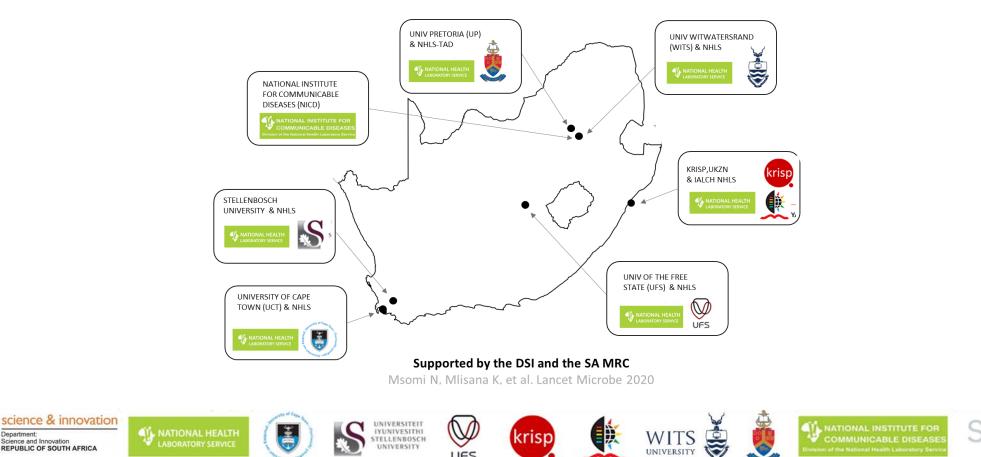


SARS-CoV-2 Sequencing Update 27 January 2023



Prepared by the National Institute for Communicable Diseases (NICD) of the National Health Laboratory (NHLS) on behalf of the Network for Genomics Surveillance in South Africa (NGS-SA)

Department

Science and Innovation

REPUBLIC OF SOUTH AFRICA

The genomic data presented here are based on South African SARS-CoV-2 sequence data downloaded from GISAID (www.gisaid.org) on 27 January 2023 at 14h08

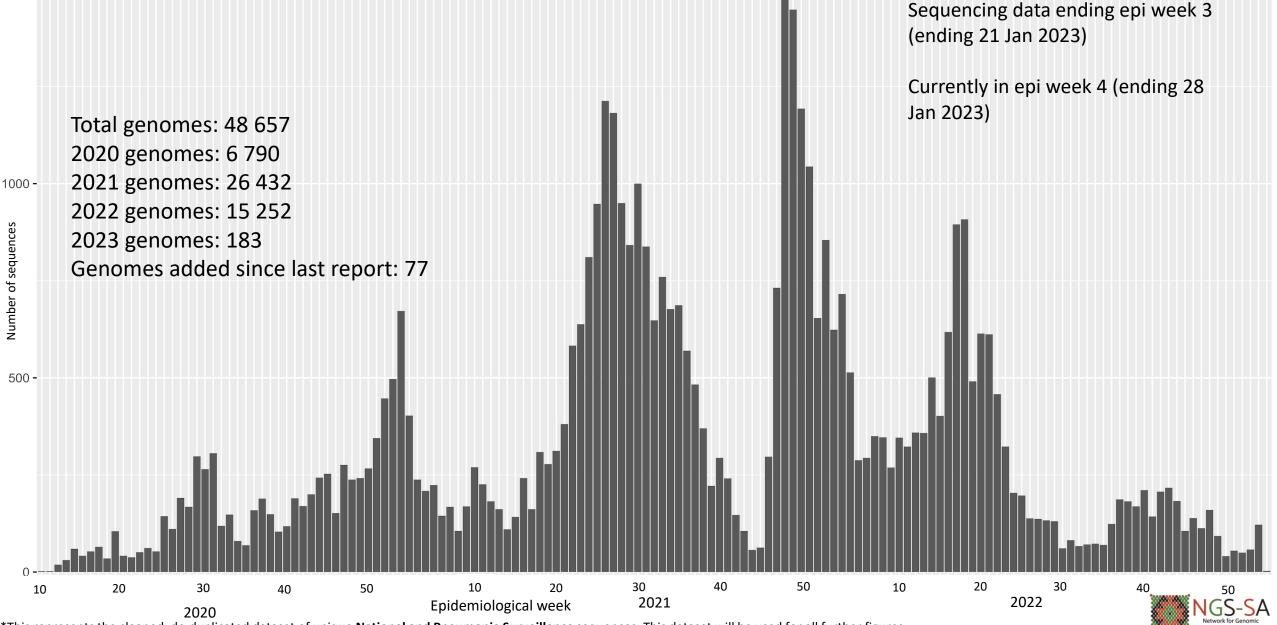


Data license: https://www.gisaid.org/registration/terms-of-use/

Elbe, S., and Buckland-Merrett, G. (2017) Data, disease and diplomacy: GISAID's innovative contribution to global health. Global Challenges, 1:33-46. DOI: 10.1002/gch2.1018 PMCID: 31565258

Shu, Y., McCauley, J. (2017) GISAID: Global initiative on sharing all influenza data – from vision to reality. EuroSurveillance, 22(13) DOI: 10.2807/1560-7917.ES.2017.22.13.30494 PMCID: PMC5388101

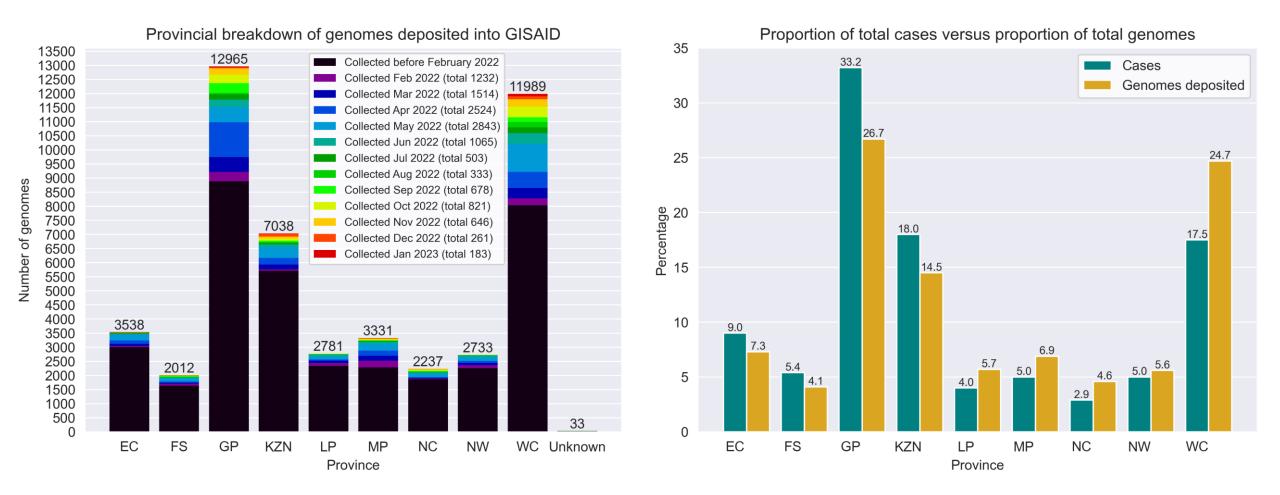
Number of South African genomes deposited on GISAID, by specimen collection week, 2020 – 2023 (N=48 657*)



*This represents the cleaned, de-duplicated dataset of unique National and Pneumonia Surveillance sequences. This dataset will be used for all further figures.

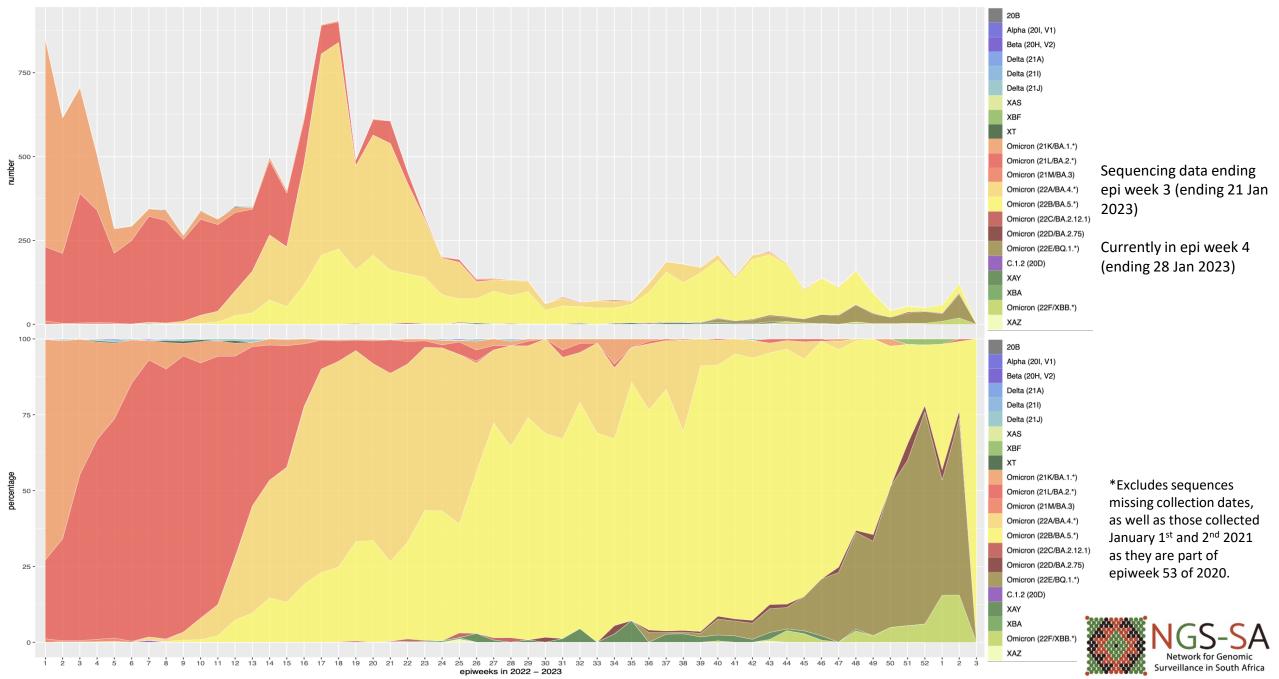
1500 -

GISAID genomes vs total cases, 2020 – 2023 (N=48 657)

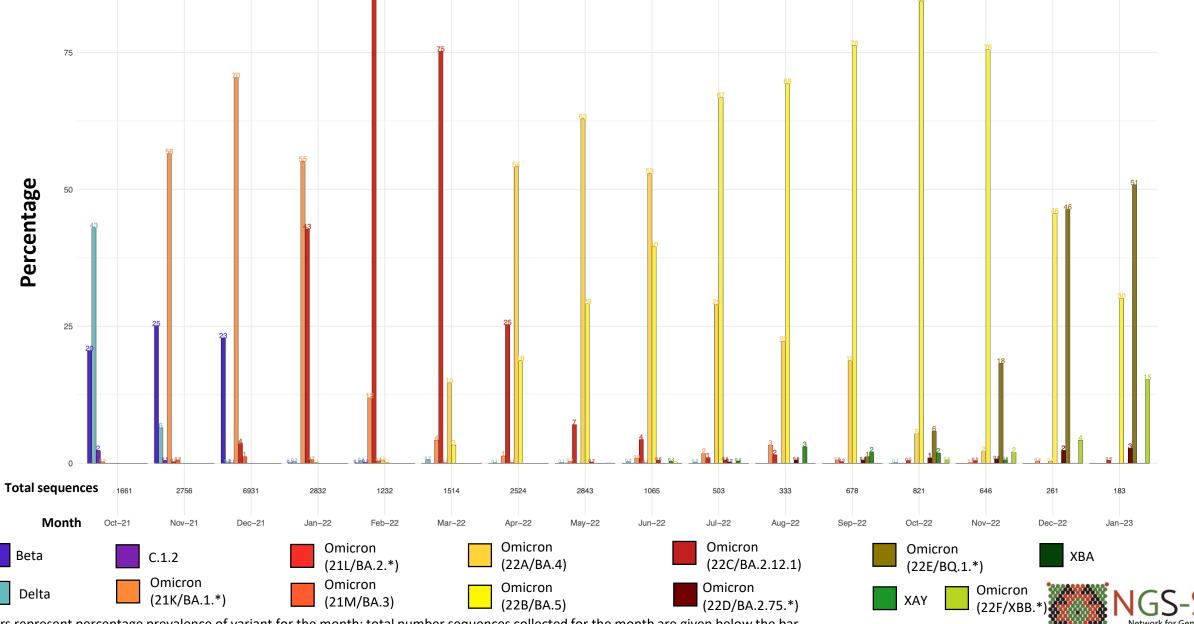




Number and percentage of clades by epiweek in South Africa, 2022-2023 (15 364*)

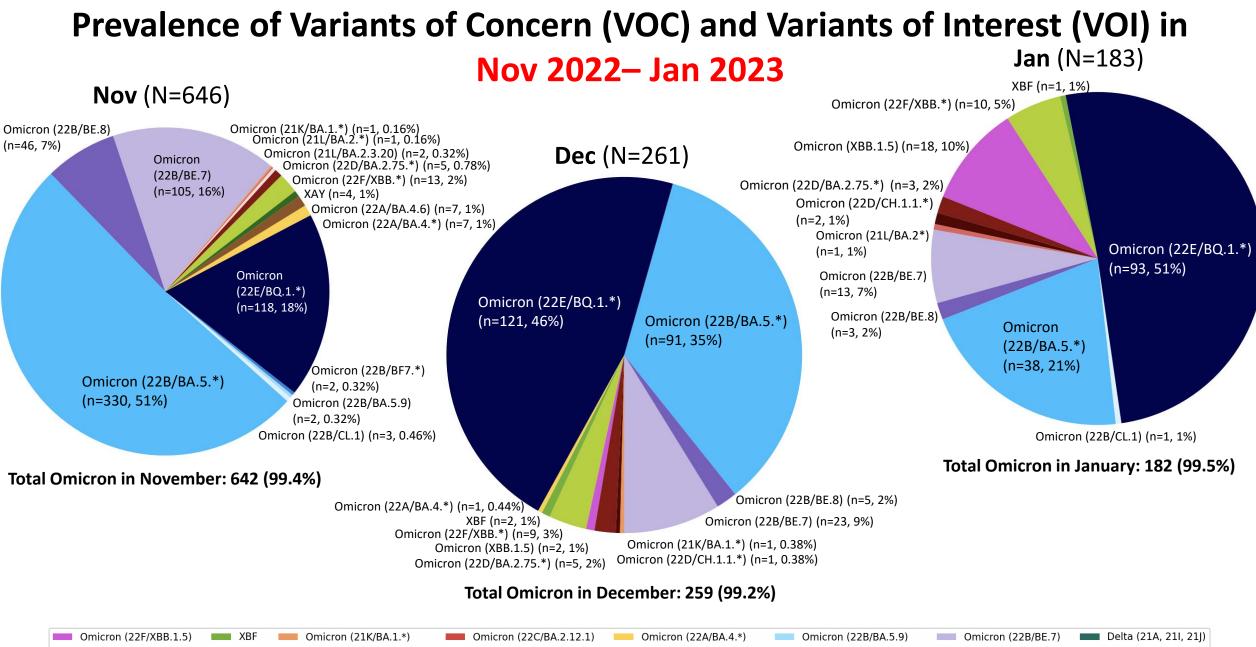


Detection Rates: Beta, Delta, C.1.2, recombinants, and Omicron



Surveillance in South Africa

*Bars represent percentage prevalence of variant for the month; total number sequences collected for the month are given below the bar



Omicron (22F/XBB.*) XAS Omicron (21L/BA.2.*) Omicron (22D/BA.2.75) XAZ Omicron (21L/CM.4) Omicron (22D/CH.1.1.*) XT Omicron (21L/BA.2.3.20) Omicron (21M/BA.3)

XAY

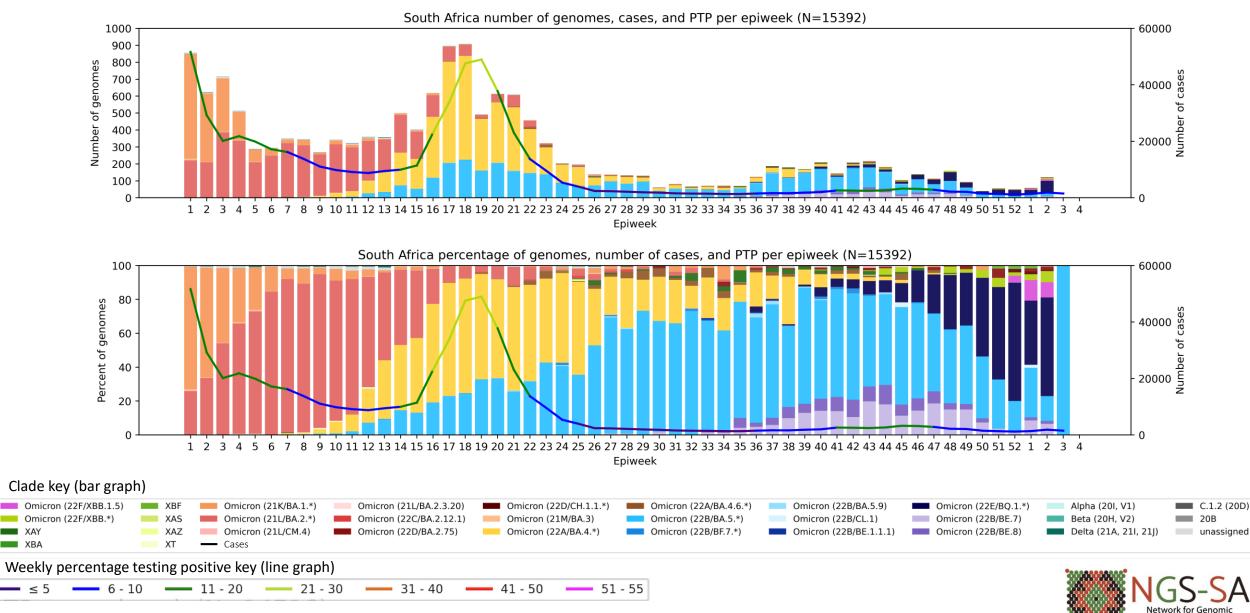
XBA

Omicron (22A/BA.4.6.*) Omicron (22B/BA.5.*) Omicron (22B/BF.7.*)

Omicron (22B/CL.1) Omicron (22B/BE.1.1.1) Omicron (22E/BQ.1.*)

Omicron (22B/BE.8) C.1.2 (20D) 20B Alpha (201, V1) Beta (20H, V2) unassigned

South Africa, 2022-2023, n = 15 392*

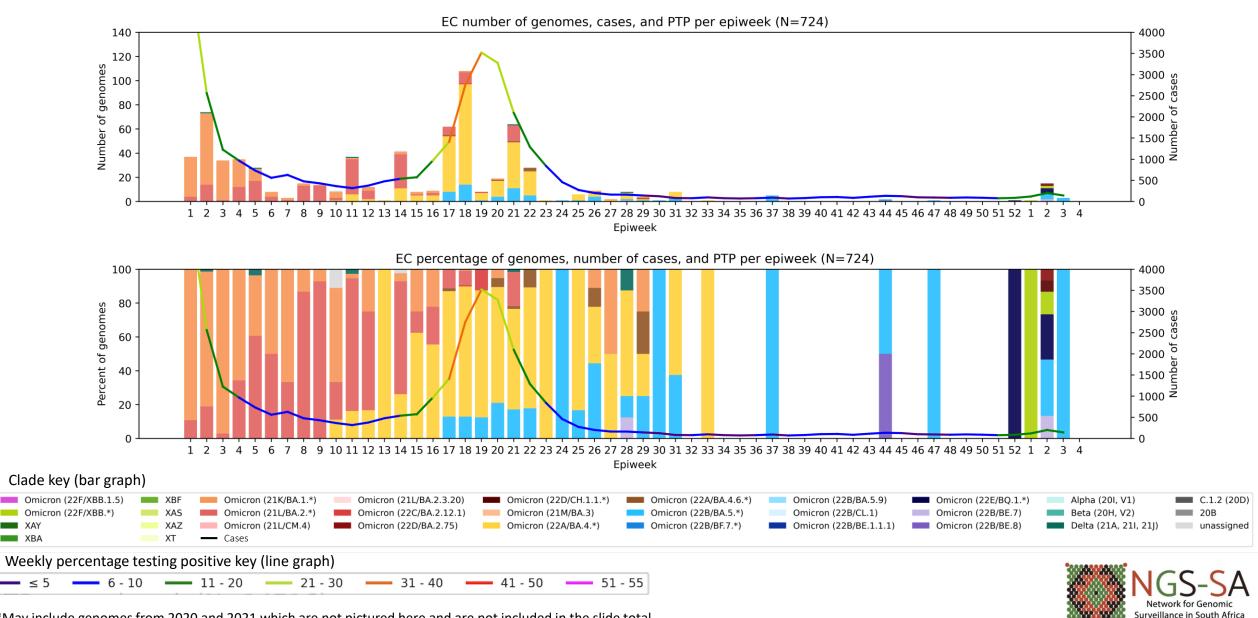


Surveillance in South Africa

*Excludes sequences missing collection dates. Lineages of particular interest (mainly WHO Omicron subvariants under monitoring) are separate from the main clade groupings.

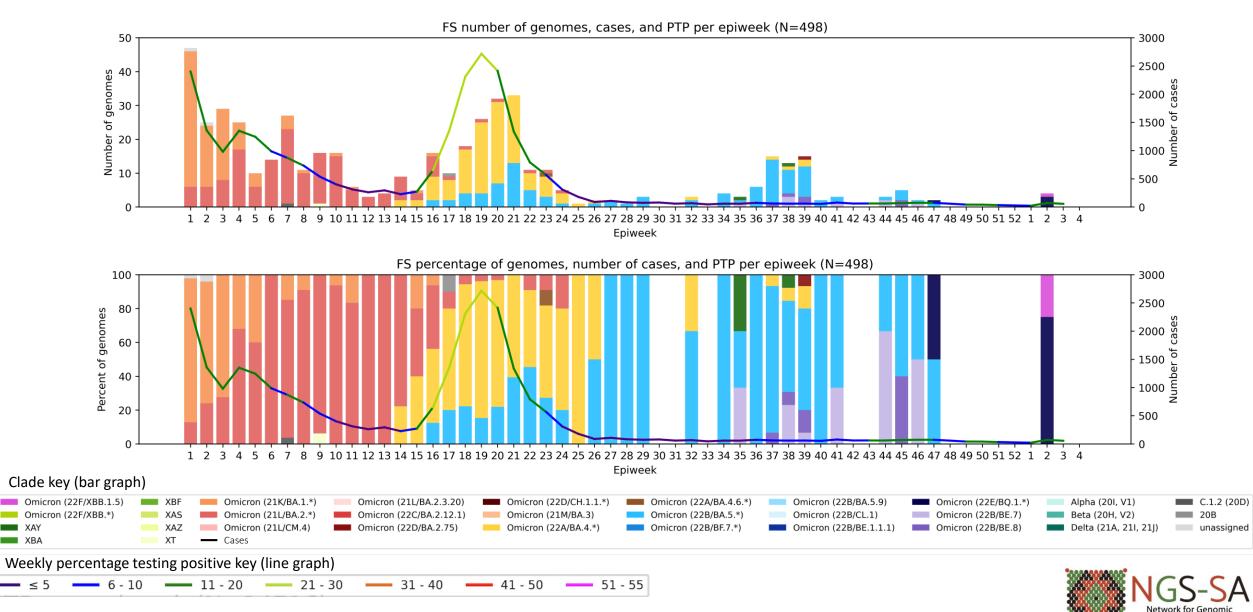
Eastern Cape Province, 2022-2023, n = 724

Genomes added since last report: 14*



Free State Province, 2022-2023, n = 498

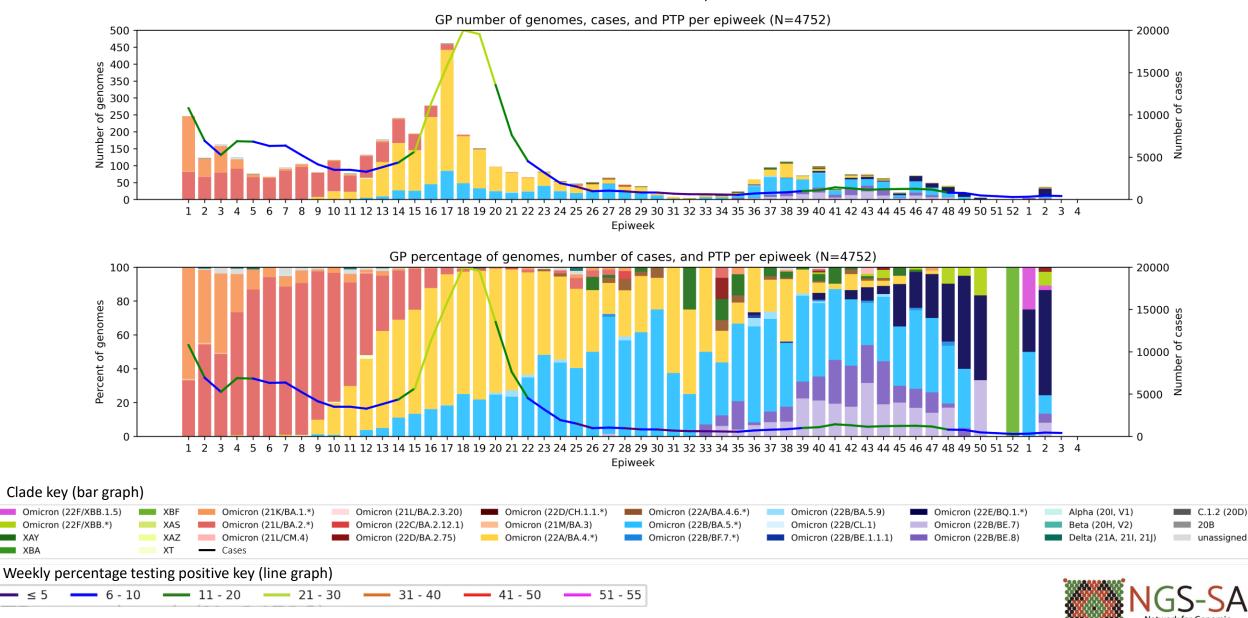
Genomes added since last report: 0*



Surveillance in South Africa

Gauteng Province, 2022-2023, n = 4752

Genomes added since last report: 25*



Surveillance in South Africa

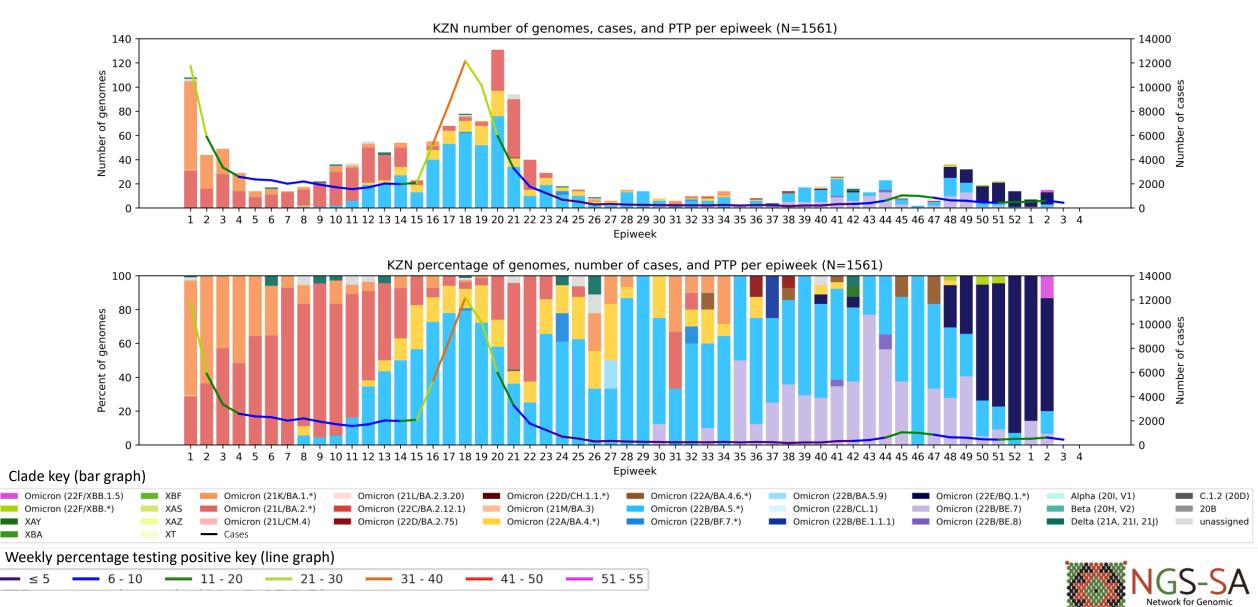
*May include genomes from 2020 and 2021 which are not pictured here and are not included in the slide total.

XAY

XBA

KwaZulu-Natal Province, 2022-2023, n = 1561

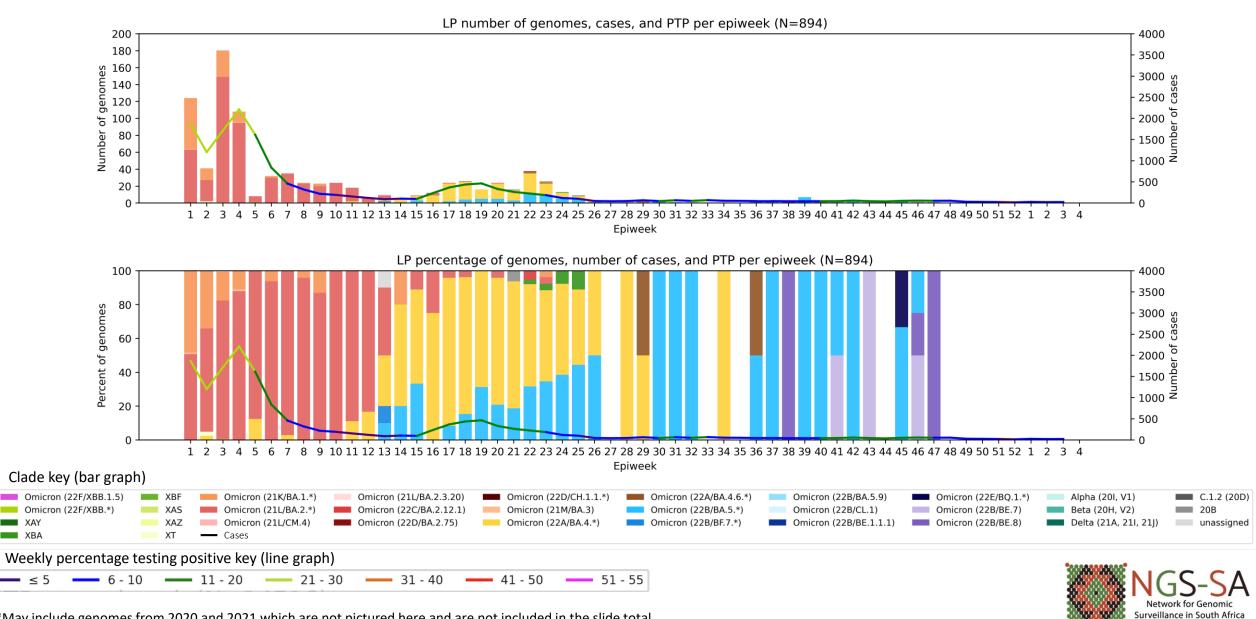
Genomes added since last report: 0*



Surveillance in South Africa

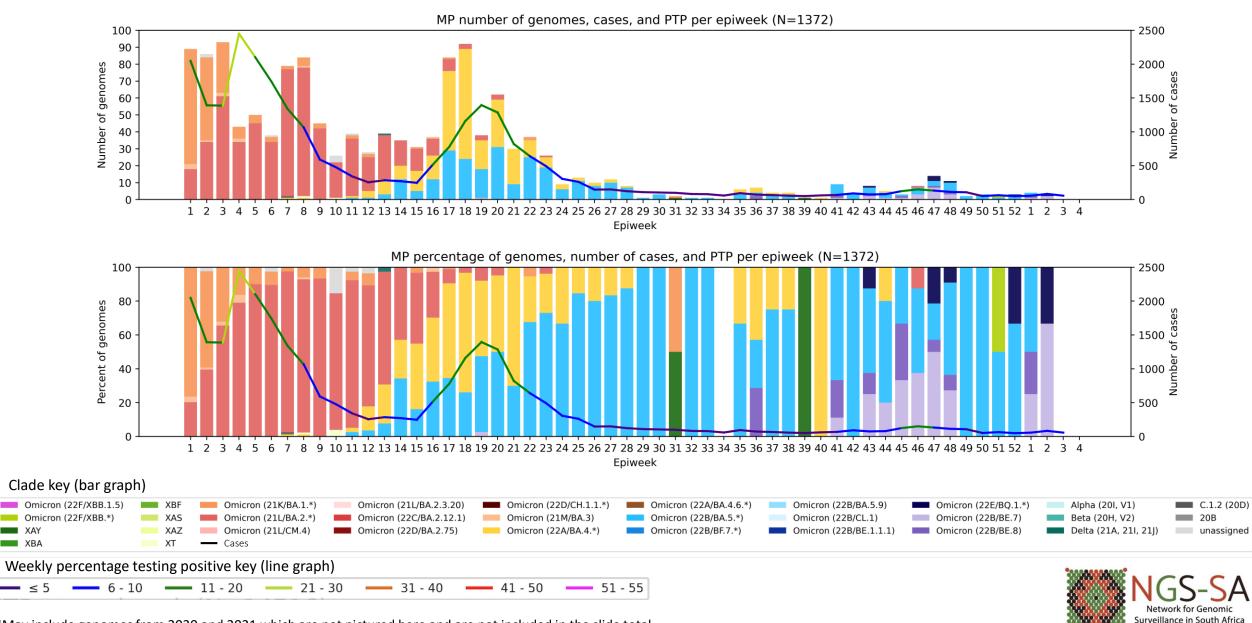
Limpopo Province, 2022-2023, n = 894

Genomes added since last report: 0*



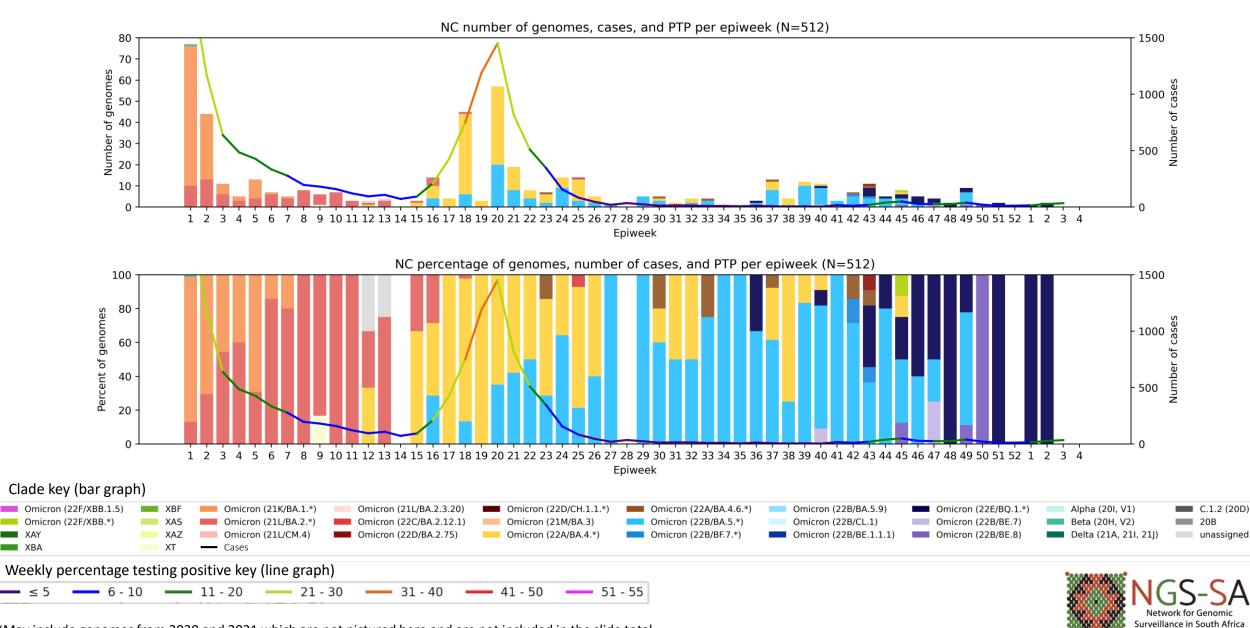
Mpumalanga Province, 2022-2023, n = 1372

Genomes added since last report: 1*



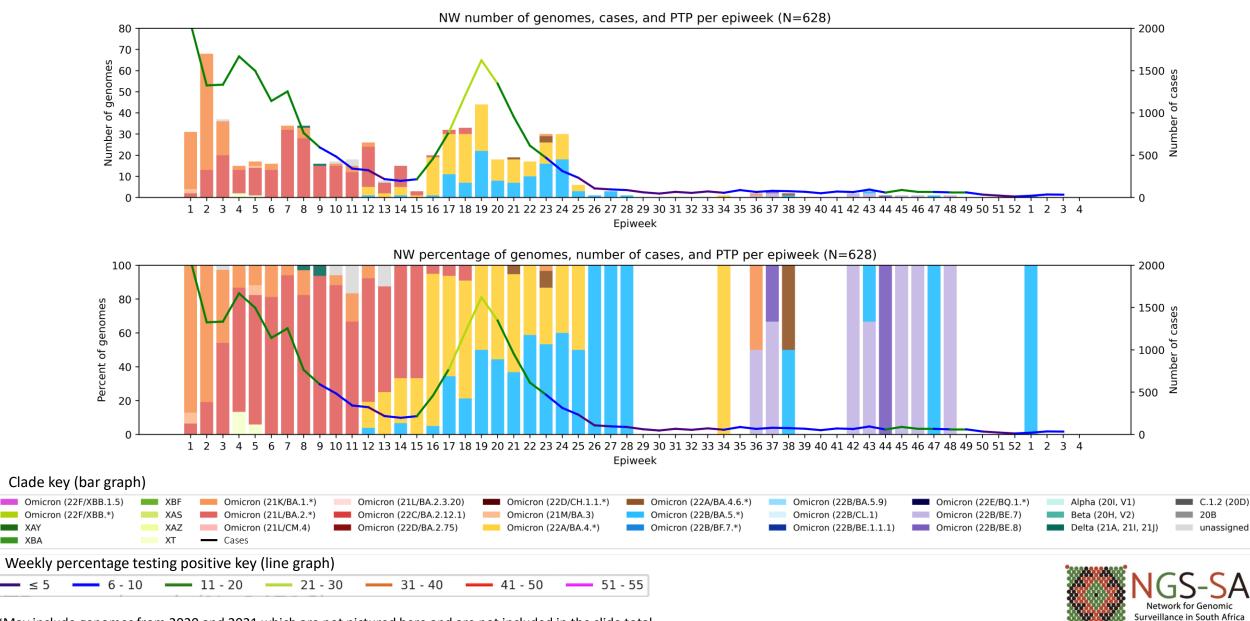
Northern Cape Province, 2022-2023, n = 512

Genomes added since last report: 16*



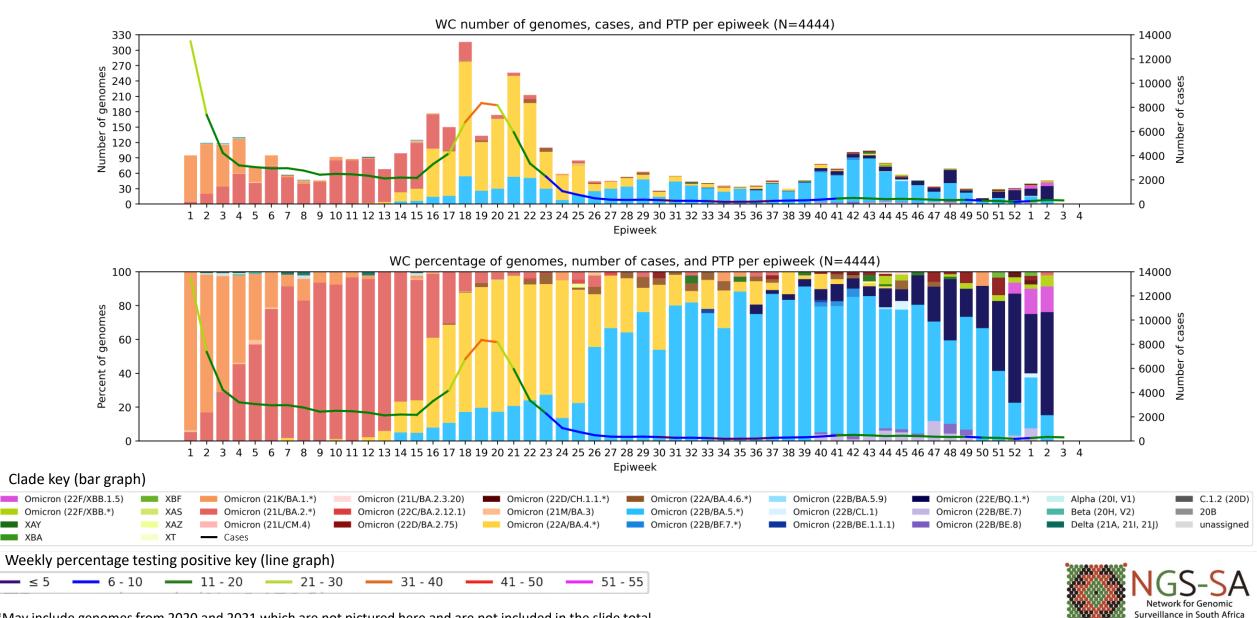
North West Province, 2022-2023, n = 628

Genomes added since last report: 0*



Western Cape Province, 2022-2023, n = 4444

Genomes added since last report: 21*



Summary

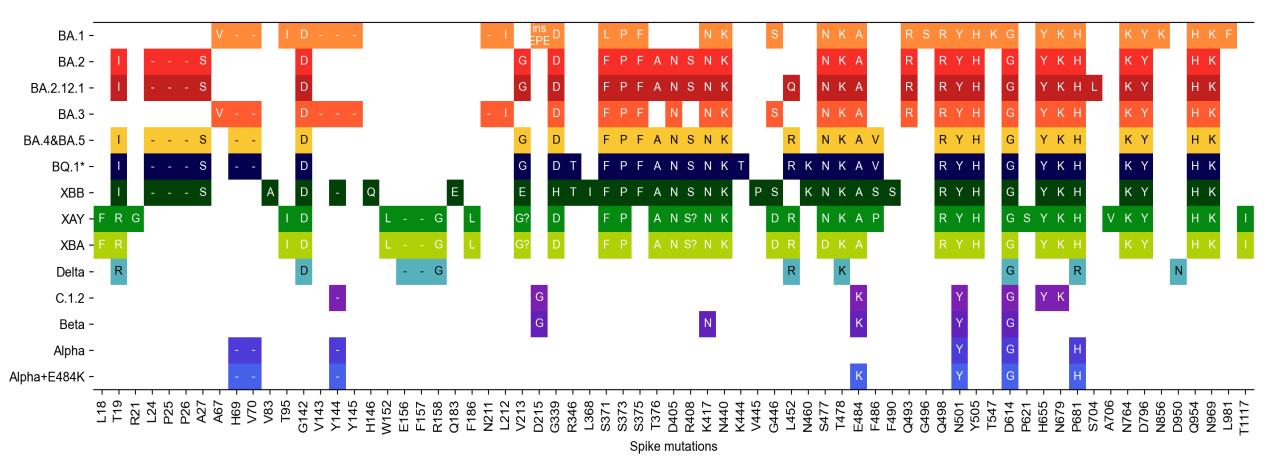
- Sequencing update
 - Eastern Cape, Gauteng, KwaZulu-Natal, Mpumalanga, Northern Cape and Western Cape have sequences for December 2022. All provinces, except for Limpopo, have sequences for January 2023.

• Variant of Concern Omicron in South Africa

- Omicron continued to dominate in November (99%), December (99%) and makes up 99% of January sequences
- Omicron BA.5 and sub-lineages were dominant in November (94%), December (92%) and January (81%)
- BA.2.75.* has been detected in July through January at a low prevalence (<4%)
- XAY has not been detected since November
- BQ.1 and sub-lineages increased in prevalence in October (6%), November (18%) and December (46%), and are currently the dominant Omicron lineage in January (51%)
- XBB.1.5 has been detected in December 2022 (n=2, 1%) and January 2023 (n=18, 10%)
 - (n=1 in the Free State, n=2 in Gauteng, n=2 in KwaZulu-Natal and n=15 in the Western Cape)



Spike protein mutation* profile of Variants of Interest and Concern

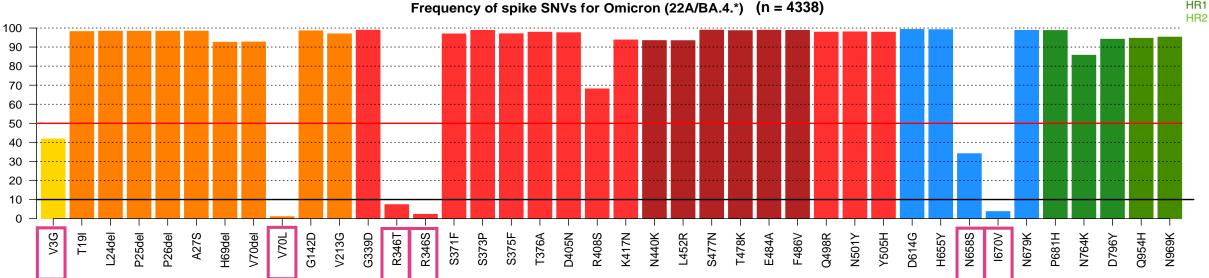


- Multiple changes within the two immunogenic regions in S1 (NTD and RBD)
 - Including a three amino acid insertion
- Accumulation of mutations surrounding the furin cleavage site
 - Including combination of N679K and P681H
- Effect of most spike S2 subunit changes have not been defined, but may be linked to immune escape

*Only mutations present in Omicron, Delta, or recombinant sequences are pictured



BA.4 and BA.5 spike mutations*



Frequency of spike SNVs for Omicron (22B/BA.5.*) (n = 4120)

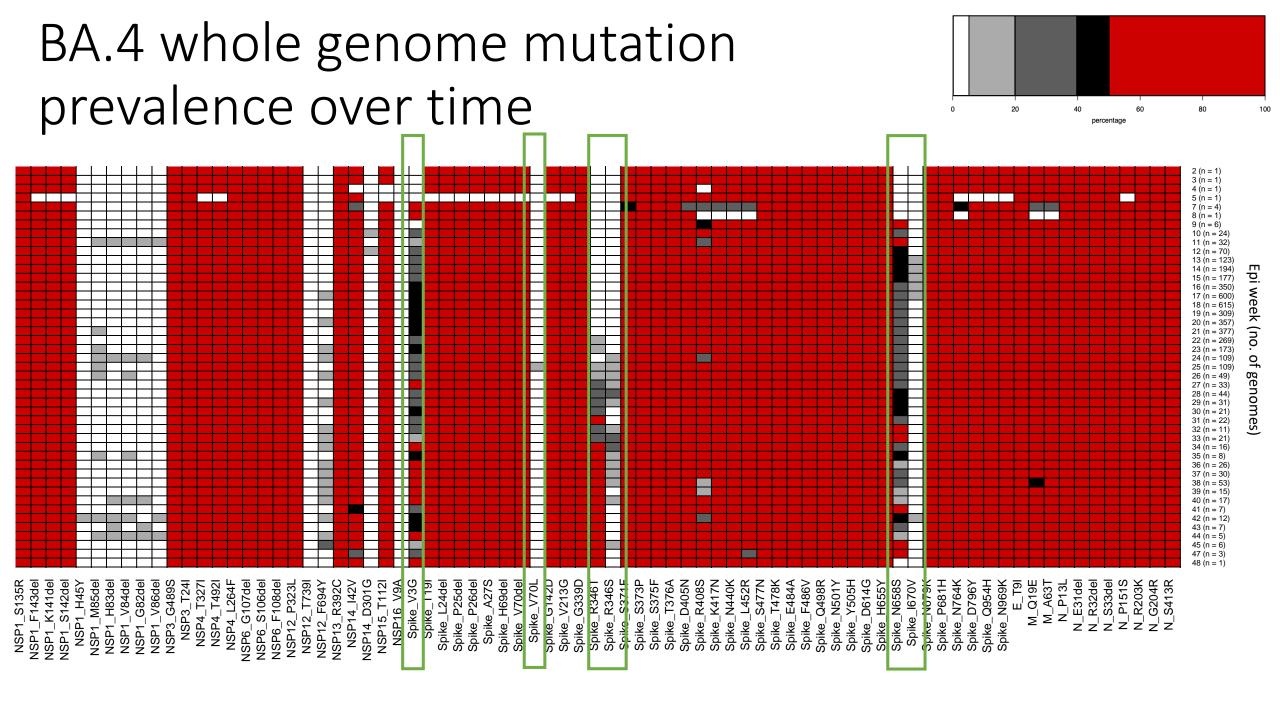


*Only mutations present in \geq 1% of sequences are shown

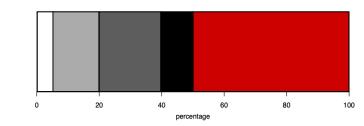
Percentage

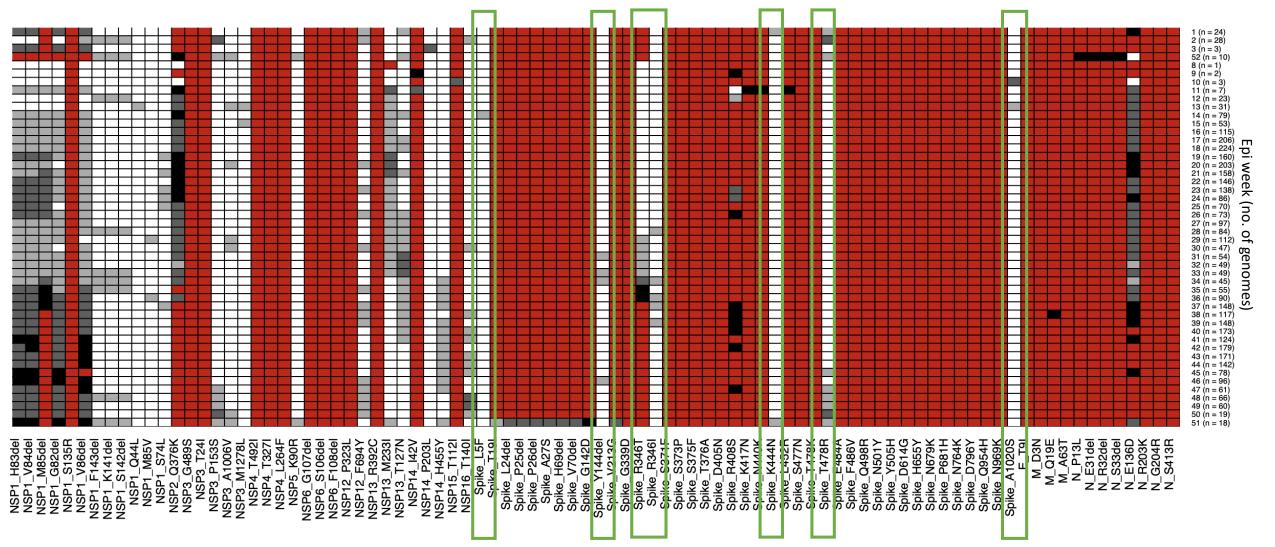
Mutation

NTD RBD RBM S1 S2 HR1

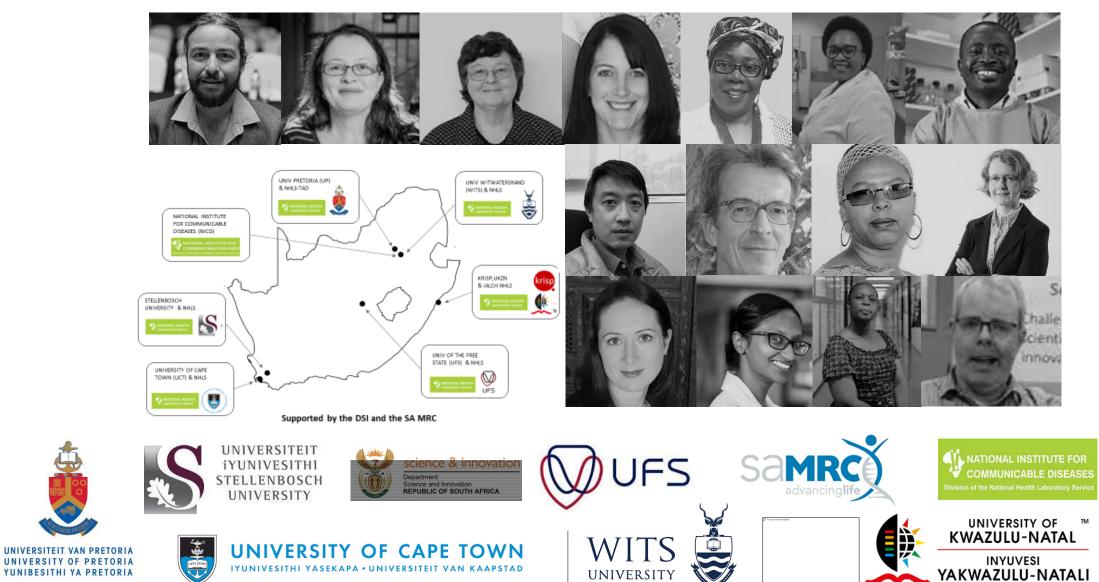


BA.5 whole genome mutation prevalence over time









This project (RIA2020EF-3030) is part of the EDCTP2 programme supported by the European Union"

XX

ЕDСТР

NATIONAL HEALTH LABORATORY SERVICE

University of Stellenbosch & NHLS Tygerberg Virology



Susan Engelbrecht Wolfgang Preiser Gert van Zyl Tongai Maponga **Bronwyn Kleinhans** Shannon Wilson Karabo Phadu Tania Stander Kamela Mahlakwane Mathilda Claassen **Diagnostic laboratory staff**

101046041

AA

EDCTP

W

UKZN-Inkosi Albert Luthuli Central Hospital



Dr Khanvi Msomi Dr Neli Ngcaba Dr Kerusha Govender Dr Tshepiso Mosito Dr Pravi Moodley Mr Malcolm Ellapen Dr Aabida Khan Mr Kubendran Reddy Dr Lili Gounder The COVID-19 Bench team Dr Kerri Francois Dr Cherise Naicker Dr Joedene Chetty

University of KwaZulu-Natal & Africa **Health Research Institute**



Tulio de Oliveira Richard Lessels Houriivah Tegally Eduan Wilkinson Jennifer Giandhari Sureshnee Pillav **Emmanuel James San**

KRISP at UKZN:



National Institute for Communicable Diseases

Cathrine Scheepers

Thandeka Movo

Frances Ayres

Zanele Molaudzi

Bronwen Lambson

Tandile Hermanus

Prudence Kgagudi

Brent Oosthuysen

Mashudu Madzivhandila

Tandile Hermanus



Diseases & Meningitis Anne von Gottberg Thabo Mohale Daniel Amoako Boitshoko Mahlangu Noxolo Ntuli Anele Mnguni Amelia Buys Cardia Fourie Noluthando Duma Linda de Gouveia Nicole Wolter



Sequencing Core Facility Zamantungwa Khumalo Annie Chan **Constantinos Kurt Wibmer** Morne du Plessis Stanford Kwenda Phillip Senzo Mtshali **Mushal Allam** Florah Mnyameni







Sabeehah Vawda Felicity Burt Thokozani Mkhize **Diagnostic laboratory staff**

NHLS Division of Virology

University of the

Free State

Dominique Goedhals

Emmanuel Ogunbayo

Makgotso Maotoana

Lutfiyya Mohamed

Armand Bester

Martin Myaga

Peter Mwangi

Milton Mogotsi

UNIVERSITY OF THE FREE STATE UNIVERSITEIT VAN DIE VRYSTAAT YUNIVESITHI Y FREISTA'

UFS



University of Cape Town, NHLS & Western Cape Government

health

Department. Health REPUBLIC OF SOUTH ATRICA



NHLS-UCT Carolyn Williamson Nei-yuan Hsiao Diana Hardie Kruger Marais





Stephen Korsman

UCT. IDM and CIDRI-Africa

Deelan Doolabh Arash Iranzadeh Lynn Tyers Innocent Mudau Nokuzola Mbhele Fezokuhle Khumalo Thabang Serakge Bruna Galvão Arghavan Alisoltani (U. California)







Darren Martin Nicola Mulder Wendy Burgers Ntobeko Ntusi Rageema Joseph Sean Wasserman Linda Boloko



Samrc CAPE TOWN HVTN

> cience & innovation etrent: nos and knovation VELIC OF SOUTH AFRICA

Zoonotic arbo and respiratory virus program **Centre for Viral Zoonoses Department Medical Virology/ NHLS Tshwane Academic division University of Pretoria**

> ZARV research program/UP Marietjie Venter (Head: ZARV) Adriano Mendes (Postdoc) Amy Strydom (Postdoc) Michaela Davis (MSc, intern medical scientist) Carien van Niekerk

NHLS Tshwane Prof Simnikiwe Mayaphi (HOD)

Funders:

GIZ/BMBF: African Network for Improved diagnostics and epidemiology of common and emerging infectious agents (ANDEMIA) G7 Global Health fund, Robert Koch Institute, Dr Fabian Leendertz

Centre for Respiratory Centre for HIV and STIs Jinal Bhiman

Josie Everatt Jackie Kleynhans Sibongile Walaza

Mignon du Plessis Stefano Tempia Mvuyo Makhasi **Cheryl Cohen**

Penny Moore Lynn Morris

NICD Groups NICD COVID-19 response team NICD SARS-CoV-2 Sequencing Group











Key to Diagnostic Excellent

AMPATH

LABORATORIES

PathCare

1

Vermaak

africa

aboratorie

FIOCRUZ

NHLS Koeleka Mlisana Zinhle Makatini Eugene Elliot Florette K. Treurnicht Kathleen Subramoney Oluwakemi Laguda-Akingba Shareef Abrahams Greta Hoyland Gloria Selabe Elias Bereda

Jeannette Wadula

Hyrax Biosciences Simon Travers

Cape Town HVTN Laboratory Erica Anderson-Nissen Anneta Naidoo

Ndlovu Research Hugo Tempelman CJ Umunnakwe

Lancet Allison J. Glass Raquel Viana

Ampath Terry Marshall Cindy van Deventer Eddie Silberbauer

Pathcare Vermaak Andries Dreyer Howard Newman Riaan Writes Marianne Wolfaardt Warren Lowman

Bridge-the-Gap Raymond Rott

Cytespace Africa Laboratories Christa Viljoen

ARC-OVI Lia Rotherham **CAPRISA** Salim Abdool Karim Nigel Garret

Additional support and collaborators

UKZN - Big Data Francesco Pettruccione Ilya Sinayskiy

University of Oxford José Lourenço

FioCruz, Brazil Vagner Fonseca Marta Giovanetti Luiz Carlos Junior Alcantara Africa CDC John Nkengasong Sofonias Tessema

> Netcare Richard Friedland Craig Murphy Caroline Maslo Liza Sitharam

DSI

Glaudina Loots

SA MRC Glenda Gray

Pathcare N1 City Jean Maritz Nadine Cronje Petra Raimond Kim Hoek











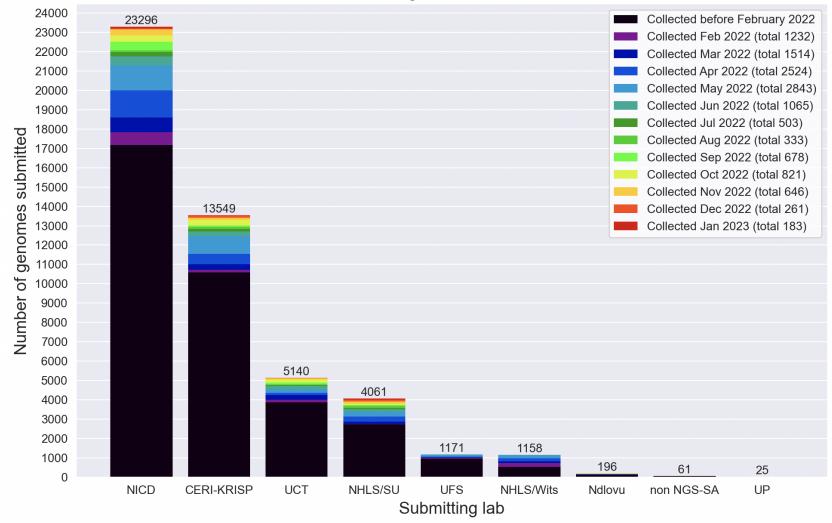






South African genomes submitted per submitting lab, 2020 - 2023 (N=48 657)

Submitting labs in South Africa



NGS-SA Labs

CERI: Centre for Epidemic Response and Innovation **KRISP:** KZN Research Innovation and Sequencing Platform NDLOVU: Ndlovu Research Laboratories **NICD**: National Institute for Communicable Diseases **NHLS**: National Health Laboratory Service **SU**: Stellenbosch University UCT: University of Cape Town **UFS**: University of the Free State **UP**: University of Pretoria

Multiple labs from NGS-SA and collaborating public and private laboratories are contributing to sequencing, both as originating and as submitting (pictured here) laboratories.



Currently circulating Variants of Concern (VOC)

WHO label	Pango lineage∙	GISAID clade	Nextstrain clade	Additional amino acid changes monitored°	Earliest documented samples	Date of designation
Omicron*	B.1.1.529	GR/484A	21K, 21L, 21M, 22A, 22B, 22C, 22D	+S:R346K +S:L452X +S:F486V	Multiple countries, Nov-2021	VUM: 24-Nov-2021 VOC: 26-Nov-2021

* Includes BA.1, BA.2, BA.3, BA.4, BA.5 and descendent lineages. It also includes BA.1/BA.2 circulating recombinant forms such as XE. WHO emphasizes that these descendant lineages should be monitored as distinct lineages by public health authorities and comparative assessments of their virus characteristics should be undertaken.

• Only found in a subset of sequences

https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/ accessed 12 August 2022

Omicron subvariants under monitoring

Pango lineage [#] (+ mutation)	GISAID clade	Nextstrain clade	Relationship to circulating VOC lineages	Spike genetic features	Earliest documented samples
BA.5** (+R346X or +K444X or +V445X or +N450D or +N460X)	GRA	22B	BA.5 sublineages (e.g. BF.7, BF.14, BQ.1)	BA.5 + one or more of these mutations: S:R346X, S:K444X, S:V445X , S:N450D or S:N460X	07-02-2022
BA.2.75***	GRA	22D	BA.2 sublineage	BA.2.75: BA.2 + S:K147E, S:W152R, S:F157L, S:I210V, S:G257S, S:D339H, S:G446S, S:N460K, S:Q493R reversion BA.2.75.2: BA.2.75 + S:R346T, S:F486S, S:D1199N	31-12-2021
BJ.1****	GRA	21L	BA.2 sublineage (B.1.1.529.2.10.1.1)	BA.2+S:V83A, S:Y144-, S:H146Q, S:Q183E, S:V213E, S:G339H, S:R346T, S:L368I, S:V445P, S:G446S, S:V483A, S:F490V, S:G798D, S:S1003I	06-09-2021
BA.4.6	GRA	22A	BA.4 sublineage	BA.4+S:R346T, S:N658S	20-07-2020
ХВВ ^{\$}		recombinant	Recombinant of BA.2.10.1 and BA.2.75 sublineages, i.e. BJ1 and BM.1.1.1, with a breakpoint in S1	BA.2+ S:V83A, S:Y144-, S:H146Q, S:Q183E, S:V213E, S:G252V, S:G339H, S:R346T, S:L368I, S:V445P, S:G446S, S:N460K, S:F486S, S:F490S	13-08-2022
BA.2.3.20 [§]	GRA	21L	BA.2 sublineage	BA.2+ S:M153T, S:N164K, S:H245N, S:G257D, S:K444R, S:N450D, S:L452M, S:N460K, S:E484R	15-08-2022

* these subvariants are tracked under Omicron unless/until sufficient evidence arises that the virus characteristics are substantially different from what is known about the VOC they belong to. If this evidence arises, WHO will decide, in consultation with the TAG-VE, if designation of the emerging variant warrants a separate WHO label.

includes descendent lineages

** additional mutations outside of the spike protein: N:G30-, N:S33F, N:E136D, ORF1a:Q556K, ORF1a:L3829F, ORF1b:Y264H, ORF1b:M1156l, ORF9b:D16G, ORF9b:M26-, ORF9b:A29I, ORF9b:V30L.

*** additional mutation outside the spike protein: ORF1a:S1221L, ORF1a:P1640S, ORF1a:N4060S; ORF1b:G662S; E:T11A

**** additional mutations outside of the spike protein: Mutations: M:D3Y, N:T282I, ORF1a:K47R, ORF1b:G662S, ORF1b:S959P, ORF7a:I110T

^{\$} additional mutations outside of the spike protein: E:T11A, ORF1a:K47R, ORF1b:G662S, ORF1b:S959P, ORF8:G8*

§ additional mutations outside of the spike protein: ORF1a:T727I, ORF1a:I1714T, ORF1a:M2169V, ORF1a:T2174I, ORF1a:T2648I, ORF1a:A2909V, ORF1a:Q3922R, ORF1b:T1404M, ORF3a:L140F, ORF9b:D89E

https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/ accessed 14 October 2022

Previously circulating Variants of Concern

WHO label	Pango lineage●	GISAID clade	Nextstrain clade	Earliest documented samples	Date of designation
Alpha	B.1.1.7	GRY	20I (V1)	United Kingdom, Sep-2020	VOC: 18-Dec-2020 Previous VOC: 09-Mar-2022
Beta	B.1.351	GH/501Y.V2	20H (V2)	South Africa, May-2020	VOC: 18-Dec-2020 Previous VOC: 09-Mar-2022
Gamma	P.1	GR/501Y.V3	20J (V3)	Brazil, Nov-2020	VOC: 11-Jan-2021 Previous VOC: 09-Mar-2022
Delta	B.1.617.2	G/478K.V1	21A, 21I, 21J	India, Oct-2020	VOI: 4-Apr-2021 VOC: 11-May-2021 Previous VOC: 7-Jun-2022

• Includes all descendant lineages. See the cov-lineages.org and the Pango network websites for further details.

https://www.who.int/en/activities/tracking-SARS-CoV-2-variants/ accessed 12 August 2022

Submission of routine specimens for sequencing

- representative of multiple geographic regions (provinces/districts/health facilities) from individuals of
 - all ages
 - over as many time periods during the SARS-CoV-2 epidemic in South Africa
- requested that testing laboratories in both the private and public sectors, submit respiratory samples to their closest NGS-SA sequencing laboratory on a routine basis (ideally every week) as follows, depending on the capacity of the testing laboratory:
 - All positives samples should be sent every week (NGS-SA laboratory will perform random sampling as described below) OR
 - A weekly selection of approximately 10%-20% of randomly selected positive samples should be sent every week. Number of selected samples will depend on the size of laboratory and how many other laboratories are drained by the submitting laboratory.

Submission of special interest specimens for sequencing

In addition to routine samples mentioned above, please send specimens separately to above and clearly marked if:

- Suspected vaccine breakthrough (≥14 days after vaccine), especially if hospitalised and clinically severe
- Suspected re-infection (≥90 days after previous episode), especially if hospitalised and clinically severe
- Prolonged shedding with high SARS-CoV-2 viral loads (i.e. Ct values less than 30 for more than 1 month post-primary diagnosis) in immunocompromised individuals
- Possible animal-to-human transmission
- Suspected cases of importation from another country, especially countries known to harbour SARS-CoV-2 variants of concern or countries with little available information
- Clusters of "unusual" cases (e.g., in terms of disease presentation, patient groups affected, etc.)