdool Karim  
Nigel Garret

## UKZN - Big Data

Francesco Pettruccione  
Ilya Sinayskiy

## University of Oxford

José Lourenço

## FioCruz, Brazil

Vagner Fonseca  
Marta Giovanetti  
Luiz Carlos Junior Alcantara

## Africa CDC

John Nkengasong  
Sofonias Tessema

## Netcare:

Richard Friedland  
Craig Murphy  
Caroline Maslo  
Liza Sitharam

## DSI

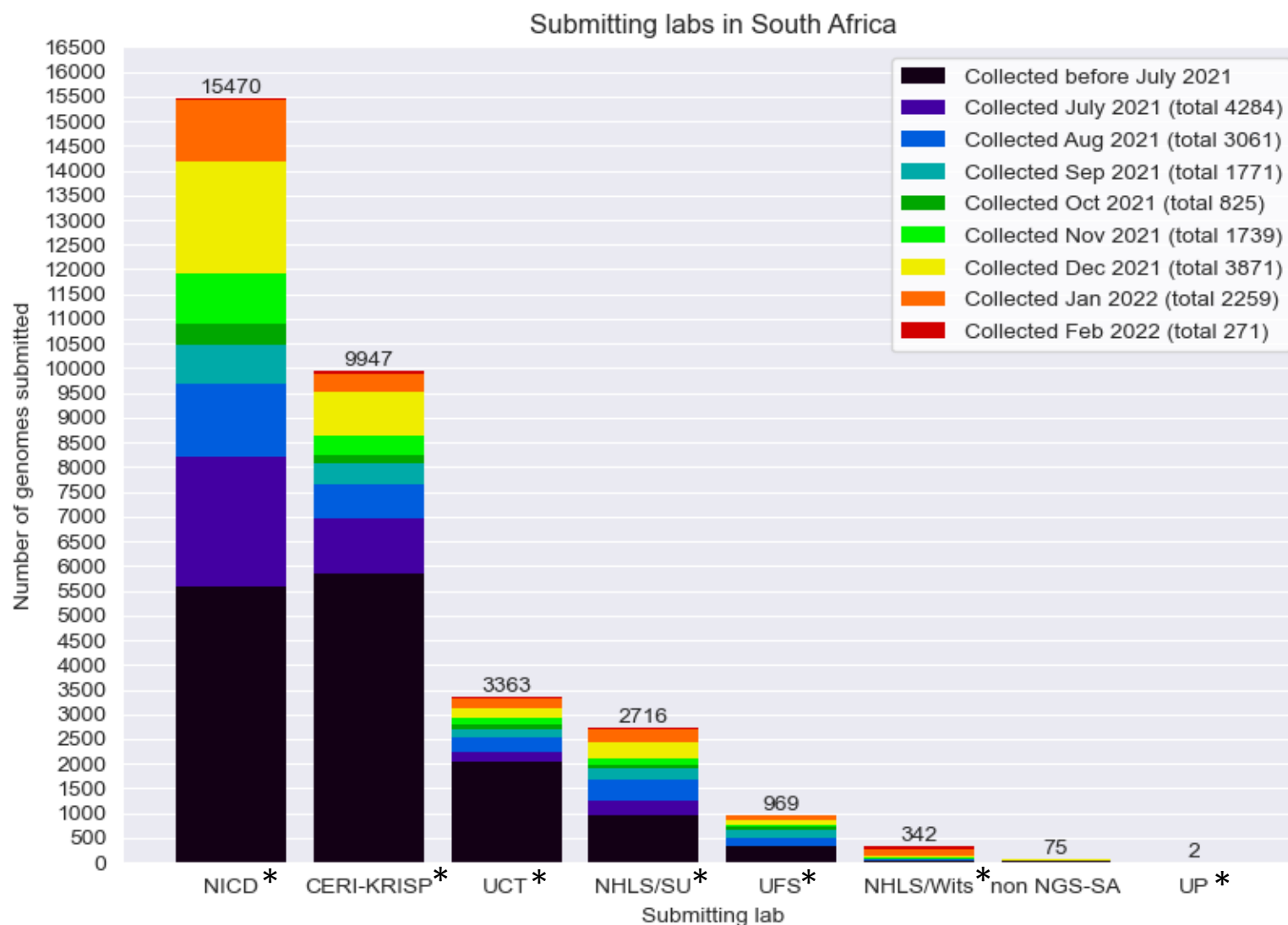
Glaudina Loots

## SA MRC

Glenda Gray



# South African genomes submitted per submitting lab, 2020 - 2022 (N=32 884)



## \*NGS-SA Labs

**CERi:** Centre for Epidemic Response and Innovation

**KRISP:** KZN Research Innovation and Sequencing Platform

**NICD:** National Institute for Communicable Diseases

**NHLS:** National Health Laboratory Service

**SU:** Stellenbosch University

**UCT:** University of Cape Town

**UFS:** University of the Free State

**UP:** University of Pretoria

Multiple labs from NGS-SA and collaborating public and private laboratories are contributing to sequencing, both as originating and as submitting (pictured here) laboratories.

# Variants of Concern (VOC)

WHO label	Pango lineage•	GISAID clade	Nextstrain clade	Additional amino acid changes monitored°	Earliest documented samples	Date of designation
Alpha	B.1.1.7	GRY	20I (V1)	+S:484K +S:452R	United Kingdom, Sep-2020	18-Dec-2020
Beta	B.1.351	GH/501Y.V2	20H (V2)	+S:L18F	South Africa, May-2020	18-Dec-2020
Gamma	P.1	GR/501Y.V3	20J (V3)	+S:681H	Brazil, Nov-2020	11-Jan-2021
Delta	B.1.617.2	G/478K.V1	21A	+S:417N +S:E484K	India, Oct-2020	VOI: 4-Apr-2021 VOC: 11-May-2021
Omicron*	B.1.1.529	GRA	21K, 21L, 21M	+S:R346K	Multiple countries, Nov-2021	VUM: 24-Nov-2021 VOC: 26-Nov-2021

<https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/> accessed 14 January 2022

• Includes all descendant lineages. See the cov-lineages.org and the Pango network websites for further details.

\* See TAG-VE statement issued on 26 November 2021

° Only found in a subset of sequences

# Currently designated Variants of Interest (VOI)

WHO label	Pango lineage*	GISAID clade	Nextstrain clade	Earliest documented samples	Date of designation
Lambda	C.37	GR/452Q.V1	21G	Peru, Dec-2020	14-Jun-2021
Mu	B.1.631	GH	21H	Colombia, Jan-2021	30-Aug-2021

<https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/> accessed 17 December 2021

\* Includes all descendant lineages. See the cov-lineages.org and the Pango network websites for further details.

# Submission of routine specimens for sequencing

- representative of multiple geographic regions (provinces/districts/health facilities) from individuals of
  - all ages
  - over as many time periods during the SARS-CoV-2 epidemic in South Africa
- requested that testing laboratories in both the private and public sectors, submit respiratory samples to their closest NGS-SA sequencing laboratory on a routine basis (ideally every week) as follows, depending on the capacity of the testing laboratory:
  - All positives samples should be sent every week (NGS-SA laboratory will perform random sampling as described below) **OR**
  - A weekly selection of approximately 10%-20% of randomly selected positive samples should be sent every week. Number of selected samples will depend on the size of laboratory and how many other laboratories are drained by the submitting laboratory.

# Submission of special interest specimens for sequencing

In addition to routine samples mentioned above, please send specimens separately to above and clearly marked if:

- Suspected vaccine breakthrough ( $\geq 14$  days after vaccine), especially if hospitalised and clinically severe
- Suspected re-infection ( $\geq 90$  days after previous episode), especially if hospitalised and clinically severe
- Prolonged shedding with high SARS-CoV-2 viral loads (i.e. Ct values less than 30 for more than 1 month post-primary diagnosis) in immunocompromised individuals
- Possible animal-to-human transmission
- Suspected cases of importation from another country, especially countries known to harbour SARS-CoV-2 variants of concern or countries with little available information
- Clusters of “unusual” cases (e.g., in terms of disease presentation, patient groups affected, etc.)