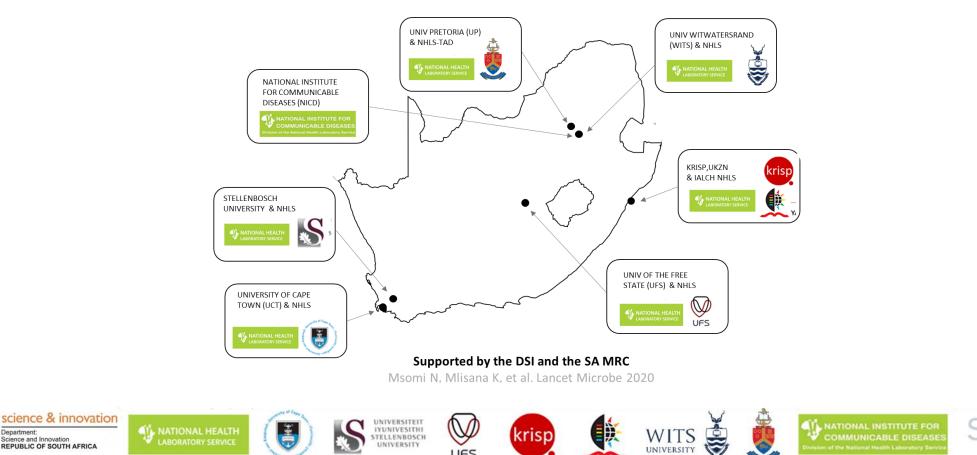


SARS-CoV-2 Sequencing Update **1 April 2022**



Prepared by the National Institute for Communicable Diseases (NICD) of the National Health Laboratory (NHLS) on behalf of the Network for Genomics Surveillance in South Africa (NGS-SA)

Department Science and Innovation

REPUBLIC OF SOUTH AFRICA

The genomic data presented here are based on South African SARS-CoV-2 sequence data downloaded from GISAID (www.gisaid.org) on 1 April 2022 at 16h05

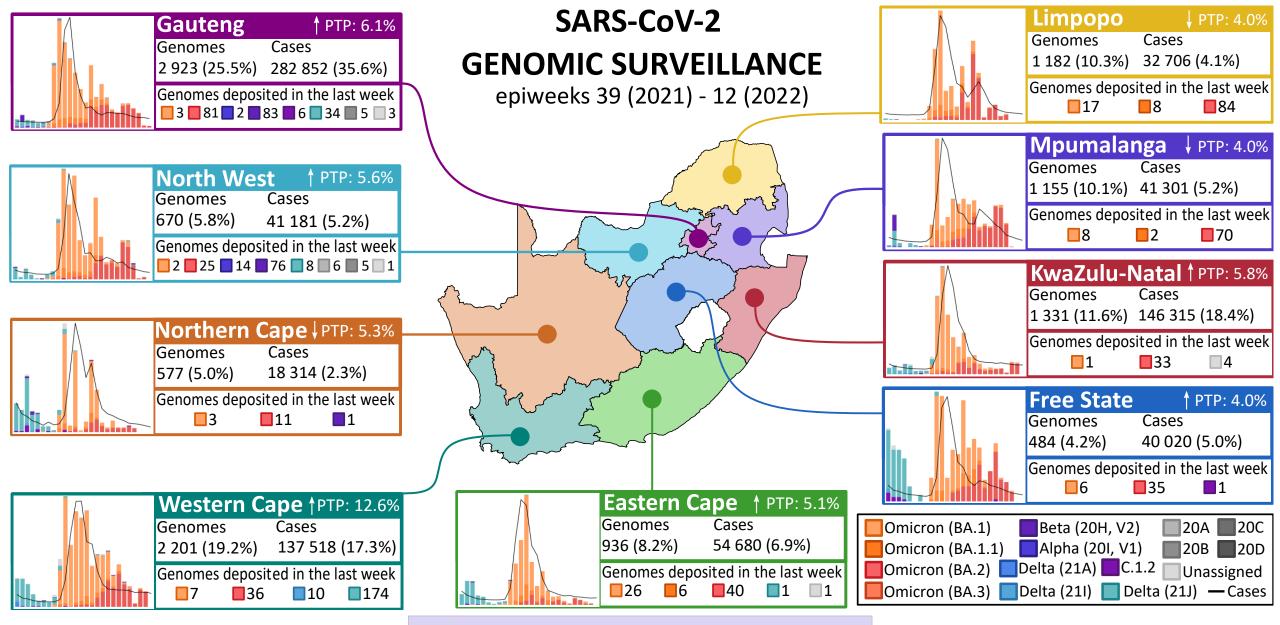


Data license: https://www.gisaid.org/registration/terms-of-use/

Elbe, S., and Buckland-Merrett, G. (2017) Data, disease and diplomacy: GISAID's innovative contribution to global health. Global Challenges, 1:33-46. DOI: 10.1002/gch2.1018 PMCID: 31565258

Shu, Y., McCauley, J. (2017) GISAID: Global initiative on sharing all influenza data – from vision to reality. EuroSurveillance, 22(13) DOI: 10.2807/1560-7917.ES.2017.22.13.30494 PMCID: PMC5388101

Case data is based on specimen collection date. Cases from <u>https://www.nicd.ac.za/diseases-a-z-index/disease-index-covid-19/surveillance-reports/weekly-epidemiological-brief/</u> Test data gives weekly percentage testing positive rates, from <u>https://www.nicd.ac.za/diseases-a-z-index/disease-index-covid-19/surveillance-reports/weekly-testing-summary/</u>



941 genomes deposited in the past week

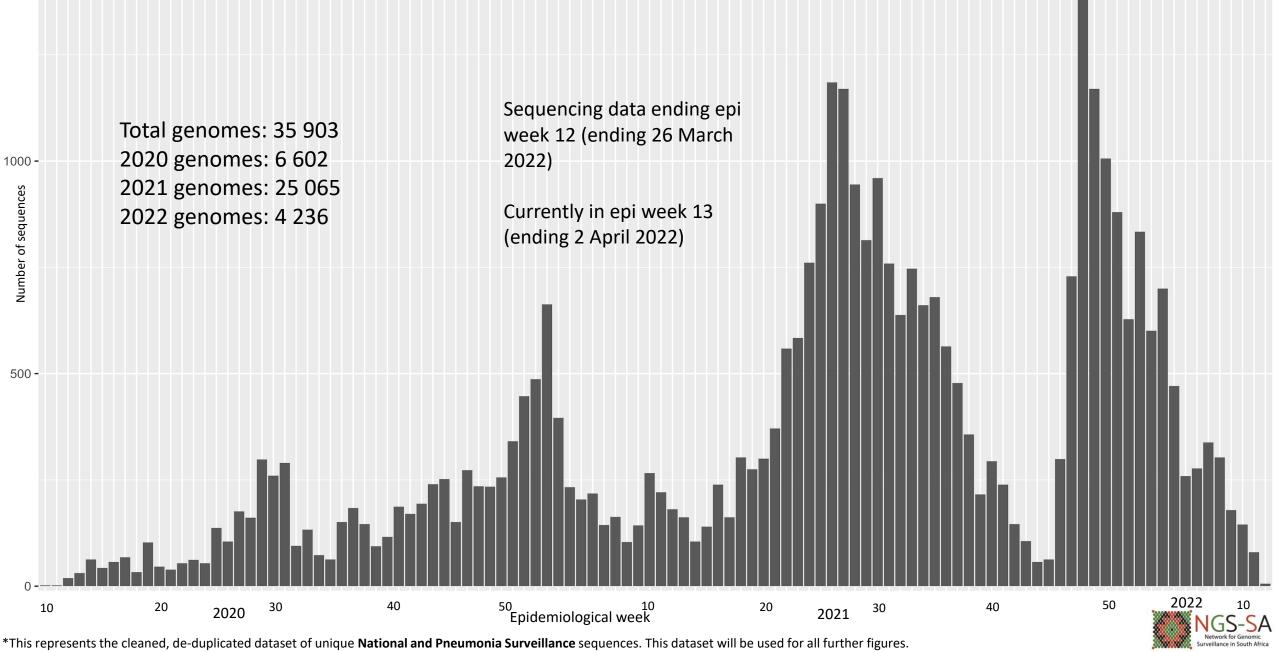
Bar graphs represent genomes sequenced per epiweek, with lines representing cases by collection date (weeks 39 [2021] – 12 [2022])

Genomes and cases presented as provincial total (percentage of national total) for epiweeks 39 (2021) – 12 (2022)

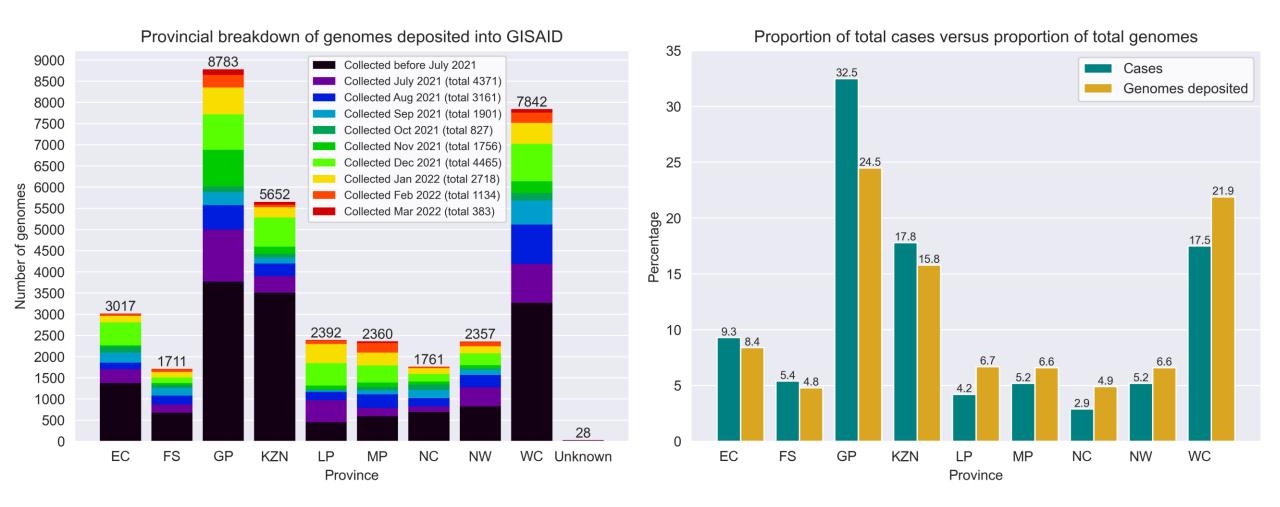
PTP: percentage testing positive in week 12 (20 Mar – 26 Mar); the arrow indicates direction of change since the previous week (13 Mar 2022 – 19 Mar 2022)



Number of South African genomes deposited on GISAID, by specimen collection week, 2020 – 2022 (N=35 903*)

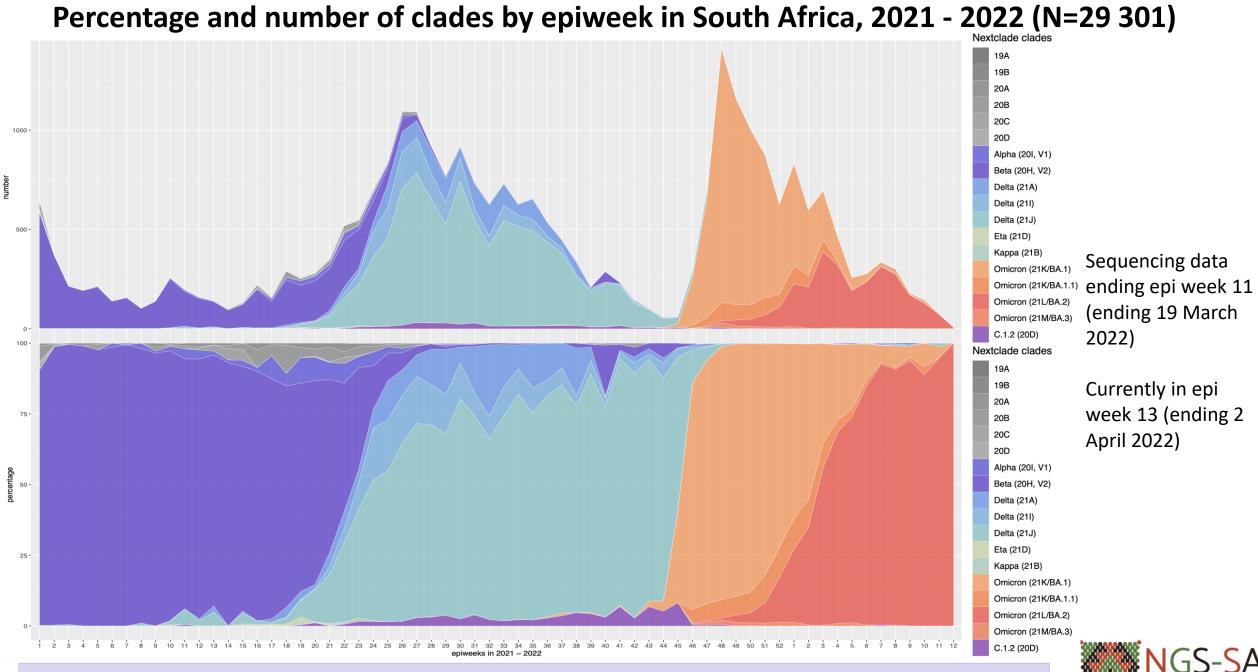


GISAID genomes vs total cases, 2020 – 2022 (N=35 903)



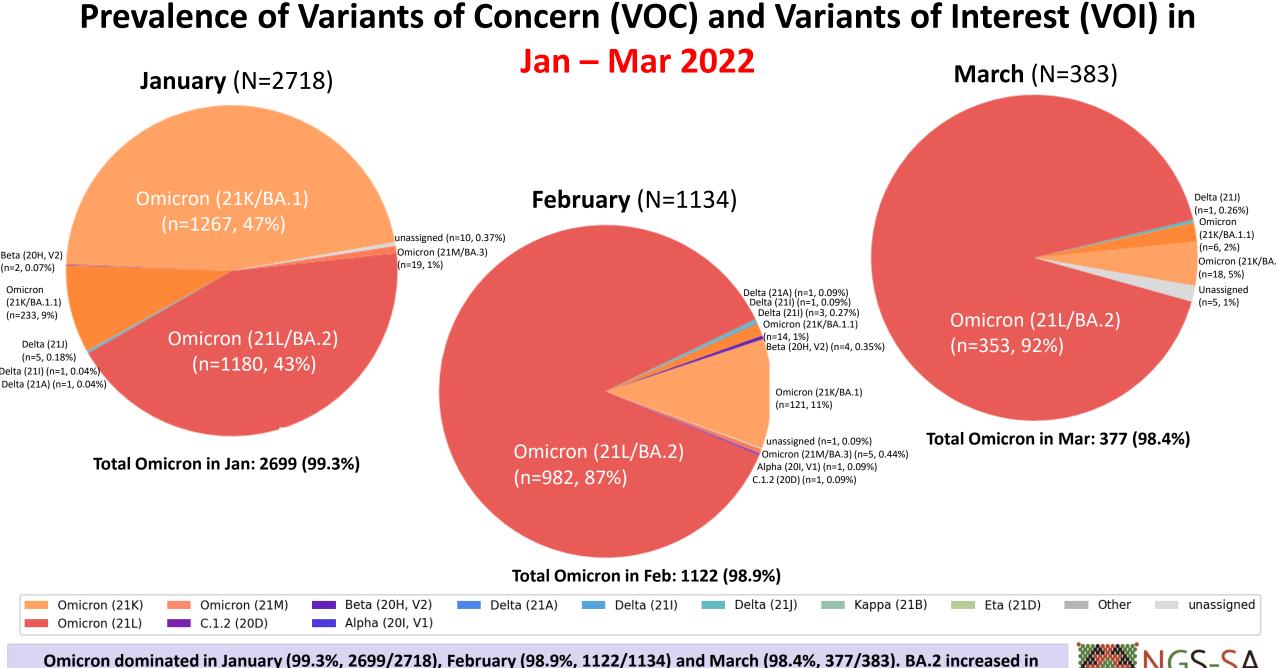
All provinces, apart from GP, LP, NC and WC, have comparable percentages of overall cases and overall sequenced genomes.





Surveillance in South Africa

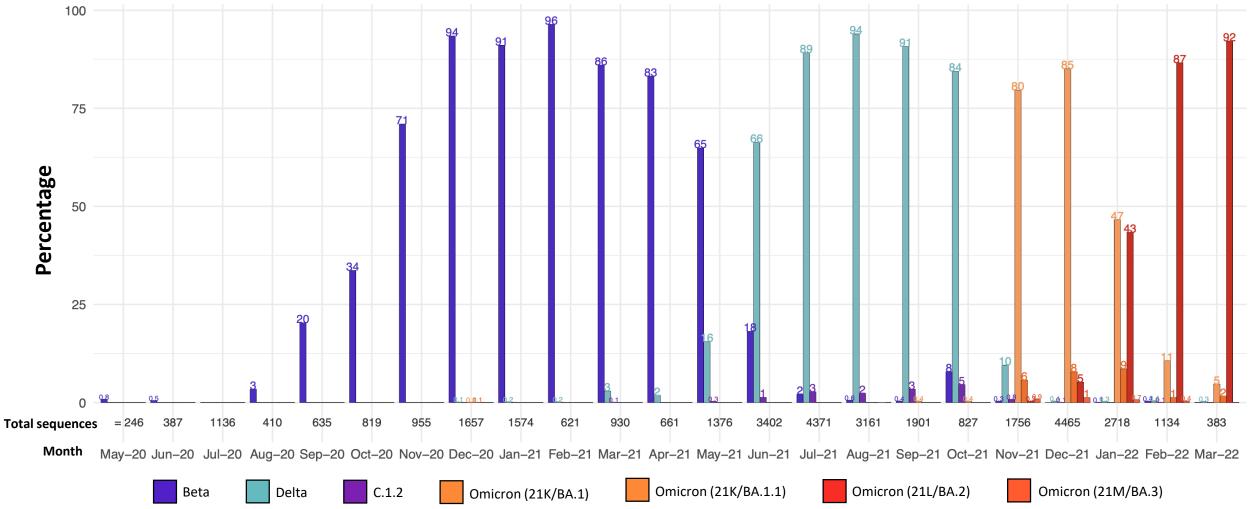
Delta dominated in South Africa until October at >80%. Omicron has dominated from November onwards.



prevalence throughout January and was dominant in February. It appears to still be dominant in March.

Detection Rates: Beta, Delta, C.1.2 and Omicron

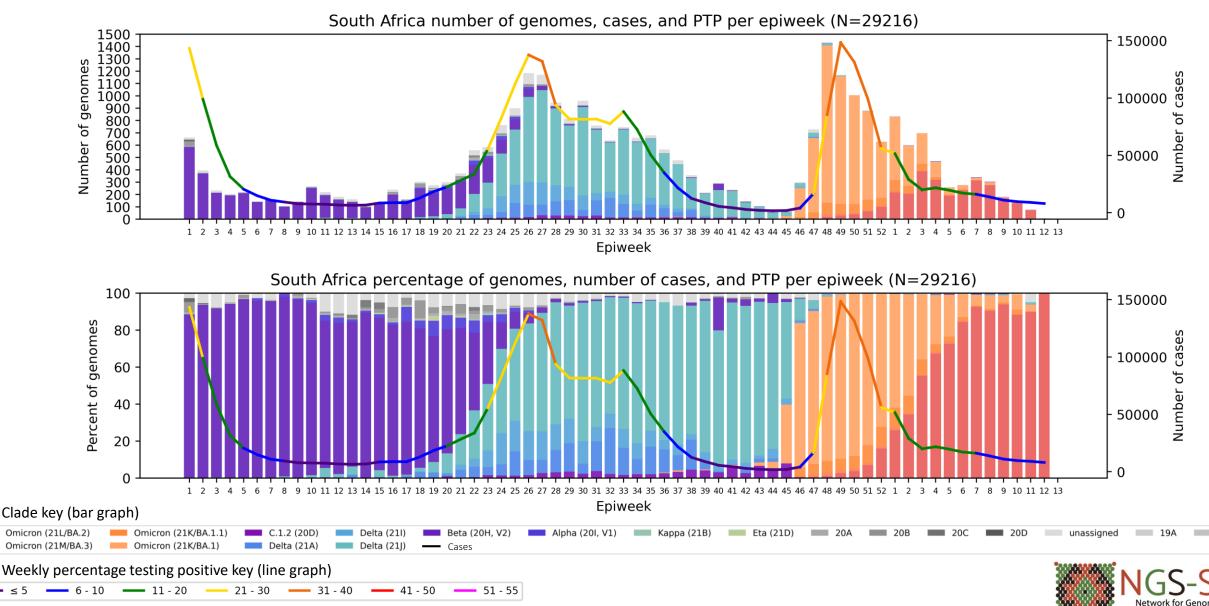
Detection rates of variants being monitored in South Africa*



*Bars represent percentage prevalence of variant for the month; total sequences collected for the month are given below

Omicron has been dominant since November (>80% in November, >98% in December, January, February and March). BA.2 has increased in frequency, making up 43% of genomes in January, 87% in February and 92% in March. BA.3 and other VOCs continue to be detected at low levels.

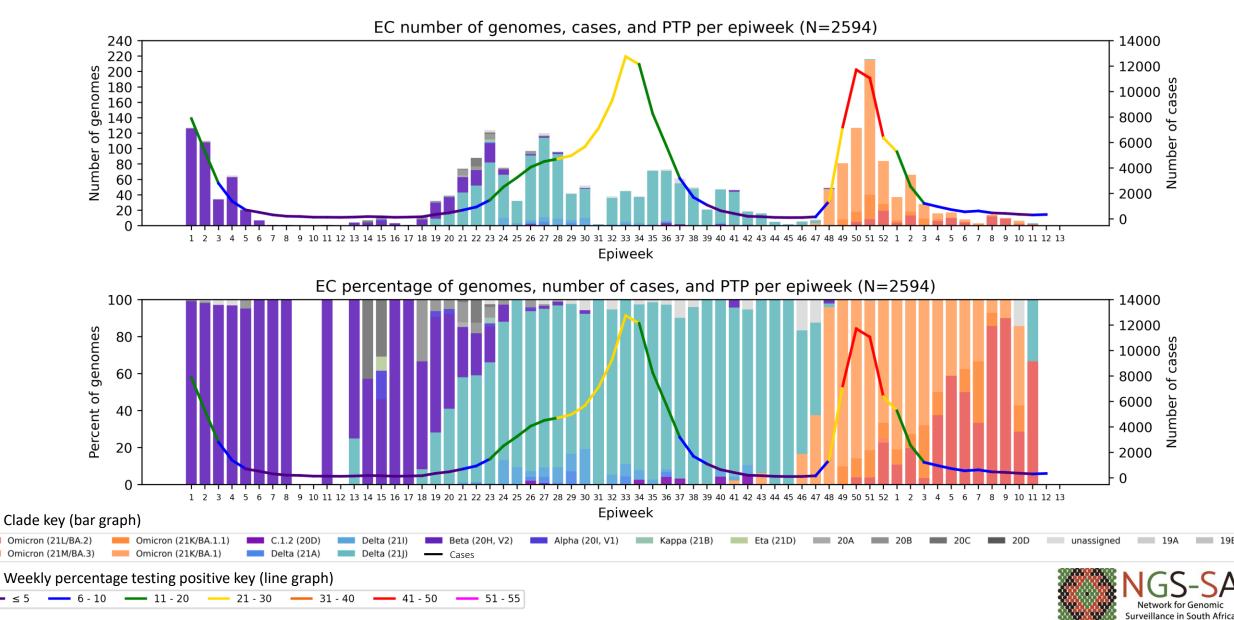
South Africa, 2021-2022, n = 29216*



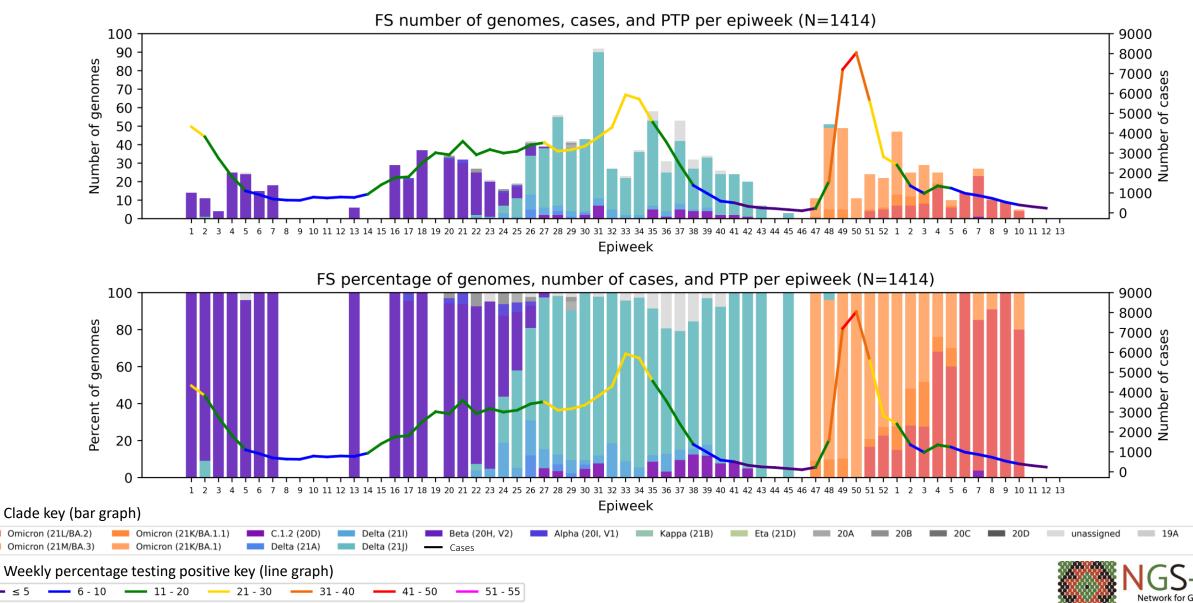
Surveillance in South Africa

*Excludes sequences missing collection dates, as well as those collected January 1st and 2nd 2021 as they are part of epiweek 53 of 2020.

Eastern Cape Province, 2021-2022, n = 2594

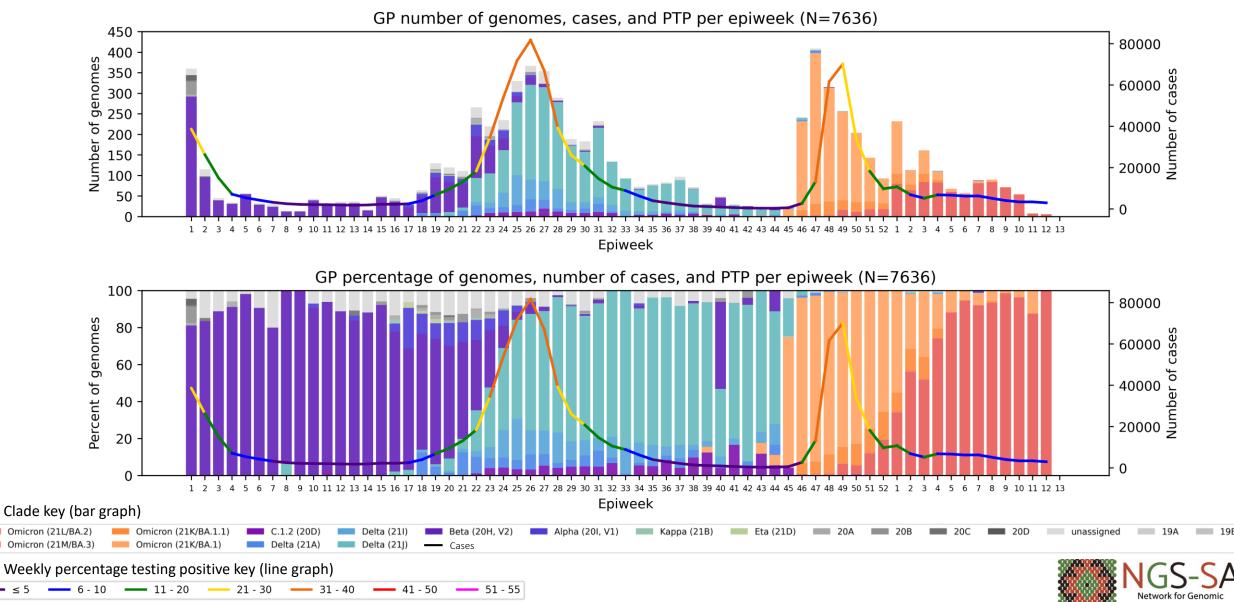


Free State Province, 2021-2022, n = 1414

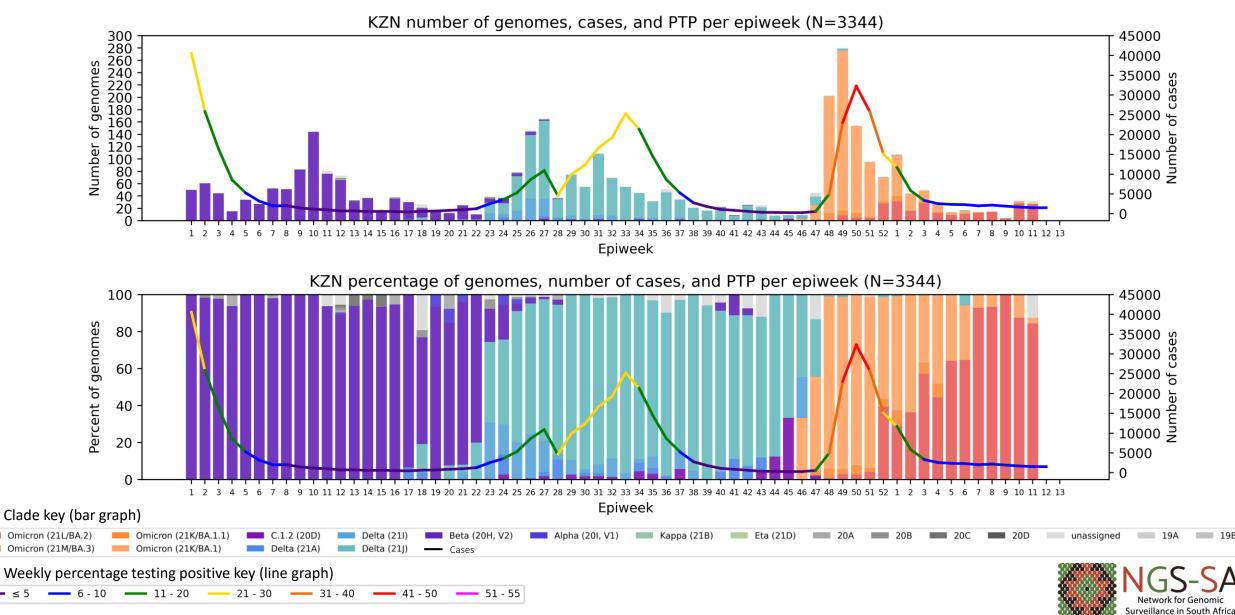


198

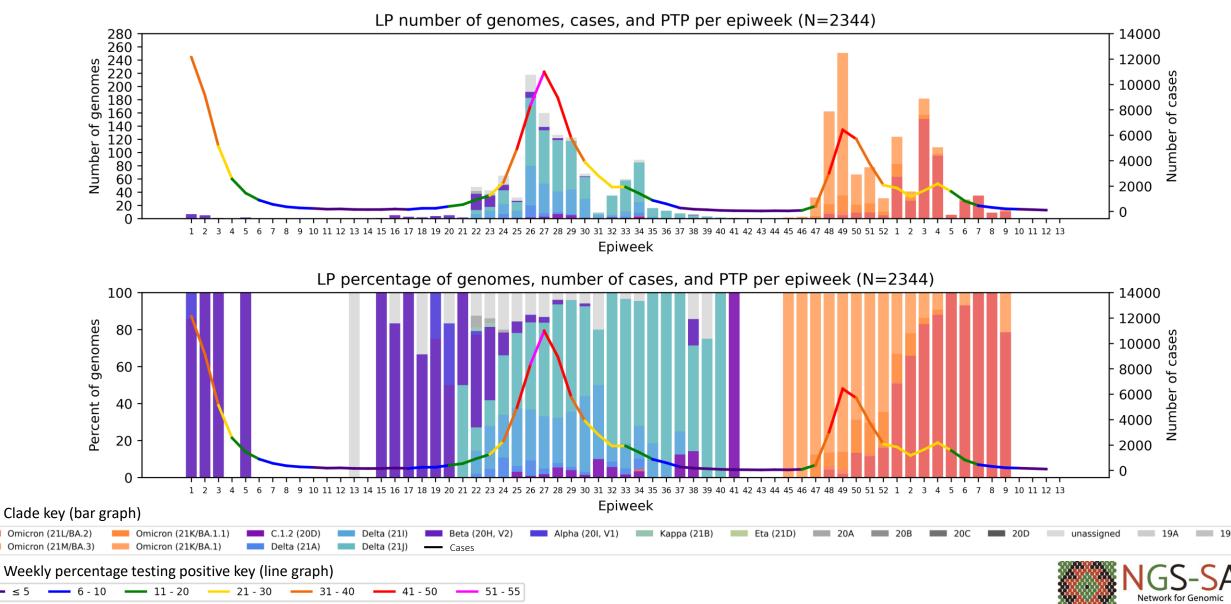
Gauteng Province, 2021-2022, n = 7636



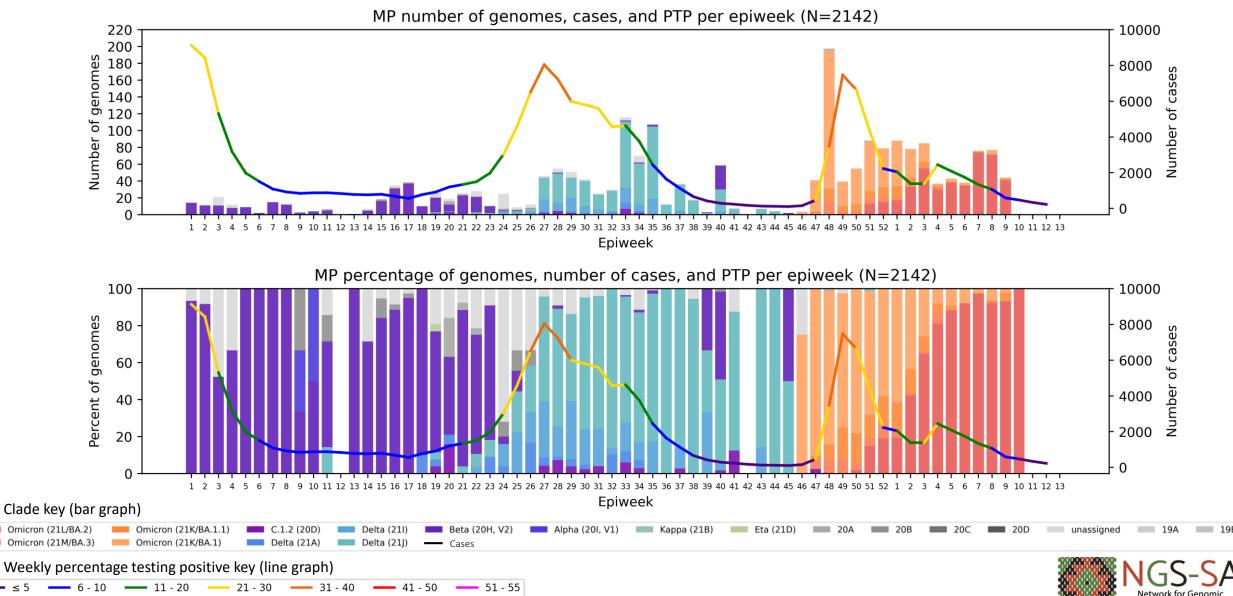
KwaZulu-Natal Province, 2021-2022, n = 3344



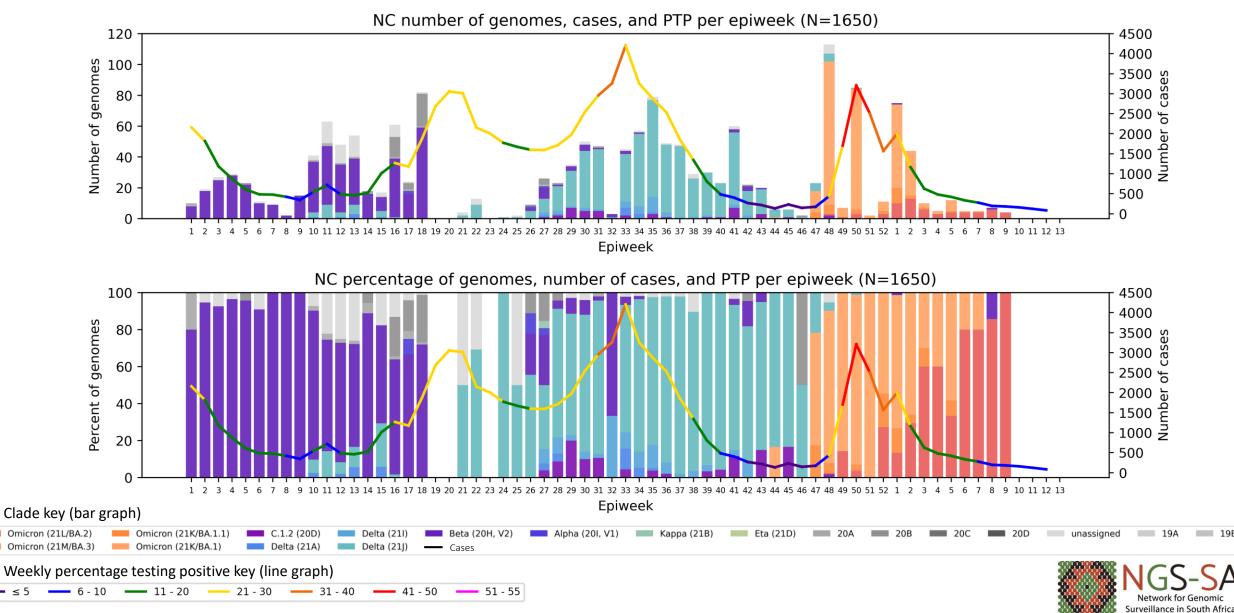
Limpopo Province, 2021-2022, n = 2344



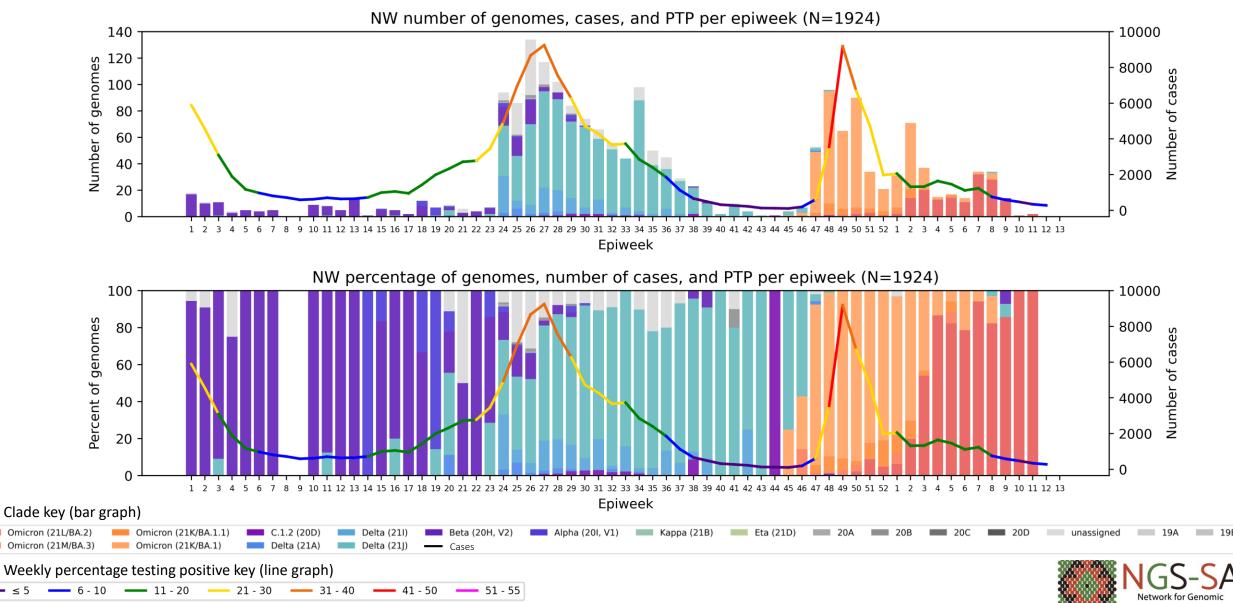
Mpumalanga Province, 2021-2022, n = 2142



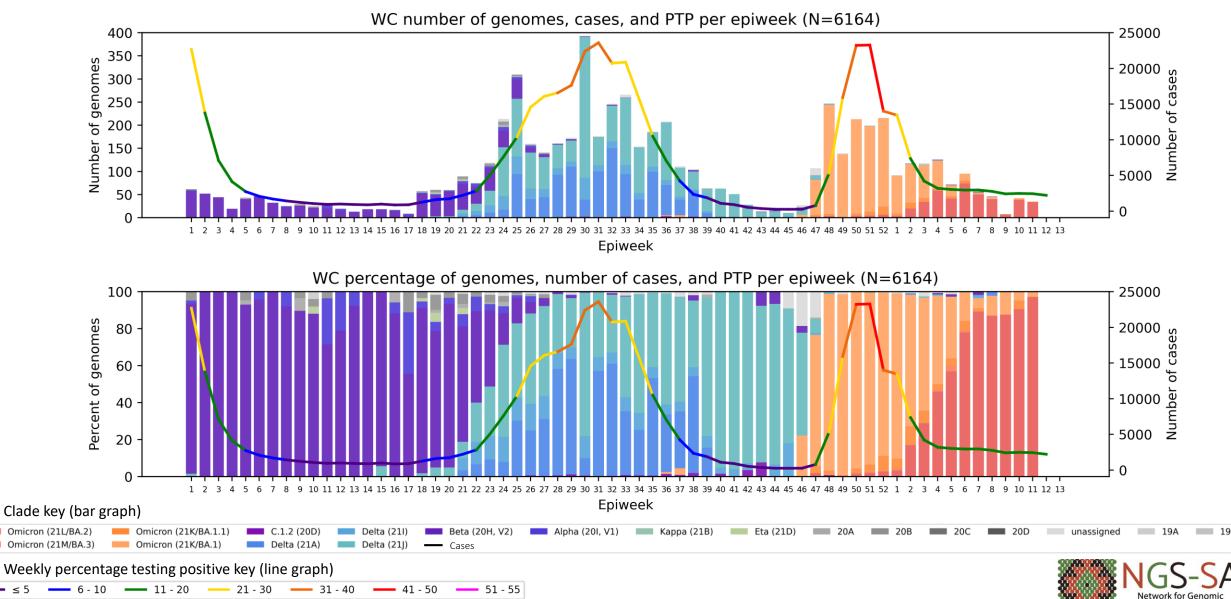
Northern Cape Province, 2021-2022, n = 1650



North West Province, 2021, n = 1924



Western Cape Province, 2021-2022, n = 6164



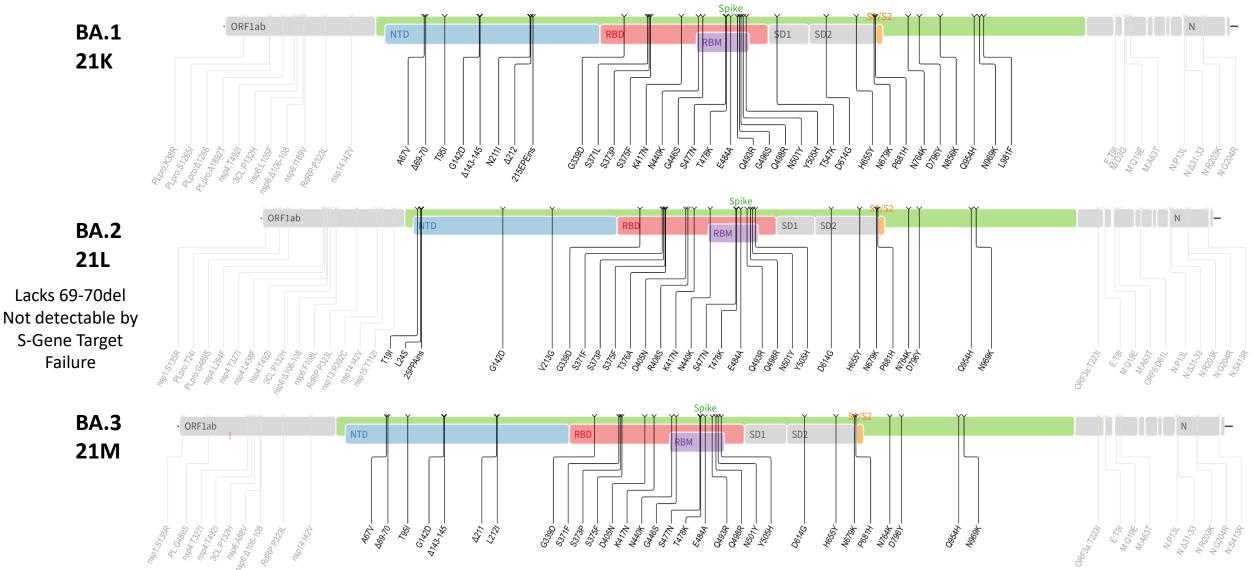
Summary

• Variant of Concern Omicron

- Detected in at least 153 countries and dominating globally
- Split into different lineages based on different mutational profiles. Predominant lineages currently BA.1 (21K), BA.1.1 (21K, BA.1+spike R346K), BA.2 (21L), BA.3 (21M). More lineages are being assigned but defining mutations are not yet available.
- South Africa (detected in all provinces):
 - Dominated January, February and March sequencing data at >99% of genomes
 - While BA.1 was the predominant sub-lineage in January (47%), the proportion of BA.2 increased from 43% in January to 87% in February and 92% in March
 - BA.3 continues to be detected at low levels
- NGS-SA teams are monitoring for recombinants
- Low frequency of previously circulating variants such as Delta and Beta still detected in recent data



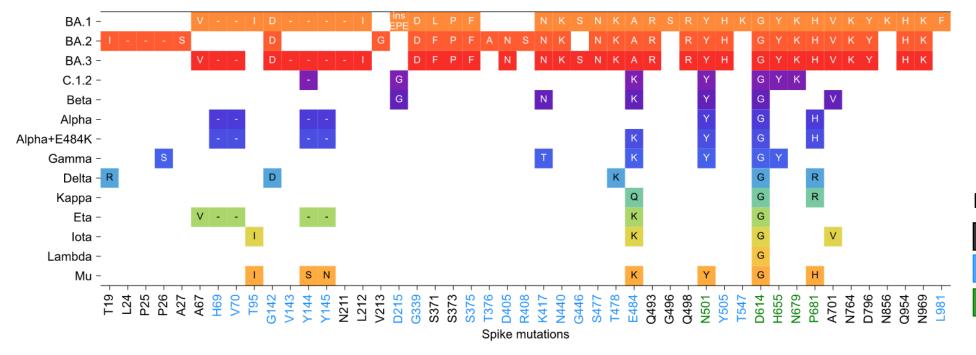
Omicron sub-lineage spike mutation profiles





Lineage definitions based on <u>https://github.com/cov-lineages/pango-designation/issues/367</u> Images from <u>https://covdb.stanford.edu/page/mutation-viewer/</u>

Omicron spike mutations compared to other VOC/VOIs



Only lineage-defining mutations are pictured here. Low prevalence mutations can be seen on the following slide.

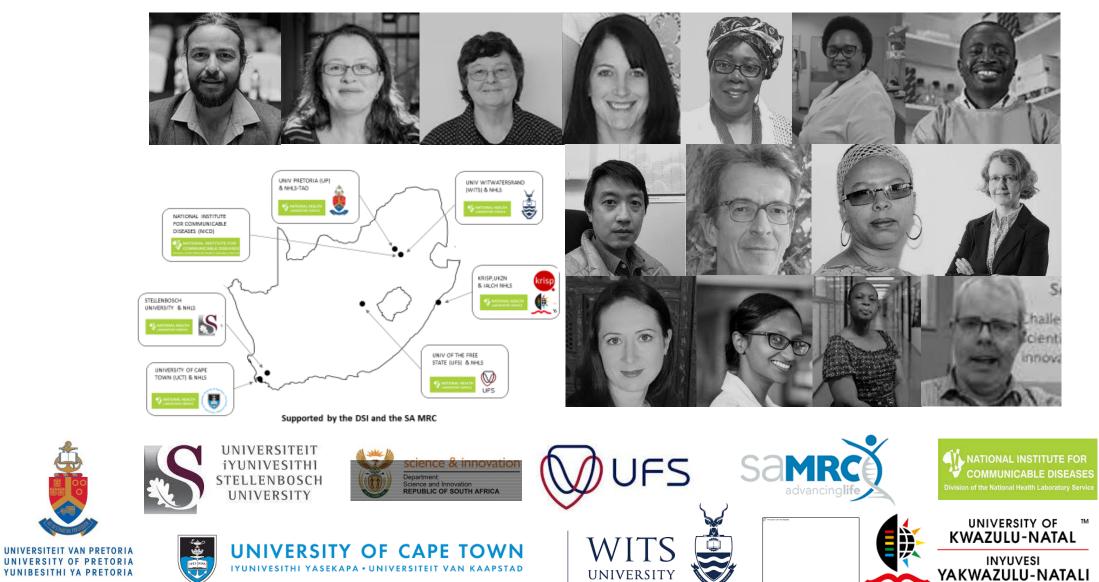
Mutation impact key

Unknown or unconfirmed impact Known/predicted immune escape Enhanced infectivity

- Multiple changes within the two immunogenic regions in S1 (NTD and RBD)
 - including a three amino acid insertion
- Accumulation of mutations surrounding the furin cleavage site
 - Including combination of N679K and P681H
- Effect of most spike S2 subunit changes have not been defined, but may be linked to immune escape







This project (RIA2020EF-3030) is part of the EDCTP2 programme supported by the European Union"

XX

ЕDСТР

NATIONAL HEALTH LABORATORY SERVICE

University of Stellenbosch & NHLS Tygerberg Virology



Susan Engelbrecht Wolfgang Preiser Gert van Zyl Tongai Maponga **Bronwyn Kleinhans** Shannon Wilson Karabo Phadu Tania Stander Kamela Mahlakwane Mathilda Claassen **Diagnostic laboratory staff**

University of Cape Town, NHLS & Western Cape Government



NHLS-UCT

Carolyn Williamson

Nei-yuan Hsiao

Diana Hardie

Kruger Marais

Stephen Korsman

Ziyaad Valley-Omar

Department. Health REPUBLIC OF SOUTH ATRICA WCG-UCT Mary-Anne Davies Hannah Hussev Andrew Boulle

health

Masudah Paleker Theuns Jacobs



UCT. IDM and CIDRI-Africa

Deelan Doolabh Arash Iranzadeh Lynn Tyers Innocent Mudau Nokuzola Mbhele Fezokuhle Khumalo Thabang Serakge Bruna Galvão Linda Boloko Arghavan Alisoltani (U. California)

NHLS Greenpoint Annabel Enoch

Erna Morden



Robert Wilkinson



cience & innovation Alment: New and Innevation

ΧŻ

EDCTP

W

Samrc

CAPE TOWN HVTN

UKZN-Inkosi Albert Luthuli Central Hospital



Dr Khanyi Msomi Dr Pravi Moodley Dr Aabida Khan Dr Lili Gounder Dr Kerri Francois Dr Cherise Naicker Dr Joedene Chetty

UNIVERSITY OF INYUVESI YAKWAZULU-NATALI

Dr Neli Ngcaba Dr Kerusha Govender Dr Tshepiso Mosito Mr Malcolm Ellapen Mr Kubendran Reddy The COVID-19 Bench team

University of KwaZulu-Natal & Africa **Health Research Institute**



Tulio de Oliveira Richard Lessels Houriivah Tegally Eduan Wilkinson Jennifer Giandhari Sureshnee Pillav **Emmanuel James San**

KRISP at UKZN:



National Institute for Communicable Diseases

Jinal Bhiman

Cathrine Scheepers

Thandeka Movo

Frances Ayres

Zanele Molaudzi

Bronwen Lambson

Tandile Hermanus

Prudence Kgagudi

Brent Oosthuysen

Penny Moore

Lynn Morris

NICD Groups

Group

Mashudu Madzivhandila

NICD COVID-19 response team

NICD SARS-CoV-2 Sequencing

Tandile Hermanus

Diseases & Meningitis Boitshoko Mahlangu Mignon du Plessis

Stefano Tempia Mvuyo Makhasi **Cheryl Cohen**



Centre for HIV and STIs Sequencing Core Facility Zamantungwa Khumalo Annie Chan **Constantinos Kurt Wibmer** Morne du Plessis Stanford Kwenda **Mushal Allam** Florah Mnyameni

SamRC







University of the **Free State**



UFS

Dominique Goedhals Armand Bester Martin Myaga Peter Mwangi Emmanuel Ogunbayo Milton Mogotsi Makgotso Maotoana Lutfiyya Mohamed



NHLS Division of Virology Sabeehah Vawda Felicity Burt Thokozani Mkhize **Diagnostic laboratory staff**



Zoonotic arbo and respiratory virus program **Centre for Viral Zoonoses Department Medical Virology/ NHLS Tshwane Academic division University of Pretoria**

> ZARV research program/UP Marietjie Venter (Head: ZARV) Adriano Mendes (Postdoc) Amy Strydom (Postdoc) Michaela Davis (MSc, intern medical scientist) Carien van Niekerk

NHLS Tshwane Prof Simnikiwe Mayaphi (HOD)

Funders:

٩

GIZ/BMBF: African Network for Improved diagnostics and epidemiology of common and emerging infectious agents (ANDEMIA) G7 Global Health fund, Robert Koch Institute, Dr Fabian Leendertz

Centre for Respiratory

Anne von Gottberg Thabo Mohale Daniel Amoako Josie Everatt Noxolo Ntuli Anele Mnguni Amelia Buys Cardia Fourie Noluthando Duma Linda de Gouveia Jackie Kleynhans Nicole Wolter Sibongile Walaza



Phillip Senzo Mtshali Arshad Ismail









Key to Diagnostic Excellent

ΑΜΡΑΤΗ

LABORATORIES

PathCare

1

Vermaak

africa

aboratorie

FIOCRUZ

NHLS Koeleka Mlisana Zinhle Makatini Eugene Elliot Florette K. Treurnicht Kathleen Subramoney Oluwakemi Laguda-Akingba Shareef Abrahams Greta Hoyland Gloria Selabe

Jeannette Wadula

Elias Bereda

Hyrax Biosciences Simon Travers

Cape Town HVTN Laboratory Erica Anderson-Nissen Anneta Naidoo

Ndlovu Research Hugo Tempelman CJ Umunnakwe

Lancet Allison J. Glass Raquel Viana

Ampath Terry Marshall Cindy van Deventer Eddie Silberbauer

Pathcare Vermaak Andries Dreyer Howard Newman Riaan Writes Marianne Wolfaardt Warren Lowman

Bridge-the-Gap Raymond Rott

Cytespace Africa Laboratories Christa Viljoen

ARC-OVI Lia Rotherham **CAPRISA** Salim Abdool Karim Nigel Garret

Additional support and collaborators

UKZN - Big Data Francesco Pettruccione Ilya Sinayskiy

University of Oxford José Lourenço

FioCruz, Brazil Vagner Fonseca Marta Giovanetti Luiz Carlos Junior Alcantara Africa CDC John Nkengasong Sofonias Tessema

Netcare: Richard Friedland Craig Murphy Caroline Maslo Liza Sitharam

DSI

Glaudina Loots

SA MRC Glenda Gray











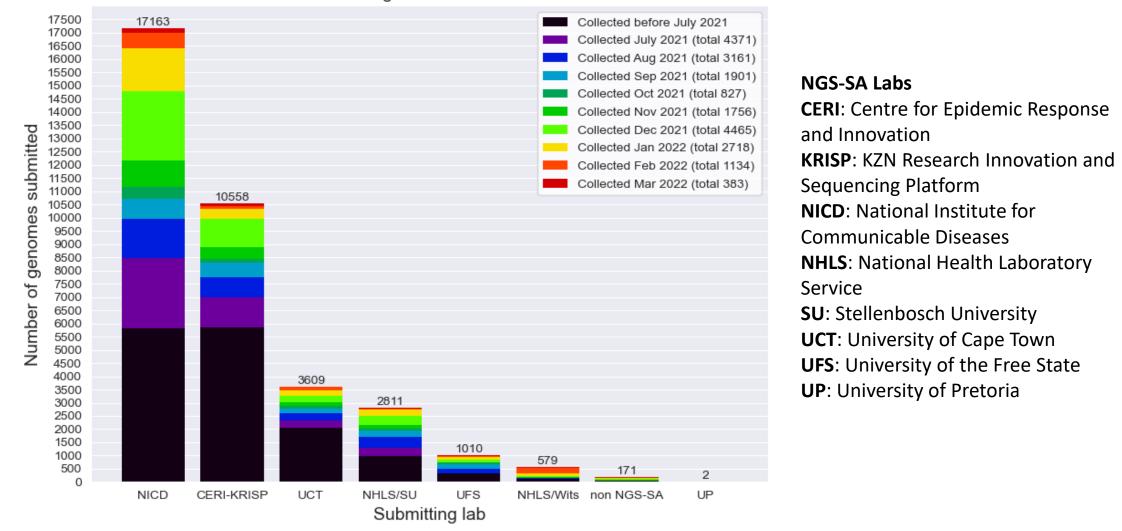






South African genomes submitted per submitting lab, 2020 - 2022 (N=35 903)

Submitting labs in South Africa



Multiple labs from NGS-SA and collaborating public and private laboratories are contributing to sequencing, both as originating and as submitting (pictured here) laboratories.



Currently circulating Variants of Concern (VOC)

WHO label	Pango lineage∙	GISAID clade	Nextstrain clade	Additional amino acid changes monitored°	Earliest documented samples	Date of designation
Delta	B.1.617.2	G/478K.V1	21A, 21I, 21J	+S:K417N +S:K484K	India, Oct-2020	VOI: 4-Apr-2021 VOC: 11-May-2021
Omicron*	B.1.1.529	GR/484A	21K	+S:R346K	Multiple countries, Nov-2021	VUM: 24-Nov-2021 VOC: 26-Nov-2021

https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/ accessed 18 March 2022

•Includes all descendant lineages. See the cov-lineages.org and the Pango network websites for further details.

• Only found in a subset of sequences

Previously circulating Variants of Concern

WHO label	Pango lineage∙	GISAID clade	Nextstrain clade	Earliest documented samples	Date of designation
Alpha	B.1.1.7	GRY	20I (V1)	United Kingdom, Sep-2020	VOC: 18-Dec-2020 Previous VOC: 09-Mar-2022
Beta	B.1.351	GH/501Y.V2	20H (V2)	South Africa, May-2020	VOC: 18-Dec-2020 Previous VOC: 09-Mar-2022
Gamma	P.1	GR/501Y.V3	20J (V3)	Brazil, Nov-2020	VOC: 11-Jan-2021 Previous VOC: 09-Mar-2022

https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/ accessed 18 March 2022

• Includes all descendant lineages. See the cov-lineages.org and the Pango network websites for further details.

Submission of routine specimens for sequencing

- representative of multiple geographic regions (provinces/districts/health facilities) from individuals of
 - all ages
 - over as many time periods during the SARS-CoV-2 epidemic in South Africa
- requested that testing laboratories in both the private and public sectors, submit respiratory samples to their closest NGS-SA sequencing laboratory on a routine basis (ideally every week) as follows, depending on the capacity of the testing laboratory:
 - All positives samples should be sent every week (NGS-SA laboratory will perform random sampling as described below) OR
 - A weekly selection of approximately 10%-20% of randomly selected positive samples should be sent every week. Number of selected samples will depend on the size of laboratory and how many other laboratories are drained by the submitting laboratory.

Submission of special interest specimens for sequencing

In addition to routine samples mentioned above, please send specimens separately to above and clearly marked if:

- Suspected vaccine breakthrough (≥14 days after vaccine), especially if hospitalised and clinically severe
- Suspected re-infection (≥90 days after previous episode), especially if hospitalised and clinically severe
- Prolonged shedding with high SARS-CoV-2 viral loads (i.e. Ct values less than 30 for more than 1 month post-primary diagnosis) in immunocompromised individuals
- Possible animal-to-human transmission
- Suspected cases of importation from another country, especially countries known to harbour SARS-CoV-2 variants of concern or countries with little available information
- Clusters of "unusual" cases (e.g., in terms of disease presentation, patient groups affected, etc.)