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OVERVIEW

This report summarises and interprets findings from detection, quantification and sequencing of SARS-CoV-2 by the National Institute for Communicable Diseases (NICD) Centre for Vaccines and Immunology from influent (untreated) wastewater in 17 wastewater treatment plants (WWTP) across five South African provinces. The results obtained and interpretations analysis of levels of SARS-CoV-2 in WWTP from 70 additional plants across South Africa were tested by SACCESS partners: the National Institute for Occupational Health (NIOH), Lumegen Laboratories, GreenHill Laboratories, SAMRC-TB Platform (until March 2022) and Praecautio to cover all provinces in South Africa. Levels of SARS-CoV-2 in wastewater correlate with population levels of SARS-CoV-2 over time and indicate the geographic distribution of disease. Variants of SARS-CoV-2 can be identified in wastewater through detection of single-nucleotide polymorphisms (SNPs) that are specific to each variant. These variants are shown to correspond to variants prevalent in clinical cases, across time and place. SARS-CoV-2 is shed from symptomatic and asymptomatic persons in stool but is not transmitted by faecal-oral route nor via wastewater. This report is based on data collected from June 2021 until 1st April 2022 (epidemiological week 13). Results from wastewater testing should be read and interpreted together with testing and genomic reports generated by the Centre for Respiratory Diseases and Meningitis found at https://www.nicd.ac.za/diseases-a-z-index/disease-index-covid-19/surveillance-reports/

- Part 1 of this report presents methods and results of quantitative testing of wastewater.
- Part 2 of this report presents methods and results from sequencing of SARS-CoV-2 RNA fragments in wastewater.

Generally, SARS-CoV-2 in wastewater remains at low levels across the country corresponding to the low numbers of clinical cases after the 4th wave. Omicron variant is present in wastewater across the country. Detailed analyses are described below.

HIGHLIGHTS

- SARS-CoV-2 levels in wastewater: Wastewater levels are increasing in Gauteng, particularly in City of Johannesburg (Goudkoppies WWTP), Tshwane North (Klipgat and Rooiwal WWTPs), Tshwane South, (Godrich WWTP and Summerplace WWTPs), with levels consistently high in Tshwane central (Daspoort WWTP) at almost 2log copies/ml for two weeks. These increases may be an early signal of an upswing in cases which will be confirmed through ongoing surveillance.
- **SARS-CoV-2 genomics in wastewater:** Sequencing data show the presence of omicron variant in all recent samples across South Africa (up to week 7, 2022). Mutations, associated with the VOC delta, were found in Gauteng (week 4, Tshwane and week 7, Johannesburg). *note this report is unchanged since week ending 1 April 2022.















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DETECTION, QUANTITATION AND GENOMIC SEQUENCING AT SENTINEL SITES IN SOUTH AFRICA, MARCH 2021- MARCH 2022

CO-FUNDED BY THE WATER RESEARCH COMMISSION, THE BILL AND MELINDA GATES FOUNDATION AND THE NICD

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- 7. Greenhill Laboratories
- 8. Praecautio
- 9. Tuberculosis Platform, South African Medical Research Council, Pretoria.
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- 11. City of Cape Town Health Department
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PART 1: DETECTION AND QUANTIFICATION OF SARS-COV-2 IN WASTEWATER

BACKGROUND

The detection and monitoring of SARS-CoV-2 through wastewater was first proposed in April 2020. Initial reports describing the feasibility and practical usefulness of this approach emerged simultaneously from several countries during August 2020. Recent evidence has shown that SARS-CoV-2 can be detected in wastewater prior to the appearance of clinical cases, and longitudinal tracking of SARS-CoV-2 viral load in wastewater correlates with the burden of clinically diagnosed cases. Furthermore, the sequencing of SARS-CoV-2 RNA fragments in wastewater has identified variants of concern as well as mutations not detected in clinical cases.

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In South Africa, SARS-CoV-2 epidemiology is monitored through laboratory testing of clinical cases using reverse-transcriptase polymerase chain reaction (RT-PCR) tests and rapid antigen tests, COVID-19 hospital admissions and COVID-19 - related deaths. Laboratory testing data is sent by testing laboratories to the National Institute for Communicable Diseases (NICD) via the DATCOV system. From these data sources, epidemiological indicators including incidence rates of testing and case detection, hospitalisation and death rates are made available to key stakeholders and the general public.

Clinical epidemiology based on reporting of laboratory-confirmed cases of SARS-CoV-2 has limitations. Household transmission studies in South African urban and rural settings have demonstrated that a large proportion of cases are asymptomatic or so mild as not to elicit health-seeking, and that laboratory-confirmed cases likely represent less than 10% of SARS-CoV-2 cases prevalent in a community at any given time. Secondly, there is increasing use of rapid antigen detection tests in clinical settings. Results of these tests may not be reported to surveillance networks. Consequently, laboratory diagnosis is increasingly less representative of the burden of disease.

In November 2020, a network of testing laboratories, which became known as the South African Collaborative COVID-19 Environmental Surveillance System (SACCESS) network, was established in order to support the development of a common testing methodology, identify and address challenges, and share best practices related to qualitative, quantitative and sequencing of SARS-CoV-2 in wastewater. Treatment of wastewater in South Africa is the responsibility of local government. Approximately 1050 wastewater treatment works (WWTPs) are administered by metropolitan councils and local government and treat industrial and domestic waste. SACCESS partners and the NICD have engaged with local government to support sample collection, interpretation and utilisation of the results for public health purposes.

The SACCESS network aims to detect and quantify SARS-CoV-2 in wastewater in urban settings in South Africa, to compare trends, temporal and geographic distribution of SARS-CoV-2 levels in wastewater with trends in clinical epidemiology so as to support the use of wastewater-based epidemiology for COVID-19 outbreak prevention and response activities.

METHODS

Outbreak context and clinical case epidemiology

Since the first case of SARS-CoV-2 in South Africa was detected on 3rd March 2020, laboratories in the country have conducted over 22 million RT-PCR and antigen tests. Four distinct waves of SARS-CoV-2 infection occurred, peaking in June 2020, December 2020, July 2021, and December 2021 respectively. The current de-duplicated and geospatially allocated national line list of laboratory-confirmed cases of SARS-CoV-2 (identified by RT-PCR or antigen test) is provided by the NICD for comparison with results from SARS-CoV-2 testing of wastewater.

Establishment of the laboratory testing network

Commencing in 2018, the NICD had been conducting testing of wastewater for poliovirus as part of the National Department of Health's polio surveillance programme. In 2020, the NICD commenced testing of influent wastewater samples from these 18 sites, including eight in Gauteng Province, two in the City of Cape Town (Western Cape Province), two in Mangaung (Free State Province), two in eThekwini (KwaZulu-Natal Province) and four in Eastern Cape Province (two in Buffalo City Metro and two in Nelson Mandela Metro). Quantitative testing results for these sites are available from week 8 of 2021, onwards.

Additional wastewater plants across all metropolitan areas as well as sentinel site plants in smaller provinces were included from February 2021. From August 2021, quantitative testing was conducted

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on all specimens submitted to partner laboratories for testing. Presently, samples from 87 WWTPs are being tested for SARS-CoV-2. The supplementary Table 1 at the bottom of the page shows all the data for these plants, including their geographical location, the surrounding suburbs, water service authority, the testing laboratory, and dates testing began in these sites.

SARS-CoV-2 detection and quantitation methodology

The general approach of SARS-CoV-2 detection in wastewater used at all network laboratories is virus concentration, followed by nucleic acid extraction and molecular detection. At the identified wastewater treatment facilities grab or passive samples of influent are collected and transported at <5°C to the testing facility. Table 1 summarises the sample collection, processing and detection methodology used by laboratories involved in the surveillance project. The levels of SARS-CoV-2 in wastewater are reported in copies/mL of wastewater. These values are log-transformed before constructing the graphs.

Table 1. Sampling and methodology used by laboratories involved in the NICD-WRC led COVID-19 wastewater surveillance project.

Name of laboratory	Sampling	Virus concentration	Nucleic acid extraction	Molecular analysis	Molecular analysis platform
National Institute for Communicable Diseases (NICD)	Grab	Ultrafiltration (Centricon® Plus-70 centrifugal ultra-filter device)	QlAamp® viral RNA mini kit	RT-qPCR³ using the Allplex™ 2019-nCoV Assay and the EDX SARS-CoV-2 standard	7500 Real-Time PCR System (Applied Biosystems
GreenHill Laboratories / Praecautio	Grab	Ultrafiltration (Amicon® Ultra-15 Centrifugal Filter Unit)	Omega Bio-Tek Mag- Bind® Viral DNA/RNA 96 Kit	RT-qPCR using the CDC 2019-Novel Coronavirus (2019- nCoV) Real-Time RT-PCR Diagnostic Panel	Rotor-Gene Q (Qiagen)
National Institute for Occupational Health (NIOH)	Grab	Skimmed milk flocculation	MagMAX Viral/ Pathogen Nucleic Acid Isolation Kit	RT-qPCR using the TaqPath COVID-19 CE-IVD RT-PCR Kit (Thermo Fisher)	QuantStudio™ 5 Real-Time PCR System 96-well, 0.1 mL, desktop (Applied Biosystems)
Waterlab/University of Pretoria	Grab	Skimmed milk flocculation	QIAamp® Ultrasens® Virus kit	RT-qPCR using the Allplex [™] 2019-nCoV Assay and the using the 2019_nCoV_N positive control plasmid (Integrated DNA Technologies, Inc, Coralville, IA)	QuantStudio™ 5 Real-Time PCR System (Applied Biosystems)
South African Medical Research Council – Tuberculosis platform (SAMRC-TB Platform)	Grab	None – sample is centrifuged then supernatant analysed	ZymoBiomics RNA Extraction Kit	RT-qPCR³ using the Allplex™ 2019-nCoV Assay and the EDX SARS-CoV-2 standard	QuantStudio 5 (Applied Biosystems)
Lumegen	Passive	Passive sampler and resuspension in phosphate buffered saline	MN DNA/RNA pathogen extraction Kit	RT-qPCR using the TaqPath COVID-19 CE-IVD RT-PCR Kit (Thermo Fisher)	QuantStudio 5 (Applied Biosystems)
Council for Scientific and Industrial Research (CSIR)	Grab	Polyethylene Glycol precipitation	Omega Bio-tek ENZA total RNA Kit II	RT-qPCR using the 2019-nCoV CDC EUA Kit	Qiagen Rotor- Gene 6000 (5-plex) (Qiagen)
Durban University of Technology – Institute of Wastewater Management	Grab	Ultrafiltration (Centricon® Plus-70 centrifugal ultra-filter device)	QIAamp® viral RNA mini kit	RT-ddPCR ^b using CDC 2019-nCoV_N2 Primers, Fam Labelled, double quenched probes	QX200 AutoDG Droplet Digital PCR System (Bio-rad)

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Interpretation of SARS-CoV-2 levels in wastewater

Interpretation of SARS-CoV-2 wastewater levels is evolving. We have elected to use interpretive principles outlined in Table 2 to support public health preparedness and response activities. In general, increasing or decreasing trends in levels are reported based on two or more results, as a single sample that increases or decreases compared with the result from the previous week may represent an outlier. Small changes (up to 0.5 log copies/ml) are not regarded as significant changes unless they form part of a general upward or downward trend. Comparison of results over time when quantification is done by the same laboratory using the same quantitative methodology is meaningful. The use of different methodologies by different laboratories precludes comparison of quantitative results across laboratories.

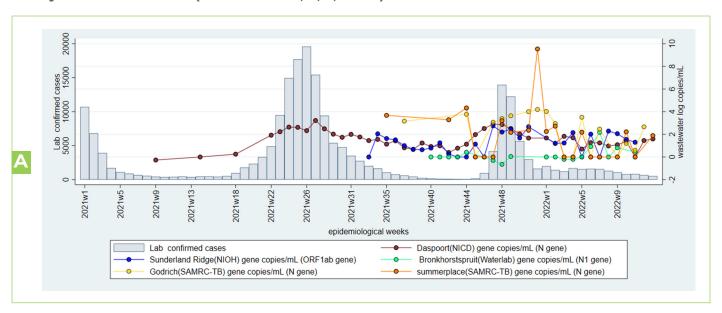
Table 2. Principles of SARS-CoV-2 detection and quantification on influent samples from wastewater treatment plants and interpretive principles to guide application of test results to support COVID-19 public health responses, South Africa.

Testing modality	Interpretive principles to support public health responses
Detection of SARS-CoV-2	 When a test result changes from positive to negative, this signifies fewer/no cases in population negative to positive, this indicates the need for increased population awareness and action Qualitative results (presence or absence) are comparable between laboratories
Quantification of SARS- CoV-2	*The concentration of SARS-CoV-2 at a particular facility may be used to infer the burden of SARS-CoV-2 in the population served by the wastewater treatment facility. *Changes in the concentration of SARS-CoV-2 give an indication of whether the burden of disease is increasing or decreasing *Quantitative results between laboratories are not comparable. *Quantitative results should be interpreted for a single wastewater treatment plant tested by the same laboratory using the same methodology over time

RESULTS

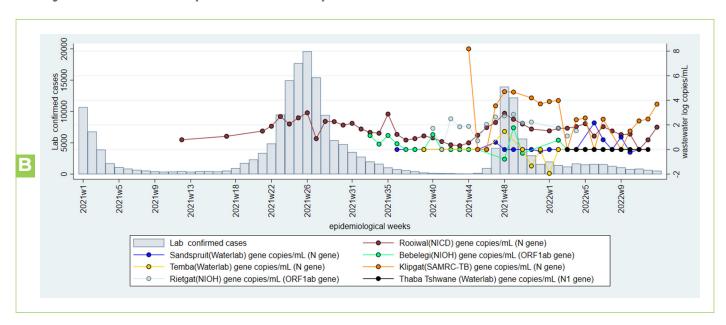
Gauteng Province

A: City of Tshwane South (sub-districts 3, 4, 6, and 7)



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B: City of Tshwane North (sub-districts 1 & 2)



C: City of Tshwane North (sub-district 5)

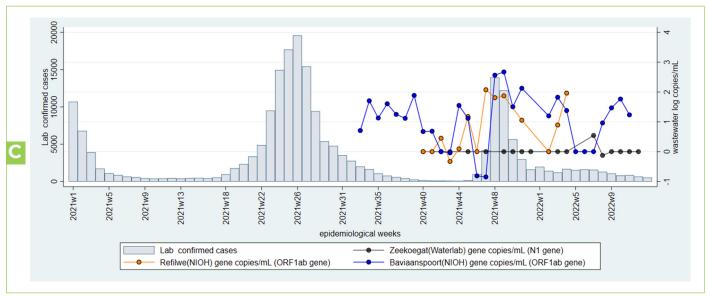


Figure 1 A-C. Laboratory confirmed cases of SARS-CoV-2 (bars) and levels of SARS-CoV-2 in log copies/ml of wastewater (coloured lines) for selected wastewater treatment plants (WWTP) and metropolitan areas in Tshwane District Municipality (Tshwane South and North), Gauteng Province during epidemiological weeks 1 of 2021 to week 13 of 2022. The testing laboratory and quantified SARS-CoV-2 gene is named in brackets after the name of the WWTP. Note that comparisons of levels over time should only be done for specimens tested in the same laboratory.

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D: City of Johannesburg Metropolitan Municipality

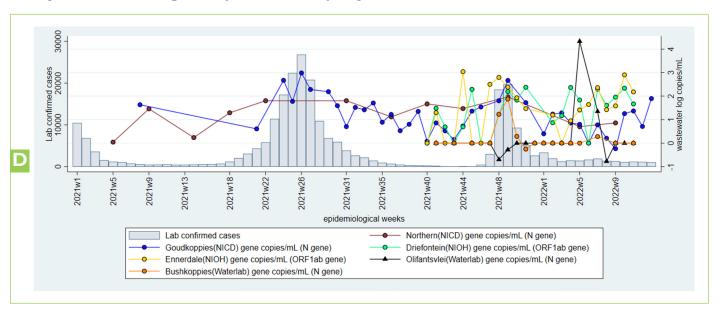
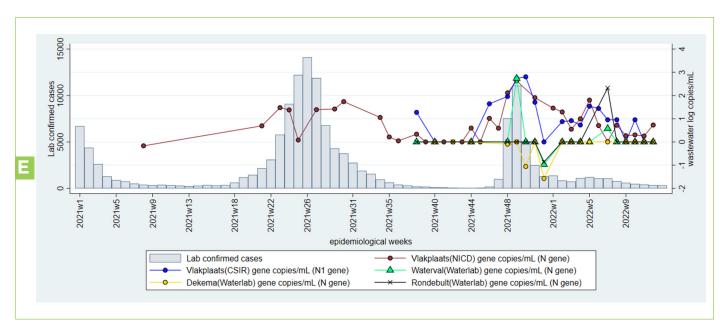


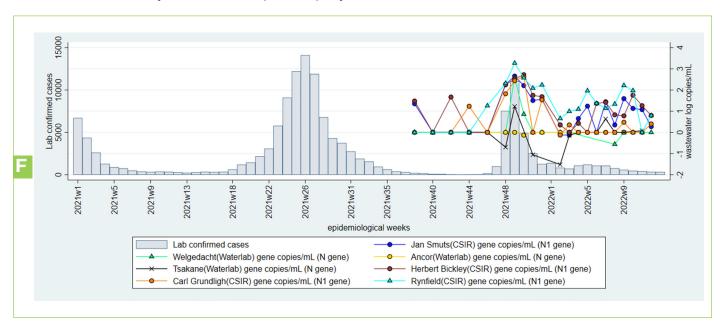
Figure 1 D. Laboratory confirmed cases of SARS-CoV-2 (bars) and levels of SARS-CoV-2 in log copies/ml of wastewater (coloured lines) for selected wastewater treatment plants (WWTPs) in the City of Johannesburg Metropolitan Municipality, Gauteng Province during epidemiological weeks 1 of 2021 to week 13 of 2022. The testing laboratory and quantified SARS-CoV-2 gene are named in brackets after the name of the WWTP. Note that comparisons of levels overtime should only be made for specimens tested in the same laboratory.

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E: Ekurhuleni East (sub-districts D, E or E1, E2)



F: Ekurhuleni South (sub-districts A, F or S1, S2)



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G: Ekurhuleni North (sub-districts B, C or N1,N2)

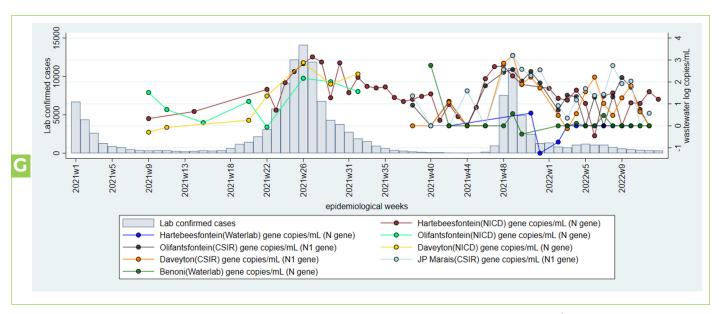


Figure 1 E-G. Laboratory confirmed cases of SARS-CoV-2 (bars) and levels of SARS-CoV-2 in log copies/ml of wastewater (coloured lines) for selected wastewater treatment plants (WWTP) in Ekurhuleni Metropolitan Municipality, Gauteng Province during epidemiological weeks 1 of 2021 to week 13 of 2022. The testing laboratory and quantified SARS-CoV-2 gene are named in brackets after the name of the WWTP. Note that comparisons of levels over time should only be made for specimens tested in the same laboratory.

H: West Rand District Municipality

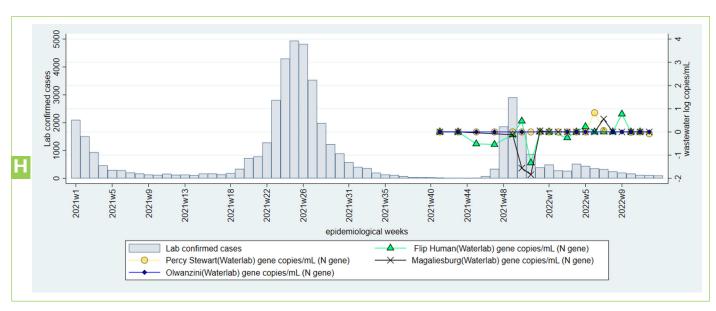


Figure 1 H. Laboratory confirmed cases of SARS-CoV-2 (bars) and levels of SARS-CoV-2 in log copies/ml of wastewater (coloured lines) for selected wastewater treatment plants (WWTP) in West Rand District Municipality, Gauteng Province during epidemiological weeks 1 of 2021 to week 7 of 2022. The testing laboratory and quantified SARS-CoV-2 gene are named in brackets after the name of the WWTP. Note that comparisons of levels over time should only be made for specimens tested in the same laboratory.

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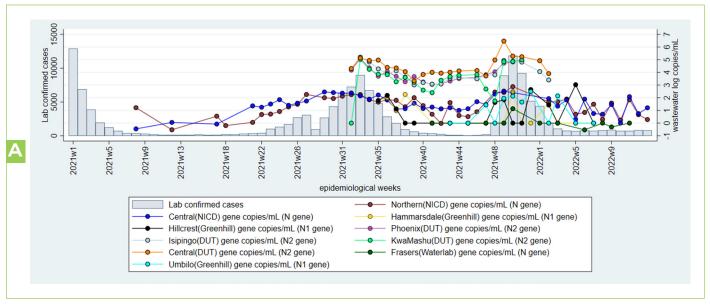
In Tshwane South (sub-district 3, 4, 6 & 7), increases by one week were observed at Godrich (in week 12) and summerplace WWTPs. The levels at Godrich WWTP has remained stable at almost 2log copies/ml for two weeks. In Tshwane North (sub-district 1& 2), four successive increases were observed at Klipgat WWTP, with the current levels at 4 log copies/ml of wastewater.

In the city of Johannesburg, the SARS-CoV-2 levels in Goudkoppies have increased to up to 2 log copies/ml. In Ekurhuleni almost all the WWTPs continue to decrease with corresponding decrease in clinical cases. However, marginal increases in SARS-CoV-2 levels were observed in Vlakplaats at Ekurhuleni South, in week 12.

The public health authorities should continue to strengthen surveillance for clinical cases in these areas, promote vaccination and non-pharmaceutical interventions in all areas.

KwaZulu-Natal Province

2A: eThekwini Metropolitan Municipality



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B: uMgungundlovu District Municipality

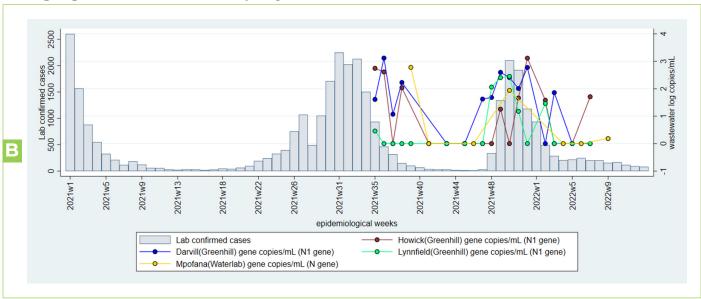


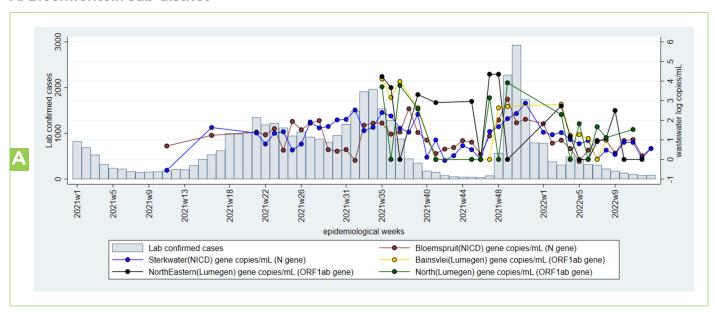
Figure 2A-B. Laboratory confirmed cases of SARS-CoV-2 (bars) and levels of SARS-CoV-2 in log copies/ml of wastewater (coloured lines) from wastewater treatment plants (WWTP) in eThekwini, (A-B) and uMgungundlovu Metro (C), Kwa-Zulu Natal Province during epidemiological weeks 1-51, 2021 and week 13, 2022. The testing laboratory and quantified SARS-CoV-2 gene are named in brackets after the name of the WWTP. Note that comparisons of levels over time should only be made for specimens tested in the same laboratory.

The SARS-CoV-2 levels have declined in all the plants with recent results in eThekwini and uMgungundlovu, corresponding to the decrease in clinical cases. A marginal increase was observed at Central WWTP in eThekwini, requiring more samples. We continue to Therefore, the authorities should continue to promote vaccination and non-pharmaceutical interventions in all areas.

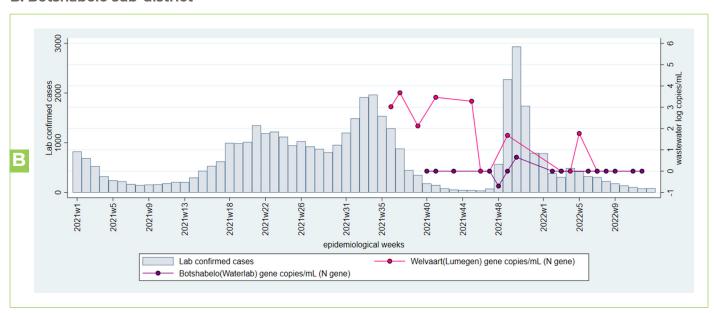
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Free State Province- Mangaung

A: Bloemfontein sub-district



B. Botshabelo sub-district



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C. Neledi & ThabaNchu sub-districts

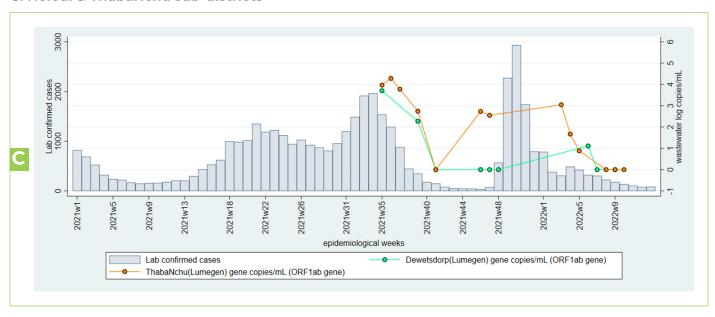


Figure 3 A-C. Laboratory confirmed cases of SARS-CoV-2 (bars) and levels of SARS-CoV-2 in log copies/ml of wastewater (coloured lines) from wastewater treatment plants (WWTPs) in Mangaung, Free State Province (Bloemfontein, Botshabelo, Naledi and ThabaNchu) during epidemiological weeks 1, 2021 to 13, 2022. The testing laboratory and quantified SARS-CoV-2 gene are named in brackets after the name of the WWTP. Note that comparisons of levels over time should only be made for specimens tested in the same laboratory.

The recent SARS-CoV-2 levels in majority of the plants in the three sub-districts in Free State continue to decline or remain stable at low levels, corresponding to the decreases in clinical cases. However, marginal increases were observed at Sterkwater and Bloemspruit WWTPs in Bloemfontein sub-district. These increases have no significance yet, until we receive subsequent samples. The public health authorities should continue surveillance for cases, promote vaccination and non-pharmaceutical interventions.

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Eastern Cape Province

A: Nelson Mandela Metropolitan Municipality

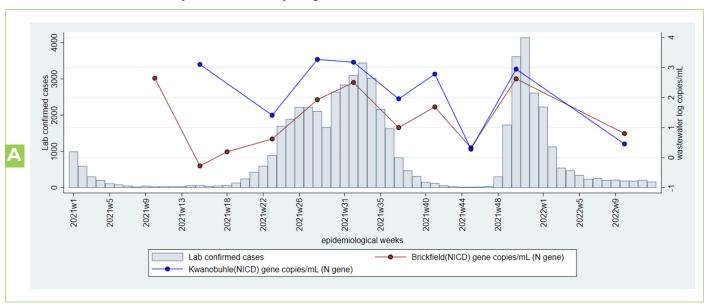


Figure 4A. Laboratory confirmed cases of SARS-CoV-2 (bars) and levels of SARS-CoV-2 in log copies/ml of wastewater (coloured lines) from wastewater treatment plants (WWTPs) in Nelson Mandela Metro, Eastern Cape Province during epidemiological weeks 1, 2021 to 10, 2022. The testing laboratory and quantified SARS-CoV-2 gene are named in brackets after the name of the WWTP. Note that comparisons of levels over time should only be made for specimens tested in the same laboratory

B: Buffalo City Metropolitan Municipality

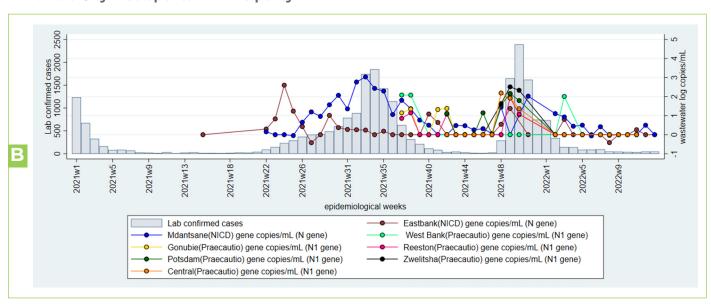


Figure 4B. Laboratory confirmed cases of SARS-CoV-2 (bars) and levels of SARS-CoV-2 in log copies/ml of wastewater (coloured lines) from wastewater treatment plants (WWTPs) in Nelson Mandela Metro, Eastern Cape Province during epidemiological weeks 1, 2021 to 13, 2022. The testing laboratory and quantified SARS-CoV-2 gene are named in brackets after the name of the WWTP. Note that comparisons of levels over time should only be made for specimens tested in the same laboratory.

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In both Nelson Mandela and Buffalo City Metros, the results show that SARS-CoV-2 levels in all WWTPs have steadily decreased and remained at zero levels, corresponding to the decline in clinical cases. The public health authorities should continue to promote vaccination and non-pharmaceutical interventions. In Nelson Mandela Metro, assessment cannot be made due to absence of recent results. Readers are referred to the SAMRC wastewater dashboard for more in-depth data regarding levels of SARS-CoV-2 in wastewater plants in Nelson Mandela Metro (https://www.samrc.ac.za/wbe/).

Western Cape Province

City of Cape Town:

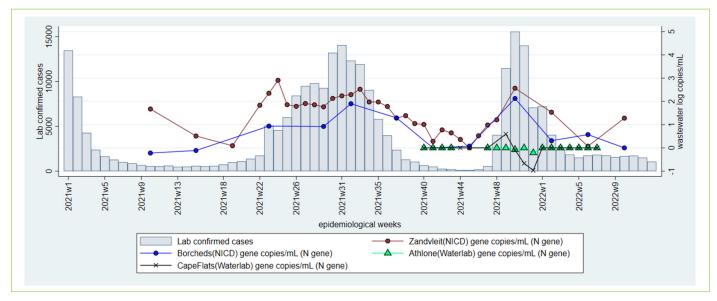


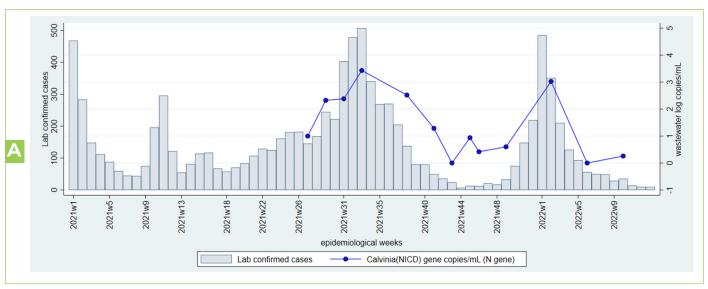
Figure 5. Laboratory confirmed cases of SARS-CoV-2 (bars) and levels of SARS-CoV-2 in log copies/ml of wastewater (coloured lines) from wastewater treatment plants (WWTPs) in the City of Cape Town, Western Cape Province during epidemiological weeks 1, 2021 to 10, 2022. The testing laboratory and quantified SARS-CoV-2 gene are named in brackets after the name of the WWTP.

While the SARS-CoV-2 levels in Borcherds Quarry remain low corresponding to the decline in clinical case load, a marginal increase in the levels in Zandvleit was observed in week 10. Readers are referred to the SAMRC website, which provides data from additional wastewater treatment plants in the City of Cape Town and other Western Cape districts (https://www.samrc.ac.za/wbe/) to contextualise the results. The public health authorities should continue surveillance for cases, promote vaccination and non-pharmaceutical interventions.

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Northern Cape Province

A: Namakwa District Municipality



B: Frances Baard District Municipality

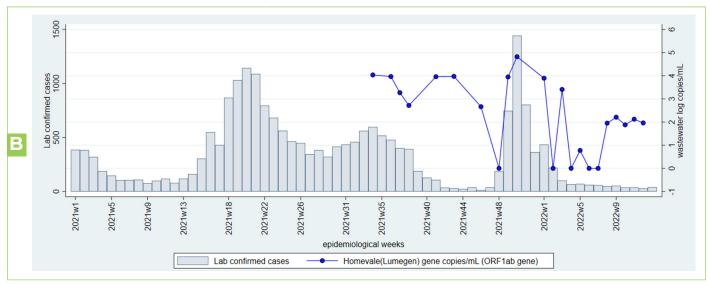


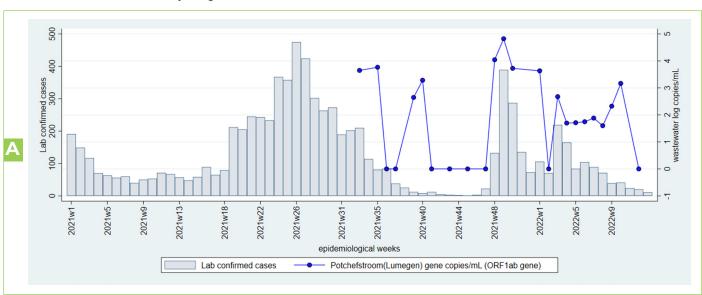
Figure 6A-B. Laboratory confirmed cases of SARS-CoV-2 (bars) and levels of SARS-CoV-2 in log copies/ml of wastewater (coloured lines) in wastewater treatment plants (WWTPs) from Calvinia in Namakwa Metro (a) and Kimberly in Frances Baard District (b), Northern Cape Province during epidemiological weeks 1, 2021 to week 10, 2022. The testing laboratory and quantified SARS-CoV-2 gene are named in brackets after the name of the WWTP. Note that comparisons of levels over time should only be made for specimens tested in the same laboratory.

In Homevale WWTP in France Baard subdistrict, the SARS-CoV-2 levels have been stable at 2 log copies/ml between weeks 9 and 12. More recent results are required. On the other hand, a current assessment cannot be made for France Baard due to the absence of recent results. The public health authorities should continue surveillance for cases, promote vaccination and non-pharmaceutical interventions.

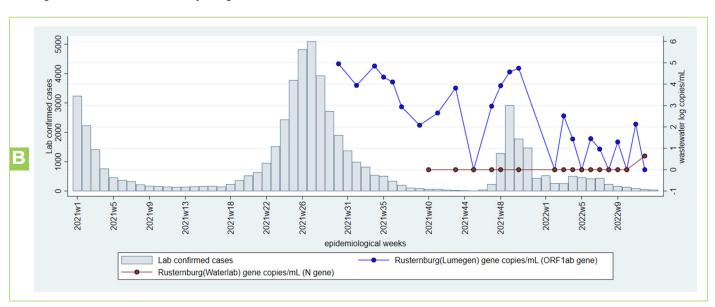
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North West Province

A: JB Marks Local Municipality



B: Bojanala District Municipality



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C: City of Matlosana Municipality

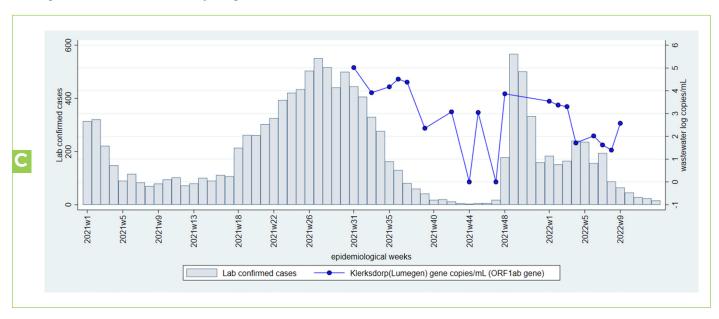


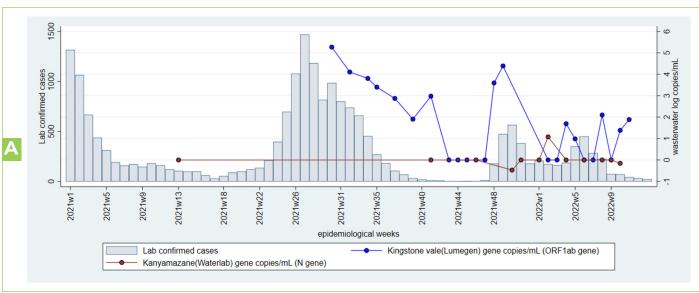
Figure 7A-C. Laboratory confirmed cases of SARS-CoV-2 (bars) and levels of SARS-CoV-2 in log copies/ml of wastewater (coloured lines) in wastewater treatment plants (WWTPs) from Potchefstroom, JB Marks District (A) Rustenberg, Bojanala District (B), and City of Matlosana, Northwest Province during epidemiological weeks 1, 2021 to 10, 2022. The testing laboratory and quantified SARS-CoV-2 gene are named in brackets after the name of the WWTP. Note that comparisons of levels over time should only be made for specimens tested in the same laboratory.

The SARS-CoV-2 levels are decreasing in all plants in the NorthWest province corresponding to the decline in clinical case. However, more recent and consistent results are required in this area. The public health authorities should continue surveillance for cases, promote vaccination and non-pharmaceutical interventions.

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Mpumalanga Province

A: Mbombela Local Municipality



B: Emalahleni Local Municipality

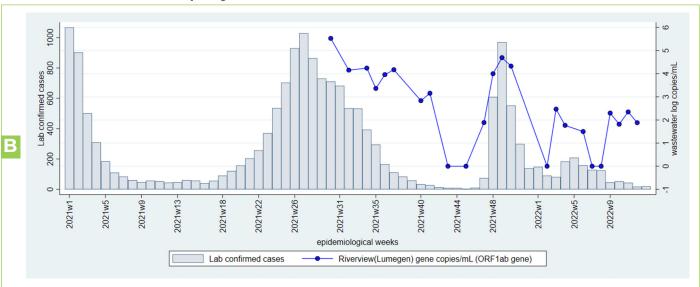


Figure 8A-B. Laboratory confirmed cases of SARS-CoV-2 (bars) and levels of SARS-CoV-2 in log copies/ml of wastewater (coloured lines) in wastewater treatment plants (WWTPs) from Mbombela and Emalahleni Local Municipality, Mpumalanga Province during epidemiological weeks 1, 2021 to 9, 2022. The testing laboratory and quantified SARS-CoV-2 gene are named in brackets after the name of the WWTP. Note that comparisons of levels over time should only be made for specimens tested in the same laboratory.

The SARS-CoV-2 levels in Mbombela (Kingstonvale) and Emalahleni (Riverview) were at 2 log copies/ml in weeks 11 and 12 respectively. More recent and consistent results are required in this area The public health authorities should continue surveillance for cases, promote vaccination and non-pharmaceutical interventions.

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Limpopo Province

Polokwane Local Municipality

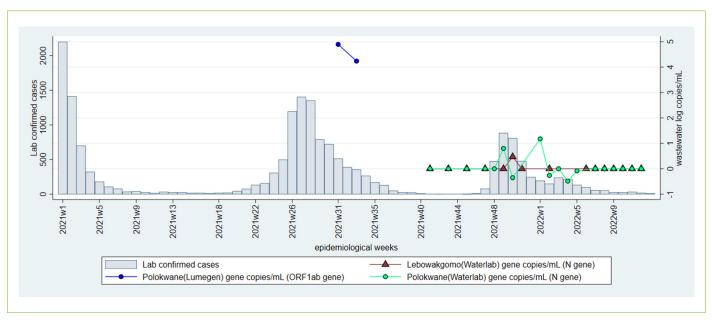


Figure 9. Laboratory confirmed cases of SARS-CoV-2 (bars) and levels of SARS-CoV-2 in log copies/ml of wastewater (coloured lines) in wastewater treatment plants (WWTPs) from Polokwane Local Municipality, Limpopo Province during epidemiological weeks 31-33, 2021.

The levels are low in Polokwane corresponding to low number of clinical cases.

LIMITATIONS

It is not possible to estimate population burden of disease using wastewater testing of SARS-CoV-2 as sources of variability are multiple, including variation in length and concentration of SARS-CoV-2 excretion by infected persons, variation in degradation rate of viral RNA in wastewater and sampling error. Interpretation of results from quantitative testing of SARS-CoV-2 in wastewater is enhanced when the population served by the wastewater treatment plants is well characterised in terms of SARS-CoV-2 testing rates, health seeking behaviour, hospital admissions and deaths due to SARS-CoV-2, as well as other general indicators of health. Further exploration of the relationship between quantitative SARS-CoV-2 results, local trends in clinical case burden, environmental factors, and test methodology will support the interpretation of observed fluctuations in RNA levels. Quality assessment and inter-laboratory comparisons are underway to ensure participating laboratories are providing consistent and comparable results.

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PART 2: RESULTS FROM SEQUENCING OF SARS-COV-2 RNA FRAGMENTS IN WASTEWATER

*note this report is unchanged since week ending 1 April 2022.

SARS-CoV-2 is shed into stools and is detectable in wastewater (sewage systems). The South African Collaborative COVID-19 Environmental Surveillance System (SACCESS) detects and quantifies SARS-CoV-2 from wastewater and used these data as a monitoring system for the SARS-CoV-2 epidemic in South Africa. Weekly reports of these data published at the NICD website: (https://www.nicd.ac.za/diseases-a-z-index/disease-index-covid-19/surveillance-reports/weekly-reports/wastewater-based-epidemiology-for-sars-cov-2-in-south-africa/).

The gold standard method to diagnose COVID-19 infection in clinical samples is reverse-transcriptase polymerase chain reaction (RT-PCR). The test, along with SARS-CoV-2 antigen detection has been used to monitor the epidemiology of the pandemic in South Africa. Epidemiological approaches based on these methods are limited by health seeking behaviour (only symptomatic patients go for testing) and testing practices of health care workers Therefore, this approach does not identify asymptomatically infected persons. Asymptomatic individuals may, however, be driving the force of infection and transmission in the community. Testing of wastewater for SARS-CoV-2 levels overcomes this limitation by allowing population levels of SARS-CoV-2 to be monitored over time, thus adding value to our understanding of SARS-CoV-2 epidemiology of the pandemic.

SARS-CoV-2 has been classified into different variants, that are continually emerging as a result of viral evolution. These variants acquire or lose mutations coding for various epitopes found on key viral proteins which lead to changes in transmissibility dynamics, response to treatment or ability to evade neutralisation by antibodies. WHO classified SARS-CoV-2 variants into variants of concerns (VOCs) and variants of interest (VOIs). VOCs have included Alpha, Beta, Delta, and Gamma, and recently Omicron. Of these, Beta and Omicron were first reported in South Africa. VOIs include Lambda and Mu (https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/).

The Network for Genomics Surveillance of South Africa (NGS-SA) monitors the epidemiology of SARS-CoV-2 variants in PCR-confirmed cases in South Africa. In clinical cases, variant detection is performed using whole genome sequencing and other methods such as real-time PCR. During the first wave (June to August 2020), the Wuhan SARS-CoV-2 strain dominated amongst clinical cases while in the second wave (November 2020 to February 2021), the Beta variant was discovered and was predominant. The third wave (May to September 2021) was characterized by the dominance of the Delta variant and the fourth wave (November 2021 to January 2022) by the Omicron variant.

Several groups have sequenced SARS-CoV-2 from wastewater including groups in the Netherlands who generated near whole genome sequence from wastewater (Lara et al., 2020). In the United States, wastewater sequencing provided comparable results to clinical testing and evidenced new mutations before they appeared in clinical samples (Crits-Christoph et al., 2021).

Here, we report on SARS-CoV-2 sequences and variants of concern present in wastewater samples collected at sentinel wastewater treatment plants in South African urban metros from week 14 in 2021 to week 7 of 2022.

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METHODS

Wastewater sites

In 2020, the National Institute for Communicable Diseases commenced testing of influent wastewater samples for SARS-CoV-2 from 15 wastewater treatment plants in metropolitan areas, including five in Gauteng Province, four in Eastern Cape province, two in the City of Cape Town (Western Cape Province), two in Mangaung (Free State Province), two in eThekwini (KwaZulu- Natal Province) (Table 1).

Table 1: Characteristics of wastewater treatment facilities and of samples submitted for SARS-CoV-2 sequencing from these sites, 2021-2022

Province	Metro or District	Plant name	Population size served by the facility		Genomic testing	
				Epidemiological week when sequencing started in 2021	# samples submitted for sequencing	# samples yielding good quality sequences
Eastern Cape			141000			
		Mdantsane	112900	25	24	
	Nelson Mandela Metro		40000			
		KwaNobuhle	100320	15	7	4
Free State	Mangaung		200000		33	
			350000			
Gauteng			100000			
			1200000			
		Goudkoppies	500000			
					33	
		Daspoort				
Kwazulu-Natal						
			350000			
Western Cape	City of Cape Town Metro		380000			
		Zandvliet	460000		25	12

Sample collection, RNA extraction, amplification and sequencing

One litre of grab sewage samples were collected and transported at 4°C. Viruses were concentrated from the sample by ultrafiltration (Ikner, Soto-Beltran and Bright, 2011), and RNA was extracted using the QIAamp Viral RNA kit (Qiagen, GmbH, Germany). SARS-CoV-2 was detected by RT-PCR using AllplexTM 2019- nCoV Assay from Seegene kit (Seoul, Korea). RNA was re-extracted from SARS-CoV-2 positive concentrates and subjected to amplicon-based whole genome sequencing using the Sinai protocol with some modifications (Gonzalez-Reiche et al., 2020). Libraries were prepared using the COVIDSeq Kit (Illumina Inc, USA), and sequencing was performed using Illumina COVIDSeq kits as described in (Bhoyar et al., 2021) at the Sequencing Core Facility at the NICD.

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Sequence analysis

The Galaxy pipeline (RC, 2005) was used for sequence analysis. Reads were trimmed and filtered according to published criteria (Khailany, Safdar and Ozaslan, 2020). At least 10 reads required at each nucleotide position for downstream analysis. Mutations present at 10% or less were removed from the analysis. Reads were mapped against the reference genome (Wuhan strain) and amino acid variation was analysed. Table 2 illustrates an example of amino acids variation file (https://usegalaxy.eu/).

Table 2: Illustration of amino acids variations (Galaxy: https://usegalaxy.eu/). A shows sample ID. B is QC filter, which is quality indicator. C is the number of reads produced for each sample. D is the effect of the mutation detected in the gene. E is the name of the gene where mutation occurred. F is the mutation detected. G is the frequency of the reads in the mutation

A	В	С	D	E	F	G
Sample	QC filtre	Number of reads	Mutation effect	Gene	Mutation	Frequency of mutations
ENV-COV-21-285_S337_001.fastq	PASS	12	NON_SYNONYMOUS_CODING	ORF1ab	K790Q	0.833333
ENV-COV-21-285_S337_001.fastq	PASS	644	NON_SYNONYMOUS_CODING	ORF1ab	K798N	0.057453
ENV-COV-21-285_S337_001.fastq	PASS	14	NON_SYNONYMOUS_CODING	ORF1ab	F800L	0.857143
ENV-COV-21-285_S337_001.fastq	PASS	44	SYNONYMOUS_CODING	ORF1ab	G45	0.863636
ENV-COV-21-285_S337_001.fastq	min_af_0.05Xmin_dp_1Xmin_dp_alt_10	44	FRAME_SHIFT	ORF1ab	Y46L?	0.045455
ENV-COV-21-285_S337_001.fastq	PASS	1347	NON_SYNONYMOUS_CODING	ORF1ab	T54P	0.123979
ENV-COV-21-285_S337_001.fastq	PASS	153	SYNONYMOUS_CODING	ORF1ab	T54	0.078431

SARS-CoV-2 in the sewage system is fragmented, and therefore, generation of a consensus sequence for each sample is not meaningful. Therefore, to identify variants at each geographic location, we analysed amino acid variation in each individual sample. For each VOC or VOI, unique single nucleotide polymorphisms were identified by comparing the new lineage with the Wuhan strain in a public database (https://outbreak.info/). Using the amino acid variation data file, we used STATA software (v 17.1) (https://www.stata.com/) to collate s-gene mutations in a matrix such that the columns represented the amino acid positions of the spike protein and each row recorded all mutations detected in each sample at every locus across the spike gene. We included all mutations, including low frequency mutations and recorded the proportion of reads where that mutation was detected (the 'read frequency') as a percentage of total reads. Using the list of unique mutations for each VOC and VOI in the spike protein region (Table 3) we interrogated the matrix for the presence or absence of known signature mutations in each sample using STATA software (Table 3). As new variants were detected and identified in clinical specimens, we added signature mutations to the STATA code, allowing us to identify the presence of new variants both retrospectively and prospectively

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Table 3: List of signature mutations which was used to identify VOC and VOI present in wastewater samples from week 14 in 2021 to week 7 of 2022

Omicron	Alpha	Beta	Delta	C.1.2	Gamma	Lambda	Mu
G339D	A570D	D80A	 T19R	P9L	T20N		
S371L	S982A		R145H	P25L	P26S		Y145N
S375F	D1118H		E156del	C136F	T1027I	D253N	
Q493R			R158G	Y449H		L452Q	
G496S			A222V			F490S	
Y505H							
N856K							
Q954H							
N969K							
L981F							

Results and discussion

Up to the 15th February 2022, a total of 315 wastewater samples from sites listed in Table 1 underwent RNA extraction, amplification and sequencing. Of these 315 samples, 160 (50.8%) yielded SARS-CoV-2 RNA sequences.

Detection of SARS-CoV-2 variants from wastewater samples using signature mutations analysis Gauteng province

In the Gauteng province, 60 samples yielded sequencing results displayed in figure 1, which illustrates how beta variant was present in north and southern Gauteng province in week 21-22, but was replaced by delta variant across the province from weeks 21 until 32. Two sub-lineages of delta and a variant present only in South Africa. C.1.2 were simultaneously present at all sites with variable but lower read frequencies. During the interwave period (weeks 34-44) most samples submitted for sequencing failed to yield good quality sequence data, most likely on account of low or absent SARS-CoV-2 RNA fragments. Omicron variant was first detected in week 46 and by week 47 was found to be present at all plants across the province. In all plants, the mutation formerly associated with C1.2 was also found to be present at high read frequencies simultaneously with Omicron variant. Omicron variant continues to be present up to week 7 of 2022.

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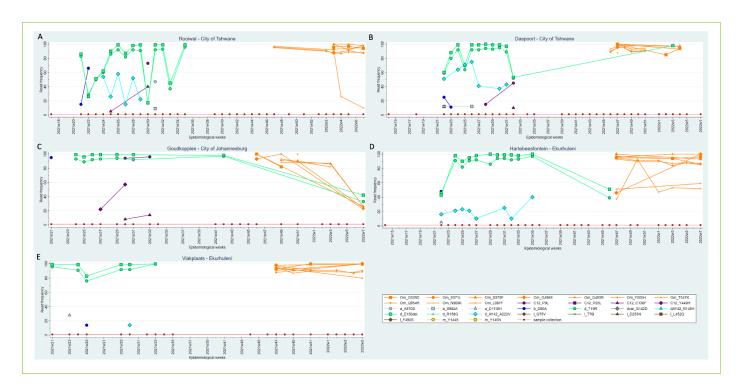


Figure 1: Beta mutation (D80A, dark blue circle). Delta mutations: T19R (green circle), G142D (pale blue diamond), R145H (green triangle), E156G (plus symbol), DEL157/158 (green square), A222V (pale blue diamond). C.1.2 mutations: P9L (purple circle), P25L (purple square), C136F (purple triangle), Y449H purple diamond. Omicron mutations: G339D (black square), S371L (black circle), S375F (black triangle), G496S (black diamond), Q493R (black dot), T547K (black small triangle), Q954H (black small diamond), N969K (black small line), L981F (black small x). Dots on the red line shows the timepoints of sample collection, absence of specific coloured lines means the mutation was not detected at that timepoint. A (Rooiwal plant), B (Daspoort plant), C (Goudkoppies plnat), D (Hartebeesfontein plant), and E (Vlakplaats plant).

KwaZulu- Natal province

In KwaZulu-Natal province, 37 samples yielded good sequences and were included in figure 2. Beta variant was detected in a single sample from Central plant in week 23. Delta variant was present in Northern and Central plants from weeks 21 to 35, and was last detected in week 43 at Central plant. As in Gauteng Province, two sub-lineages of delta and a variant present only in South Africa, C.1.2 were simultaneously present at all sites with variable but lower read frequencies from weeks 21-35. During the interwave period (weeks 34-44) most samples submitted for sequencing failed to yield good quality sequence data, most likely on account of low or absent SARS-CoV-2 RNA fragments. Omicron variant was first detected in week 49 in both plants, along with the mutation formerly associated with C1.2. Omicron variant continues to be present up to week 2 of 2022.

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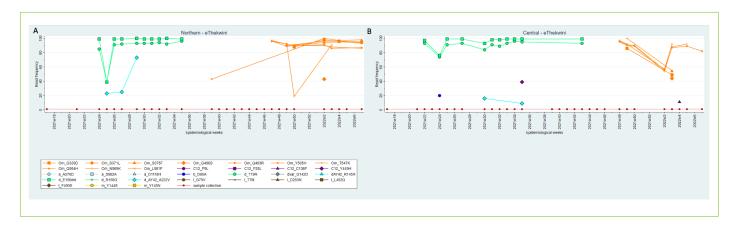


Figure 2: Beta mutation (D80A, dark blue circle). Delta mutations: T19R (green circle), G142D (pale blue diamond), R145H (green triangle), E156G (plus symbol), DEL157/158 (green square), A222V (pale blue diamond). C.1.2 mutations: P9L (purple circle), P25L (purple square), C136F (purple triangle), Y449H purple diamond. Omicron mutations: G339D (black square), S371L (black circle), S375F (black triangle), G496S (black diamond), Q493R (black dot), T547K (black small triangle), Q954H (black small diamond), N969K (black small line), L981F (black small x). Dots on the red line shows the timepoints of sample collection, absence of specific coloured lines means the mutation was not detected at that timepoint. A (Northern eThekwini plant) and B (Central eThekwini plant).

Free State province

In Mangaung, Free State province, 34 samples yielded sequencing results displayed in figure 3. The beta variant was present until week 23 (Sterkwater plant) and 25 (Bloemspruit) when it was co-detected with delta variant. As in Gauteng, delta variant dominated from weeks 23-24, along with two delta sublineanges. Variant C.1.2 was detected in week 31 in Blomespruit plant. No samples yield quality sequence data from weeks 35-46. Omicron was first detected in week 50 at both plants, along with variant C.1. 2.

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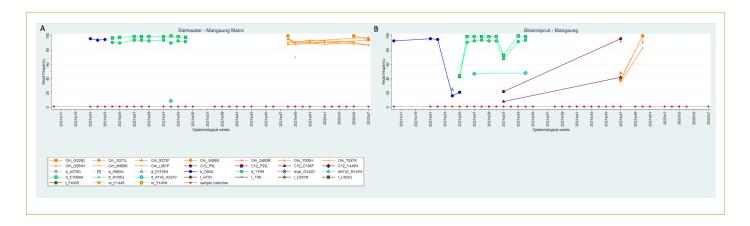


Figure 3: Beta mutation (D80A, dark blue circle). Delta mutations: T19R (green circle), G142D (pale blue diamond), R145H (green triangle), E156G (plus symbol), DEL157/158 (green square), A222V (pale blue diamond). C.1.2 mutations: P9L (purple circle), P25L (purple square), C136F (purple triangle), Y449H purple diamond. Omicron mutations: G339D (black square), S371L (black circle), S375F (black triangle), G496S (black diamond), Q493R (black dot), T547K (black small triangle), Q954H (black small diamond), N969K (black small line), L981F (black small x). Dots on the red line shows the timepoints of sample collection, absence of specific coloured lines means the mutation was not detected at that timepoint. A (Sterkwater plant) and B (Bloemspruit plant).

Western Cape province

In the Western Cape Province, 13 samples yielded sequencing results displayed in figure 4. At Zandvliet plant, delta variant along with subvariants described above were detected from week 25-35. Evidence of C1.2 was found in week 22. No samples yield quality sequence data from week 38 to week 46. In week 47, omicron and variant C.1.2 was detected. A single mutation associated with omicron was observed in week 24, without co-detection of other mutations associated with omicron. At Borcherd's Quarry, the delta variants and subvariant were detected in week 32. No samples yield quality sequence data from week 34, 2021 to week 2, 2022.

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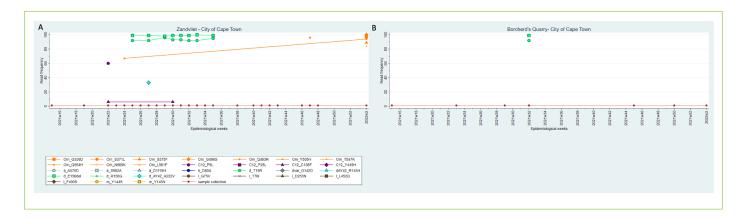


Figure 4: Beta mutation (D80A, dark blue circle). Delta mutations: T19R (green circle), G142D (pale blue diamond), R145H (green triangle), E156G (plus symbol), DEL157/158 (green square), A222V (pale blue diamond). C.1.2 mutations: P9L (purple circle), P25L (purple square), C136F (purple triangle), Y449H purple diamond. Omicron mutations: G339D (black square), S371L (black circle), S375F (black triangle), G496S (black diamond), Q493R (black dot), T547K (black small triangle), Q954H (black small diamond), N969K (black small line), L981F (black small x). Dots on the red line shows the timepoints of sample collection, absence of specific coloured lines means the mutation was not detected at that timepoint. A (Zandvliet plant) and B (Borcherd's Quarry plant).

Eastern Cape province

In the Eastern Cape Province, 16 samples yielded sequencing results displayed in figure 5. The delta variant and subvariant were detected in week 30 until week 35 (Mdantsane plant), week 24 until 29 (Brickfield plant), week 28 until 32 (Brickfield) and week 28 until 41 (Kwanobuhle). Omicron variant was first detected in week 46 and by week 48 at the Mdantsane plant, alongside C1.2 and 3 other omicron subvariants were detected in week 2, 2022. Similarly, omicron and C1.2. variants were detected in week 50 at the Kwanobuhle and Brickfield plant. No omicron and C1.2. variants were detected at the Eastbank plant.

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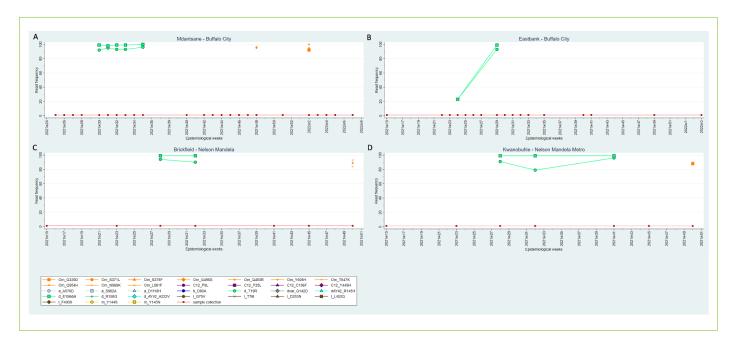


Figure 5: Beta mutation (D80A, dark blue circle). Delta mutations: T19R (green circle), G142D (pale blue diamond), R145H (green triangle), E156G (plus symbol), DEL157/158 (green square), A222V (pale blue diamond). C.1.2 mutations: P9L (purple circle), P25L (purple square), C136F (purple triangle), Y449H purple diamond. Omicron mutations: G339D (black square), S371L (black circle), S375F (black triangle), G496S (black diamond), Q493R (black dot), T547K (black small triangle), Q954H (black small diamond), N969K (black small line), L981F (black small x). Dots on the red line shows the timepoints of sample collection, absence of specific coloured lines means the mutation was not detected at that timepoint. A (Mdantsane plant), B (Eastbank plant) C (Brickfield plant) and D (Kwanobuhle plant).

The distribution of SARS-CoV-2 from wastewater has progressed from the predominance of beta variant in January 2021, to delta variant dominance (June 2021) to Omicron and Cl.2 in early 2022 (Figure 6).

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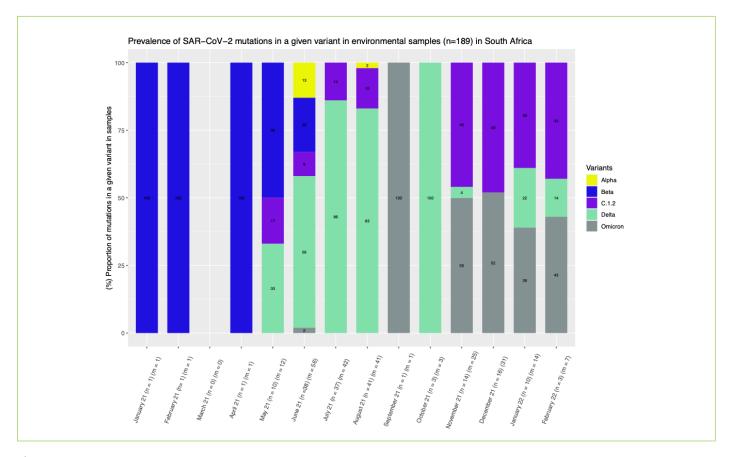


Figure 6: The proportion of mutations in a given variant in the environmental samples by month and year (January 2021-February 2022) from all South African provinces. The number of samples processed each month are indicated as n and the number of mutations present in a sample are represented as m i.e. n = 10 indicates that mutations were successfully detected 10 sample in May, 2021 and m = 12 indicates that 12 mutations were found in which 13% of the 12, are mutations in the alpha variant.

CONCLUSION

Sequencing of SARS-CoV-2 fragments from wastewater is able to identify the presence of variants in known to be present in clinical samples. These data show the presence of omicron variant in all recent samples across South Africa (up to week 7). The significance of the mutation associated with the C1.2 variant (P25L) is presently unclear as other mutations associated with this variant are not present, nor is the mutation found in omicron VOC. These results must be read along with the reports generated by the Centre for Respiratory Diseases and Meningitis found at (https://www.nicd.ac.za/wp-content/uploads/2022/03/Update-of-SA-sequencing-data-from-GISAID-18-Mar-2022_2.pdf).

Sequencing of wastewater has limitations. The ability to identify variants in wastewater relies on the identification of single nucleotide polymorphisms found in clinical strains and which are uniquely associated with these variants. We are not yet able to detect new variants. Sequencing of SARS-CoV-2 from wastewater may not yield good quality sequence data when viral concentration in wastewater is low.

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CONCLUSION

SARS-CoV-2 data from wastewater at South African sentinel sites show concordance with clinical, epidemiologic curves and sequencing data (not shown) in the respective locations, illustrating the potential of the SACCESS network to provide descriptive epidemiological data pertaining to geographic variation, burden and variants of SARS-CoV-2.

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ACKNOWLEDGEMENTS

- The contributions of local government and wastewater treatment staff to sample collection and transport is acknowledged and appreciated.
- Students support with sample collections and processing the samples: Mr Thoriso Mooa, Ms Unarine Matodzi, Ms Phiwinhlanhla Nkosi SAMRC-TB Platform
- The Water Research Commission is thanked for their vision and support.
- The NICD SARS-CoV-2 epidemiology and IT team members are thanked for sharing district and sub-district case burdens in order to generate graphs. These team members include Andronica Moipone Shonhiwa, Genevie Ntshoe, Joy Ebonwu, Lactatia Motsuku, Liliwe Shuping, Mazvita Muchengeti, Jackie Kleynhans, Gillian Hunt, Victor Odhiambo Olago, Husna Ismail, Nevashan Govender, Ann Mathews, Vivien Essel, Veerle Msimang, Tendesayi Kufa-Chakezha, Nkengafac Villyen Motaze, Natalie Mayet, Tebogo Mmaborwa Matjokotja, Mzimasi Neti, Tracy Arendse, Teresa Lamola, Itumeleng Matiea, Darren Muganhiri, Babongile Ndlovu, Khuliso Ravhuhali, Emelda Ramutshila, Salaminah Mhlanga, Akhona Mzoneli, Nimesh Naran, Trisha Whitbread, Mpho Moeti, Chidozie Iwu, Eva Mathatha, Fhatuwani Gavhi, Masingita Makamu, Matimba Makhubele, Simbulele Mdleleni, Tsumbedzo Mukange, Trevor Bell, Lincoln Darwin, Fazil McKenna, Ndivhuwo Munava, Muzammil Raza Bano, Themba Ngobeni.
- The NICD Centre for Respiratory Disease and Meningitis are thanked for their assistance in setting up and troubleshooting PCR testing, and ongoing supportive collaboration.
- Staff of SACCESS network laboratories are thanked for their assistance in generating these results.

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Supplementary Table: Data for all wastewater treatment plants tested by SACCESS network

S/ No	Wastewater plant name	Province	Metro or District	Official subdistrict SD or Local municipality	Subdistrict as represented on the graphs	Water service authority	Suburbs in drainage reticulation	Testing laboratory	Date quantitative testing started
		Eastern Cape			No subdistrict	Buffalo City Local Municipality	Wisonia, Dawn, Summer Pride, Amalinda Forest, Haven Hills, Buffalo flats ext, Scenery Park		20-10-2021
2		Eastern Cape			No subdistrict	Buffalo City Local Municipality			
	Gonubie	Eastern Cape			No subdistrict	Buffalo City Local Municipality	Sunrise-on-Sea, Gonubie Manor, Thorn Ridge, Cyprus Dale, Bay View, Donny-brook, Gonubie, Gonubie Park		15-09-2021
		Eastern Cape			No subdistrict	Buffalo City Local Municipality	Mdantsane Newlands		01-06-2021
		Eastern Cape			No subdistrict	Buffalo City Local Municipality	Zone 12 to Zone 18, Unit P, Potsdam, Khayelitsha, WSU Potsdam, Campus, Mbekweni		20-10-2021
		Eastern Cape			No subdistrict	Buffalo City Local Municipality			15-09-2021
		Eastern Cape			No subdistrict	Buffalo City Local Municipality	Duncan Village, Leach Bay, Nahoon		15-09-2021
		Eastern Cape			No subdistrict	Buffalo City Local Municipality			20-10-2021
		Eastern Cape	Nelson Mandela Metropolitan Municipality		No subdistrict	Nelson Mandela Metropolitan Municipality	KwaNobuhle, Uitenhage, Van Riebeekhoogte		
10		Eastern Cape	Nelson Mandela Metropolitan Municipality		No subdistrict	Nelson Mandela Metropolitan Municipality	KwaNobuhle, Uitenhage		

S/ No	Wastewater plant name	Province	Metro or District	Official subdistrict SD or Local municipality	Subdistrict as represented on the graphs	Water service authority	Suburbs in drainage reticulation	Testing laboratory	Date quantitative testing started
			Mangaung			Mangaung		Lumegen	01-09-2021
12			Mangaung			Mangaung	Langenhoven Park, Bloemfontein		16-03-2021
13			Mangaung			Mangaung		Lumegen	01-09-2021
14			Mangaung			Mangaung	Fonteintjie, Rooidam		16-03-2021
15	Botshabelo		Mangaung	Botshabelo SD	Botshabelo	Mangaung	Bonolo, Botshabelo, Poklenberg, Dankbaar, Roodekop	Lumegen	04-10-2021
16			Mangaung	Botshabelo SD	Botshabelo	Mangaung	Kagisanong, Fichardtpark, Bochebela, Phahameng, Generaal deWet, Willows, Batho, Rocklands, Universitas	Lumegen	09-09-2021
17			Mangaung			Mangaung	Midway, Bloemspruit, Grasslands	Lumegen	01-09-2021
18	Dewetsdorp		Mangaung		Naledi and Thabanchu	Mangaung	Dewetsdorp, Frankfort, Glengary	Lumegen	01-09-2021
19	Thaba Nchu		Mangaung	Thaba N'chu SD	Naledi and Thabanchu	Mangaung	Thaba Nchu, Mokwena, Selosesha, Abramskraal, Roodekop, Strydom College, Bultfontein Number Three, Ratlau, Serwalo, Bultfontein Number One, Bultfontein Number Two, Motlala, Lusaka	Lumegen	01-09-2021
20			City of Johannesburg Metropolitan Municipality		No subdistrict	City of Johannesburg Metropolitan Municipality	Walkerville, Hartsenbergfontein, Althea, Golfview, Blignautrus		04-10-2021

Northern Wastewater Treatment Works Gauteng Metropolitan Municipality Zigorian Sunshippies Gauteng Sunsh										
Northern Northern Gauteng Gauteng Gauteng Gauteng Metropolitan Municipality Metropolitan Municip	Date quantitative testing started	Testing laboratory			represented	subdistrict SD or Local	Metro or District	Province		
23 Bushkoppies Gauteng Metropolitan Municipality DSD No subdistrict Metropolitan Municipality DSD No subdistrict Metropolitan Municipality Soweto, Rivasdale NICD 24 Olifantsylei Gauteng City of Johannesburg G SD No subdistrict Metropolitan Municipality Display Disp	06-04-2021		Rivonia, Jukskei Park, Douglasdale, Ferndale, Lone Hill, Sandton, North Riding, Fourways,	Johannesburg Metropolitan	No subdistrict					21
24 Olifantsvlei Gauteng Gauteng Metropolitan Municipality GSD No subdistrict Metropolitan Municipality GSD No subdistrict Metropolitan Municipality Metropolitan Municipality Metropolitan Municipality GSD No subdistrict Metropolitan Municipality Metropolitan Municipality GSD No subdistrict Metropolitan Metropolitan Municipality GSD No subdistrict Metropolitan Metropolit	24-05-2021		Soweto, Rivasdale	Johannesburg Metropolitan	No subdistrict				Goudkoppies	22
24 Olifantsvlei Gauteng Johannesburg Metropolitan Municipality 25 Driefontein Gauteng Gauteng Gauteng Gauteng Gauteng Gauteng Gauteng Metropolitan Municipality 26 Bronkhortspruit Gauteng Gauteng Gauteng Gauteng Gauteng Gauteng Gauteng Gauteng Metropolitan Municipality 27 Klipgat Gauteng Gauteng City of Tshwane Metropolitan Municipality 28 Sandspruit Gauteng Gauteng Gauteng Gauteng Gauteng City of Tshwane Metropolitan Municipality 28 Sandspruit Gauteng Gauteng Gauteng Gauteng Gauteng Gauteng City of Tshwane Metropolitan Municipality 28 Sandspruit Gauteng Gauteng Gauteng City of Tshwane Metropolitan Municipality 29 Sandspruit Gauteng City of Tshwane Metropolitan Municipality 20 Sandspruit Gauteng City of Tshwane Metropolitan Municipality 20 Sandspruit Gauteng Metropolitan Metropolitan Metropolitan Municipality 30 Sandspruit Gauteng Metropolitan	11-10-2021		Johannesburg South, Dube, Willowdene,	Johannesburg Metropolitan	No subdistrict				Bushkoppies	23
25 Driefontein Gauteng Gauteng Gauteng Metropolitan Municipality Mogale City LM No subdistrict Metropolitan Municipality Mogale City LM No subdistrict Metropolitan Municipality Manor, Edenburg, Lone Hill, Rivonia, Sandton, Northdene, Fourways, Paulshof City of Tshwane Metropolitan Municipality Thembisile Hani (sub-districts 3,4,6,7) Tshwane North (sub-districts 3,4,6,7) Klipgat Gauteng Gauteng City of Tshwane Metropolitan Municipality City of Tshwane North (sub-districts 1,2) Tshwane North (sub-districts 1,2) Tshwane North (sub-districts 1,2) Tshwane North (sub-districts 1,2) Tshwane North Metropolitan Municipality Klipgat, Boekenhoutfontein, Soshanguve, Mabopane, Honeyvale, Boekenhoutfontein, Lebaleng City of Tshwane North (sub-districts 1,2) Tshwane North (sub-districts Metropolitan Municipality Manor, Edenburg, Lone Hill, Rivonia, Sandton, Northdene, Fourways, Paulshof Wilgerivier, Wonderfontein, Graley Crown Douglas, Bronkhorst Klipgat, Boekenhoutfontein, Soshanguve, Mabopane, Honeyvale, Boekenhoutfontein, Lebaleng City of Tshwane North (sub-districts Metropolitan Municipality City of Tshwane Medunsa, Hebron, Rosslyn, Strydfontein, IID	11-10-2021			Johannesburg Metropolitan	No subdistrict		Johannesburg Metropolitan			24
Bronk- hortspruit Gauteng Gaut	04-10-2021		Manor, Edenburg, Lone Hill, Rivonia, Sandton, Northdene, Fourways,	Johannesburg Metropolitan	No subdistrict	Mogale City LM				25
27 Klipgat Gauteng City of Tshwane Metropolitan Municipality Tshwane 1 SD (sub-districts 1,2) Tshwane North (sub-districts 1,2) Metropolitan Municipality Boekenhoutfontein, Soshanguve, Mabopane, Honeyvale, Boekenhoutfontein, Lebaleng SAMRC-TE Metropolitan Municipality Boekenhoutfontein, Lebaleng City of Tshwane Tshwane 1 SD (sub-districts Metropolitan Metropolitan Metropolitan Metropolitan Metropolitan Metropolitan North City of Tshwane Medunsa, Hebron, Rosslyn, Strydfontein, IID (sub-districts Metropolitan Metropolitan North North North Metropolitan Metropolitan North	04-10-2021		Wonderfontein, Graley Crown Douglas,	Johannesburg Metropolitan						26
28 Sandspruit Gauteng Metropolitan Tshwane 1 SD (sub-districts Metropolitan Rosslyn, Strydfontein, ID	02-11-2021	SAMRC-TB Platform	Boekenhoutfontein, Soshanguve, Mabopane, Honeyvale, Boekenhoutfontein,	Metropolitan					Klipgat	27
	11-09-2021		Rosslyn, Strydfontein,	Metropolitan					Sandspruit	28
Rooiwal East- ern Works City of Tshwane Tshwane North City of Tshwane Atteridgeville, Pretoria CBD, Pretoria North, NICD Municipality 1,2) Municipality Rosslyn	23-03-2021			Metropolitan			Metropolitan			29
City of Tshwane Tshwane North City of Tshwane Majanen, Hammanskraal, Waterlab/ 30 Temba Gauteng Metropolitan Tshwane 2 SD (sub-districts Metropolitan Mabopane, Soshanguve, Municipality 1,2) Municipality Pyramid, Doornpoort	26-09-2021		Mabopane, Soshanguve,	Metropolitan						30

s/ No	Wastewater plant name	Province	Metro or District	Official subdistrict SD or Local municipality	Subdistrict as represented on the graphs	Water service authority	Suburbs in drainage reticulation	Testing laboratory	Date quantitative testing started
31			City of Tshwane Metropolitan Municipality			City of Tshwane Metropolitan Municipality	Groenkloof, Arcadia, Pretoria South, Gezina, Hercules, Rietfontein, Pretoria Central, Sunnyside, Pretoria East, Prinshof, Daspoort, Villieria, Capital Park, Pretoria West, Wonderboom South, Pretoria-Wes, Innesdale		02-03-2021
32	Sunderland Ridge		City of Tshwane Metropolitan Municipality			City of Tshwane Metropolitan Municipality	Centurion, Olivenhoutbosch and some parts of Midrand.		18-08-2021
33	Babelegi		City of Tshwane Metropolitan Municipality			City of Tshwane Metropolitan Municipality			18-08-2021
34			City of Tshwane Metropolitan Municipality			City of Tshwane Metropolitan Municipality	Elandsfontein, Cullinan, Sonderwater		18-08-2021
			City of Tshwane Metropolitan Municipality			City of Tshwane Metropolitan Municipality			05-10-2021
36	Zeekoegat		City of Tshwane Metropolitan Municipality			City of Tshwane Metropolitan Municipality	Zeekoegat, Magalies Water, Buffelsdrif		04-10-2021
37	Godrich		City of Tshwane Metropolitan Municipality			City of Tshwane Metropolitan Municipality	Bronkspruit town Rhema Park Caltura park Venster Park Zithobeni	SAMRC-TB Platform	13-09-2021
38	Summer Place Pack- age Plant		City of Tshwane Metropolitan Municipality			City of Tshwane Metropolitan Municipality		SAMRC-TB Platform	01-09-2021
	Rietgat		City of Tshwane Metropolitan Municipality			City of Tshwane Metropolitan Municipality			05-10-2021
40	Thaba Tsh- wane		City of Tshwane Metropolitan Municipality			City of Tshwane Metropolitan Municipality	Thaba Tshwane, Generaal Kemp Heuwel Radio Uitkyk		05-10-2022

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S/ No	Wastewater plant name	Province	Metro or District	Official subdistrict SD or Local municipality	Subdistrict as represented on the graphs	Water service authority	Suburbs in drainage reticulation	Testing laboratory	Date quantitative testing started
41			Ekurhuleni Metropolitan Municipality			Ekurhuleni Metropolitan Municipality	Welgedag, Persida		NICD:02-03- 2021 CSIR: 21-09- 2021
42			Ekurhuleni Metropolitan Municipality			Ekurhuleni Metropolitan Municipality	New Modder, Lakefield, Benoni, Boksburg, Northmead, Atlasville		21-09-2021
43			Ekurhuleni Metropolitan Municipality			Ekurhuleni Metropolitan Municipality	Welgedag, Payneville, Selcourt, Casseldale, Springs		21-09-2021
44	Carl Grun- dlingh		Ekurhuleni Metropolitan Municipality			Ekurhuleni Metropolitan Municipality	Nigel, Bultfontein, Laversburg		21-09-2021
45			Ekurhuleni Metropolitan Municipality			Ekurhuleni Metropolitan Municipality	Dalpark, Brakpan, Dersley, Dalview, Benoni, New Modder, Schapenrust		21-09-2021
46			Ekurhuleni Metropolitan Municipality			Ekurhuleni Metropolitan Municipality	Benoni, Dersley, Dalpark, Brakpan, Dalview, Schapenrust		05-10-2021
47	Welgedacht		Ekurhuleni Metropolitan Municipality			Ekurhuleni Metropolitan Municipality	KwaThema, Brakpan, Dersley, Schapenrust		21-09-2021
48	Hartebees- fontein WasteWater Treatment Works		Ekurhuleni Metropolitan Municipality		Ekurhuleni North (N1, N2)	Ekurhuleni Metropolitan Municipality	Mid-Ennerdale, Althea, Grasmere, Elandsfontein		NICD:02-03- 2021 Waterlab/UP: 05-10-2021
49			Ekurhuleni Metropolitan Municipality		Ekurhuleni North (N1, N2)	Ekurhuleni Metropolitan Municipality			21-09-2021
50			Ekurhuleni Metropolitan Municipality		Ekurhuleni North (N1, N2)	Ekurhuleni Metropolitan Municipality	Pinedene, Clayville, Tembisa, Midstream Estates, Olifantsfontein		CSIR: 21-09- 2021 NICD: 02-03- 2021
51			Ekurhuleni Metropolitan Municipality	Ekurhuleni N2 SD	Ekurhuleni North (N1, N2)	Ekurhuleni Metropolitan Municipality	Northmead, Dalpark, Dalview, Lakefield, Benoni, New Modder		05-10-2021

S/ No	Wastewater plant name	Province	Metro or District	Official subdistrict SD or Local municipality	Subdistrict as represented on the graphs	Water service authority	Suburbs in drainage reticulation	Testing laboratory	Date quantitative testing started
52			Ekurhuleni Metropolitan Municipality	Ekurhuleni N2 SD	Ekurhuleni North (N1, N2)	Ekurhuleni Metropolitan Municipality	Northmead, Atlasville, New Modder, Lakefield, Benoni		21-09-2021
53			Ekurhuleni Metropolitan Municipality			Ekurhuleni Metropolitan Municipality	Katlehong, Natalspruit, Randhart, Alrode		05-10-2021
54			Ekurhuleni Metropolitan Municipality			Ekurhuleni Metropolitan Municipality	Bartlett, Atlasville, Boksburg North, Lakefield, Bonaero Park, Ravenswood, Witfield, Boksburg		21-09-2021
55	Vlakplaats WasteWater Treatment Works		Ekurhuleni Metropolitan Municipality			Ekurhuleni Metropolitan Municipality			NICD: 22-02- 2021 CSIR: 21-09- 2021
56			Ekurhuleni Metropolitan Municipality			Ekurhuleni Metropolitan Municipality	Kliprivier, Henley on Klip, Ophir, Glen Donald, Chrissiefontein, Rothdene, Riversdale, Meyerton Farms		21-09-2021
57						Mogale City Local Municipality	Rietvallei, Bhongwem, Brink's Vlakfontein		12-10-2021
58	Magaliesburg			West Rand (Mogale City LM)		Mogale City Local Municipality	Magaliesburg, Mogale City		12-10-2021
				West Rand (Mogale City LM)		Mogale City Local Municipality	Lewisham, Krugersdorp North		12-10-2021
60	Hammarsdale		eThekwini Metropolitan Municipality			eThekwini Metropolitan Municipality	Hammarsdale, Elangeni, Mpumalanga		02-09-2021
61			eThekwini Metropolitan Municipality			eThekwini Metropolitan Municipality	New Germany, Pinetown, Clermont, Pinelands, KwaDabeka		02-09-2021

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62	Isipingo		eThekwini Metropolitan Municipality			eThekwini Metropolitan Municipality	Umbogintwini, Malukaze, Egolokodo, KwaMakhutha, Umlazi		10-08-2021
63			eThekwini Metropolitan Municipality			eThekwini Metropolitan Municipality	Brighton Beach, Grosvenor, King's Rest, Ocean View, Fynnland and Treasure Beach		NICD: 22-02- 2021 DUT: 10-08- 2021 for DUT
64			eThekwini Metropolitan Municipality			eThekwini Metropolitan Municipality	La Lucia, Umhlanga, Prestondale, Phoenix, Duff's Road, Glen Ashley, Mount Edgecombe		10-08-2021
65			eThekwini Metropolitan Municipality			eThekwini Metropolitan Municipality			22-02-2021
66			eThekwini Metropolitan Municipality			eThekwini Metropolitan Municipality	Rietrivier, KwaMashu, Duff's Road, Mount Edgecombe, Phoenix, Richmond, Inanda		10-08-2021
67			eThekwini Metropolitan Municipality			iLembe District municipality			11-10-2021
68			eThekwini Metropolitan Municipality			eThekwini Metropolitan Municipality	Acorn, Albany, Alexander Park		28-10-2021
			u Mgungundlovu District municipality	Msunduzi LM	No subdistrict	The Msunduzi Local Municipality	Pelham, Hayfields, New England, Northdale, Hay Paddock, Scottsville, Cleland, Bishopstowe, Sobantu		02-09-2021
70			uMgungundlovu District municipality	Msunduzi LM	No subdistrict	The Msunduzi Local Municipality	Thornville, Hayfields, Hay Paddock, Lynnfield Park, Cleland		02-09-2021
71			Umgungundlovu District municipality		No subdistrict	Mpofana Local Municipality			28-09-2021

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72			Umgungundlovu District municipality	uMngeni LM	No subdistrict	UMgungundlovu District municipality			02-09-2021
73		Limpopo	Capricorn District municipality		No subdistrict	Polokwane Local Municipality	Westenburg, Nirvana, Bendor, Welgelegen, Moregloed, Annadale, Ivydale, Flora Park, Fauna Park, Penina Park, Ivy Park, Ster Park, Dalmada, Broadlands, Woodlands, and Thornhill	Lumegen: /Waterlab/ UP	Lumegen: 02-08-2021 Waterlab/UP: 11-10-2021
74	Lebowakgo- mo	Limpopo	Capricorn District municipality	Lepelle-Nkumpi LM	No subdistrict	Lepelle-Nkumpi LM	Thabamoopo, Vaalboschlaagte, Lekhuswaneng, Moepeng, Makurung, Sekurwaneng, Ga- Matshele, Makurun		12-10-2021
75	Emalahleni (Riverview)	Mpuma- langa	Nkangala District Municipality		No subdistrict		Lynnville, Duvhapark, Paxton, Klipfontein	Lumegen	26-07-2021
76		Mpuma- langa		City of Mbombela LM	No subdistrict	Mbombela/ Umjindi			30-03-2021
77	Mbombela (Kingstonvale)	Mpuma- langa		Mbombela/ Umjindi	No subdistrict	Mbombela/ Umjindi	Gutshwa, eMpumalanga, eNyalungu, Dwaleni, Hlauhlau, Phasha, Ngodini	Lumegen	26-07-2021
78		Northern Cape			No subdistrict	Hantam Local Municipality			
79		Northern Cape		Sol Plaatjie Local Municipality	No subdistrict	Sol Plaatjie Local Municipality		Lumegen	28-08-2021
80					No subdistrict	JB Marks Local Municipality	Harpington, Vyfhoek, Mooibank, Wilgeboom	Lumegen	17-08-2021

S/ No	Wastewater plant name	Province	Metro or District	Official subdistrict SD or Local municipality	Subdistrict as represented on the graphs	Water service authority	Suburbs in drainage reticulation	Testing laboratory	Date quantitative testing started
81	Klerksdorp main				No subdistrict	Matlosana Local Municipality	Boetrand, Wilkoppies	Lumegen	02-08-2021
82	Rustenburg		Rustenburg Local Municipality	Rustenburg Local Municipality	No subdistrict	Rustenburg Local Municipality		Lumegen and Waterlab/ UP	Lumgen: 26- 07-2021 Waterlab: 11- 10-2021
83		Western Cape	City of Cape Town Metropolitan Municipality		No subdistrict	City of Cape Town Metropolitan Municipality		MRC-BRIP	
84	Cape Flats	Western Cape	City of Cape Town Metropolitan Municipality		No subdistrict	City of Cape Town Metropolitan Municipality			06-10-2022
85		Western Cape	City of Cape Town Metropolitan Municipality	CT Tygerberg	No subdistrict	City of Cape Town Metropolitan Municipality	Crawford, Gleemore, Rondenbosch East		06-10-2021
86	Borcherd's Quarry	Western Cape	City of Cape Town Metropolitan Municipality	CT Tygerberg SD	No subdistrict	City of Cape Town Metropolitan Municipality			09-03-2021
87		Western Cape	City of Cape Town Metropolitan Municipality		No subdistrict	City of Cape Town Metropolitan Municipality			09-03-2021